

Nano secondary ion mass spectrometry (NanoSIMS) – technique and application to biological samples

Dirk Schaumlöffel

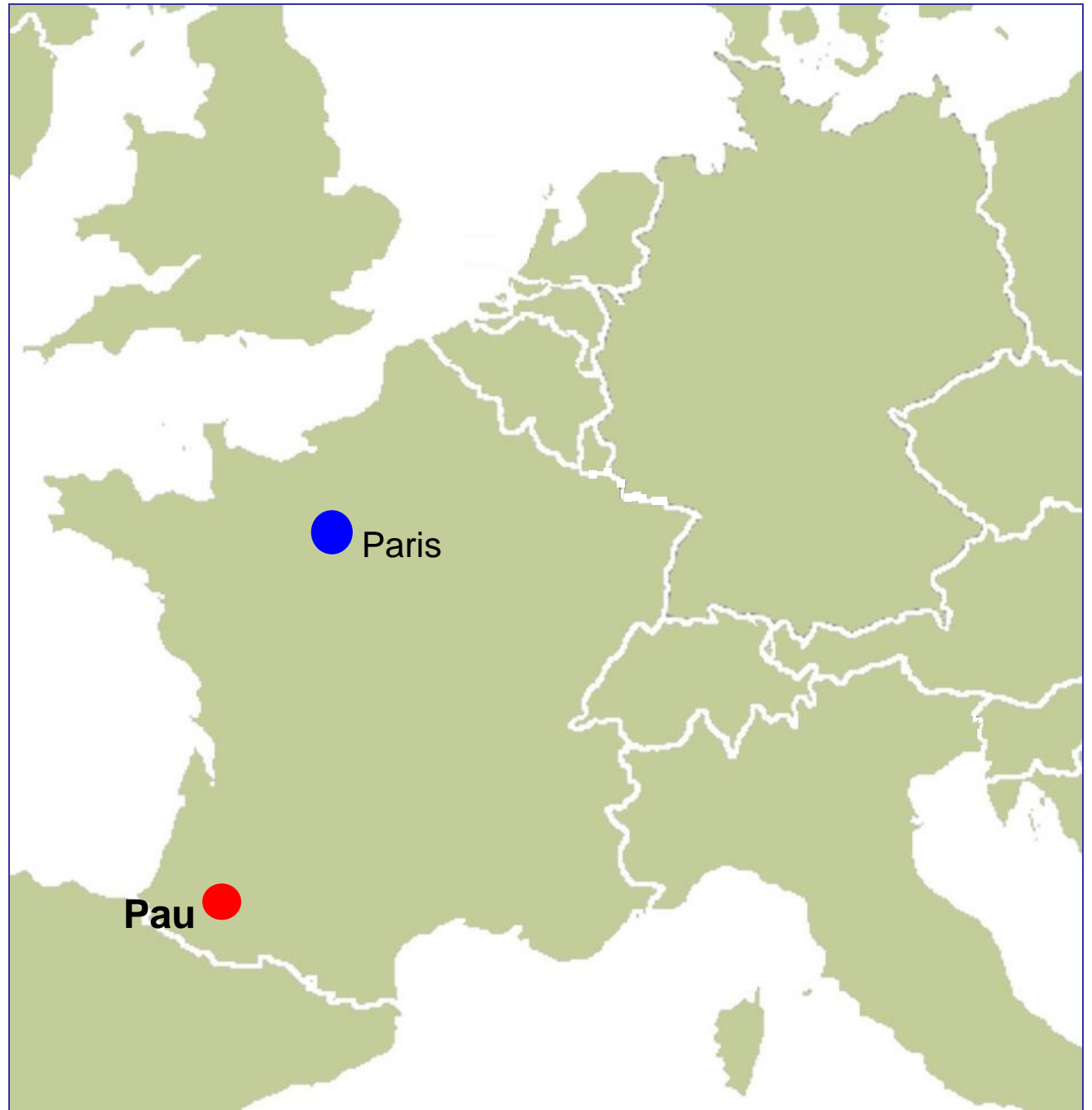
Université de Pau et des Pays de l'Adour / CNRS

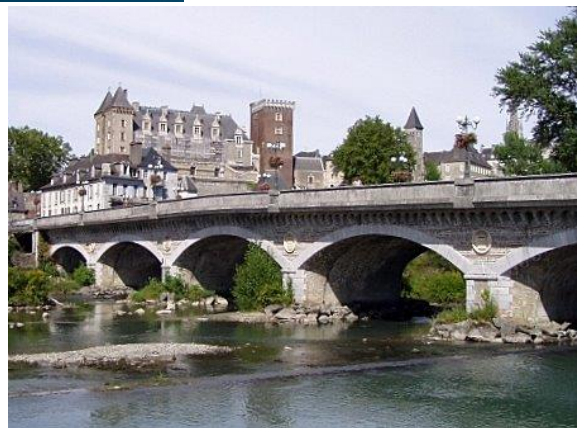
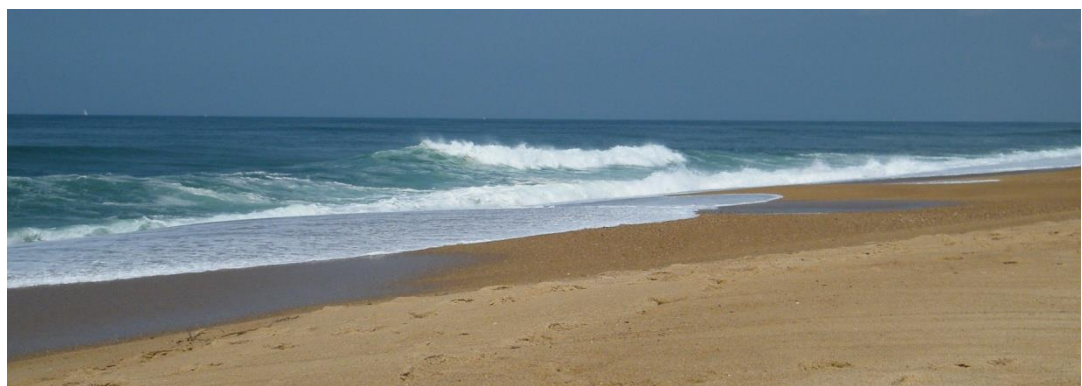
Institut des Sciences Analytiques et de Physico-Chimie

pour l'Environnement et les Matériaux, UMR 5254 IPREM/LCABIE, Pau, France

KICK-OFF MEETING

Accumulation, Subcellular Mapping and Effects of Trace Metals in Aquatic
Organisms (AQUAMAPMET)
Zagreb, 14 September 2015





IPREM - Institut des Sciences Analytiques et de Physico-chimie pour l'Environnement et les Matériaux



Staff

120 permanents

100 PostDocs and
PhD students

4 divisions

Physicochemistry

**Analytical Chemistry:
bio-inorganic and environment**

Polymer chemistry

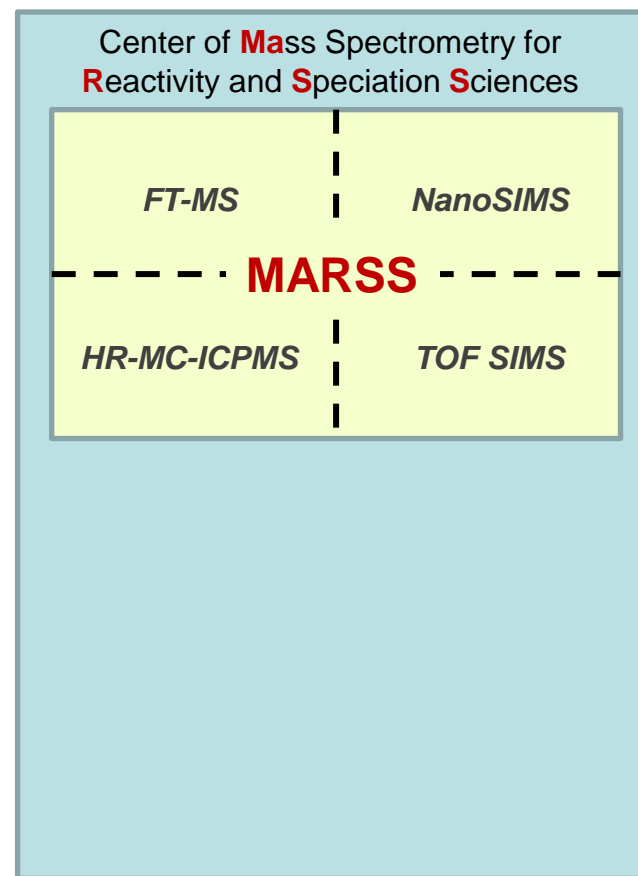
Microbiology and environment



IPREM 1



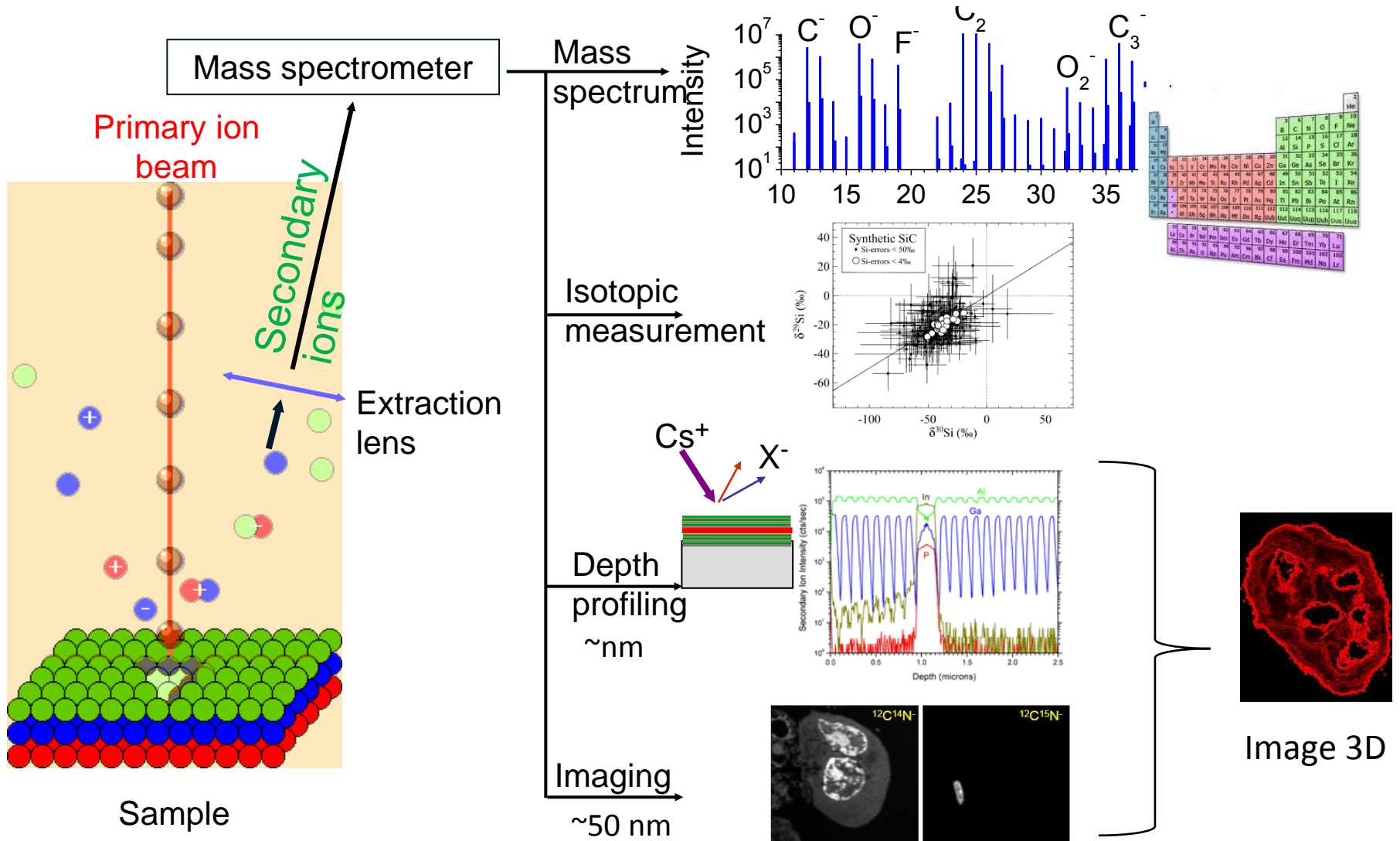
IPREM 2



Outline

- 1 Introduction to SIMS**
- 2 Element imaging by Nano Secondary Ion Mass Spectrometry (NanoSIMS)**
- 3 Application to biological samples: subcellular element imaging**

SIMS : Secondary ion mass spectrometry



Dynamic SIMS technique

- Samples analyzed under **ultra high vacuum** (UHV); biological samples must be dehydrated.

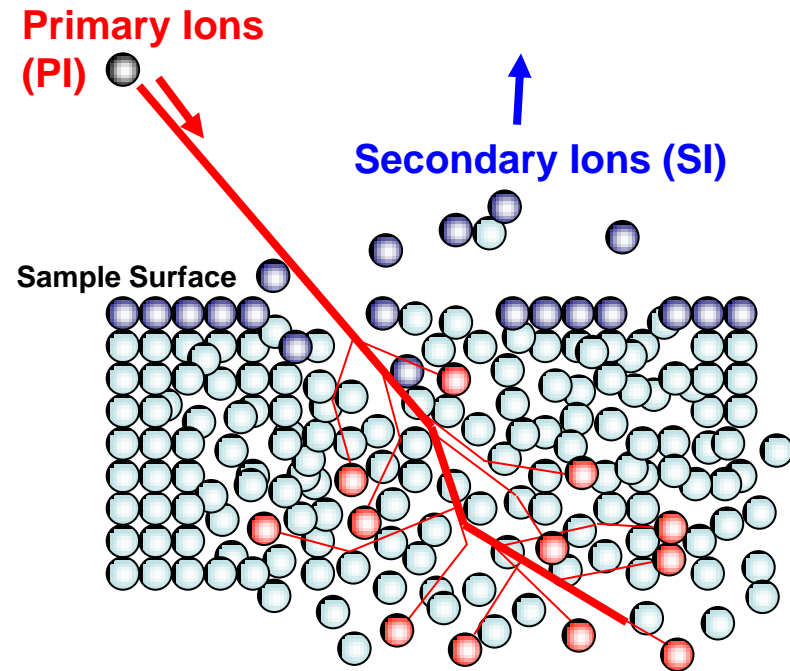
- **Bombardment** by focused **Primary Ions (PI)**:

- **Collision cascade** (10-20nm depth) with simultaneous Implantation and Sputtering.

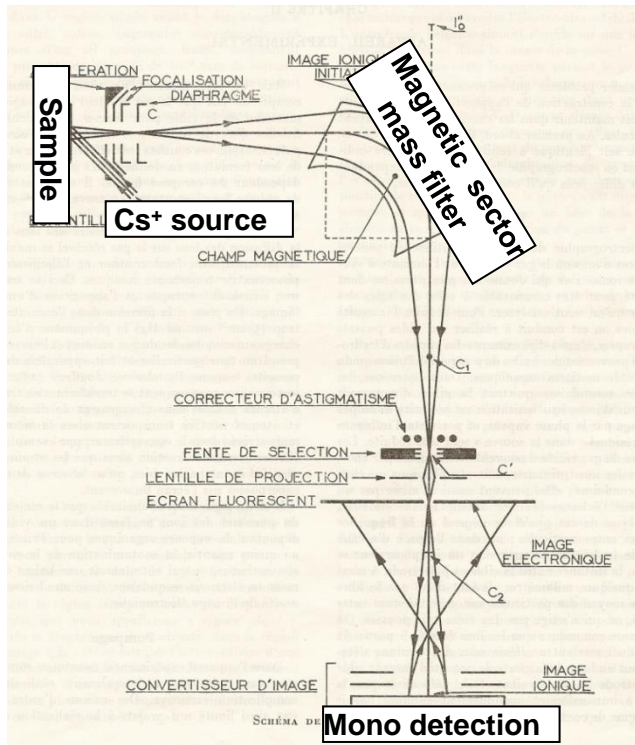
- Use of **reactive PI species** to enhance the **ionization yield** (O^- for + ions, Cs^+ for - ions)

- **All molecules are broken**, single atoms and clusters are ejected.

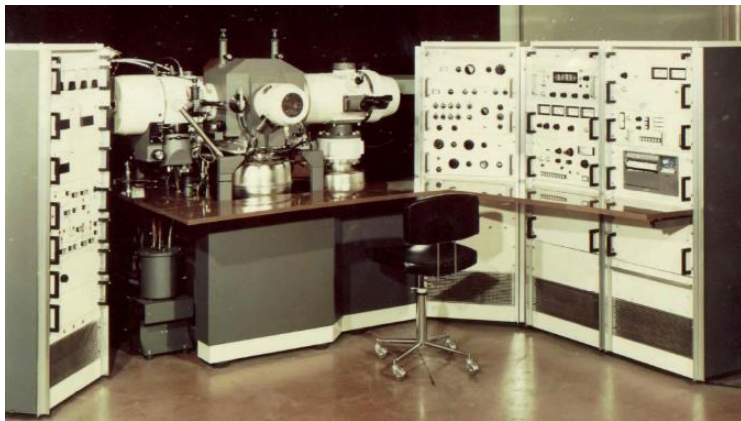
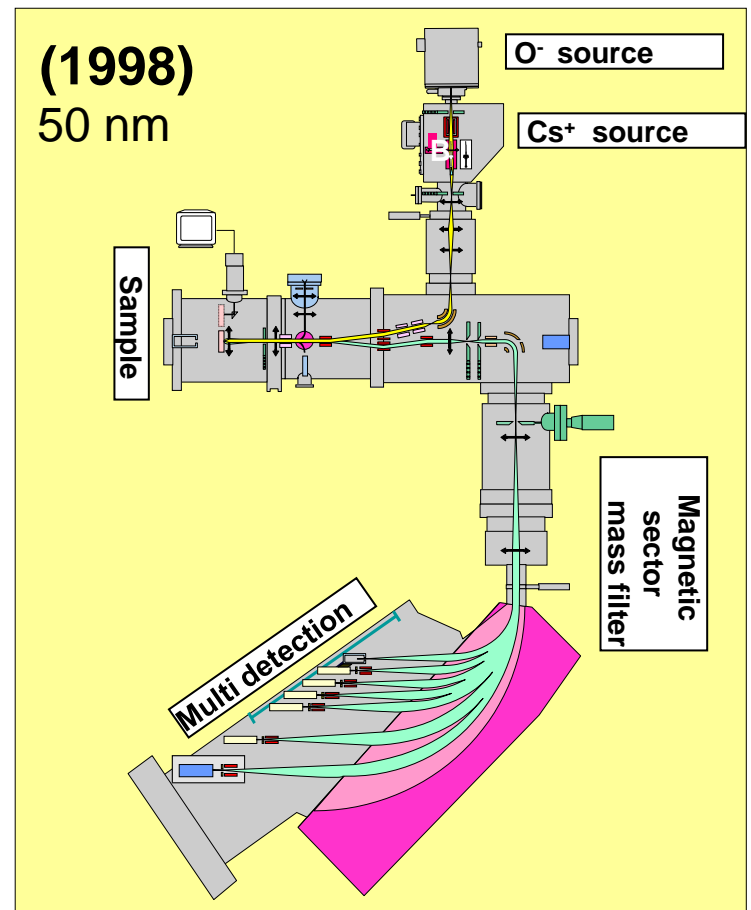
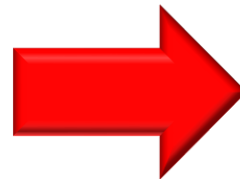
- A **small fraction is ionized** (+ or - charge), **SI** available for **Mass Spectrometry**.



The Secondary Ions, characteristic of the local composition, are collected, then separated in a magnetic sector analyzer according to their **mass/charge** ratio:
SIMS reveals **elemental** (H included) and **isotopic** surface composition



Prof. Slodzian

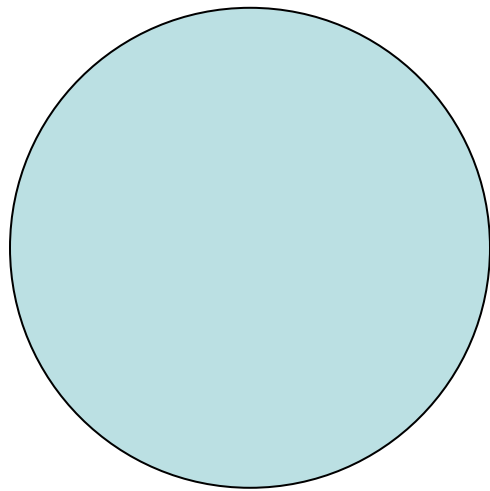


Cameca SMI 300, 1968



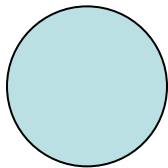
Cameca NanoSIMS50, 2000

The beam size of the microprobe



IMS 3f Ion Microprobe
2 – 3 μm

70 ‘



IMS 4f 500 nm

80 ‘



IMS 6f / Tof-SIMS 250 nm

90 ‘



NanoSIMS50 50 nm

2000 ‘

Nano Secondary Ion Mass Spectrometry (NanoSIMS)



The NanoSIMS 50L instrument
part of the new Mass Spectrometry Center in Pau (MARSS)



- Reactive primary ions; lateral resolution: **50nm** in Cs^+ , **200nm** (50nm) in O^-
- Parallel Detection: **7 masses**
- High **Sensitivity** together with **High Mass Resolution** and **small spot size**

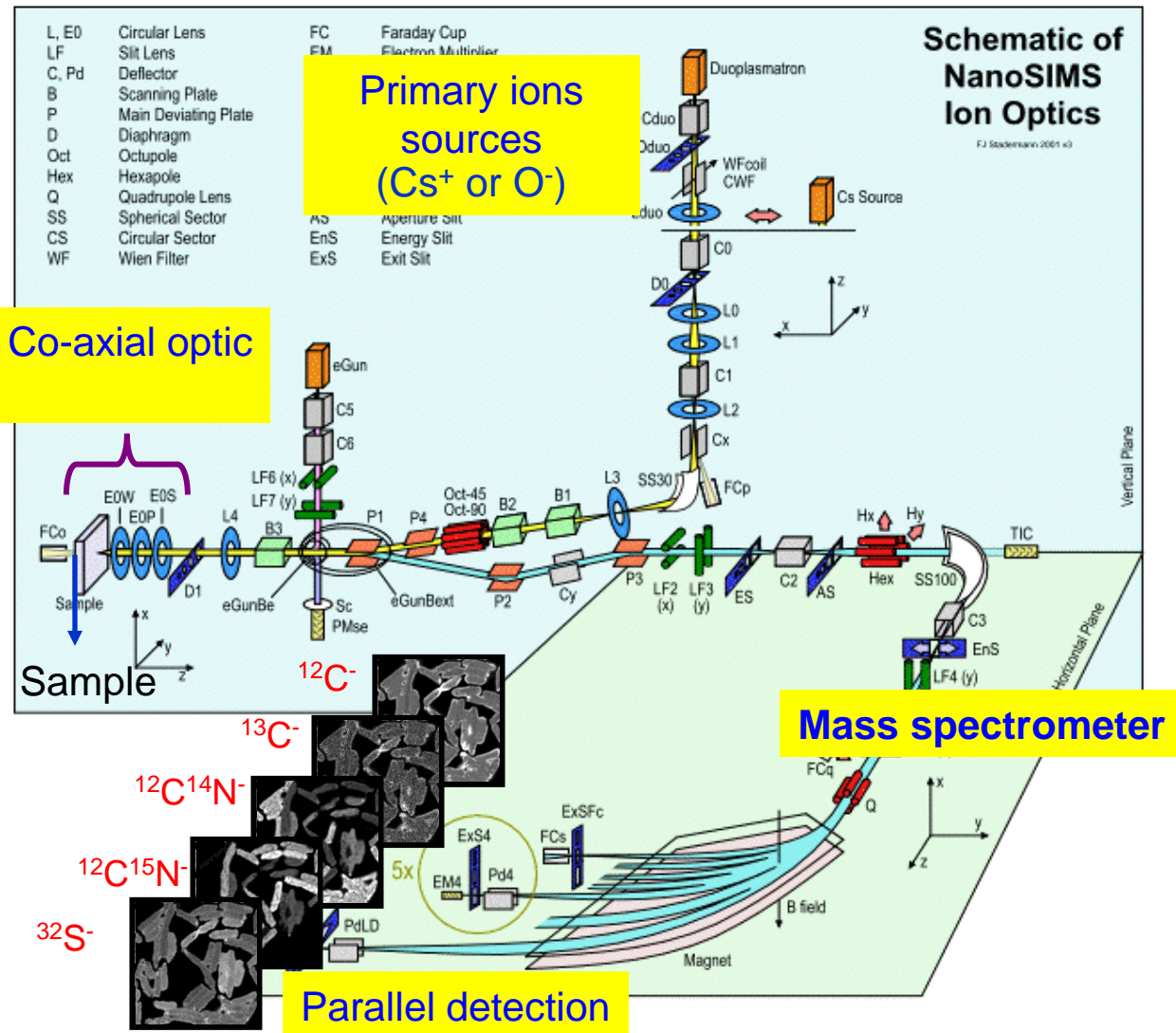
NanoSIMS : Ionic microprobe

(<http://presolar.wustl.edu/nanosims/schematic.html>)

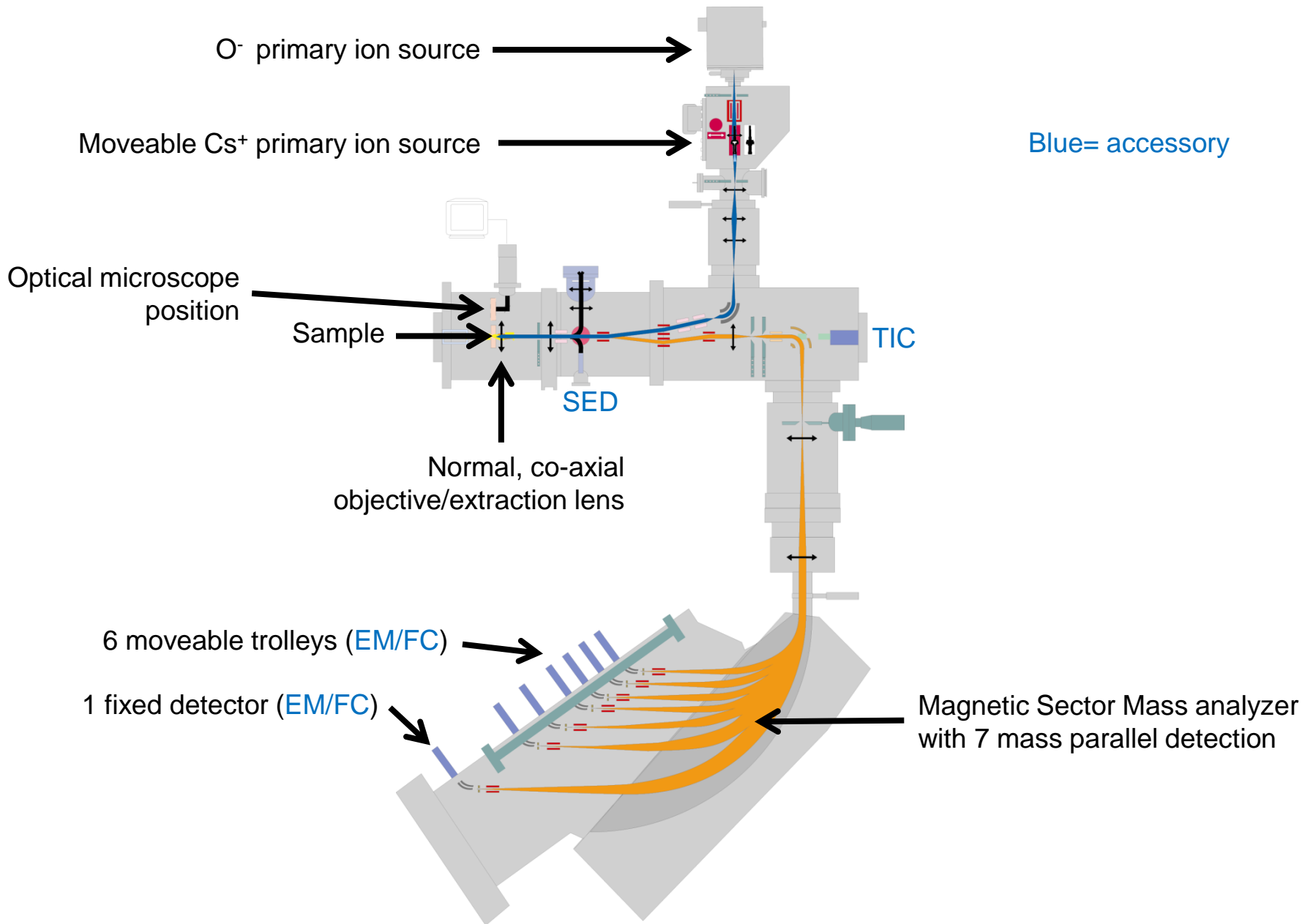
Analysis by scanning of a fine probe (50nm with Cs⁺)



Parallel ions counting and reconstruction of the elemental distribution (and isotopic)



NanoSIMS 50L scheme



SIMS Signal : Secondary ion intensity

$$I_{\text{ionM}} = T \cdot Y_i \cdot d_b \cdot S \cdot S_y \cdot [M]$$

d_b : bulk density,

Surface,

S_y : Rdt Ion I/II

T: instrumental transmission

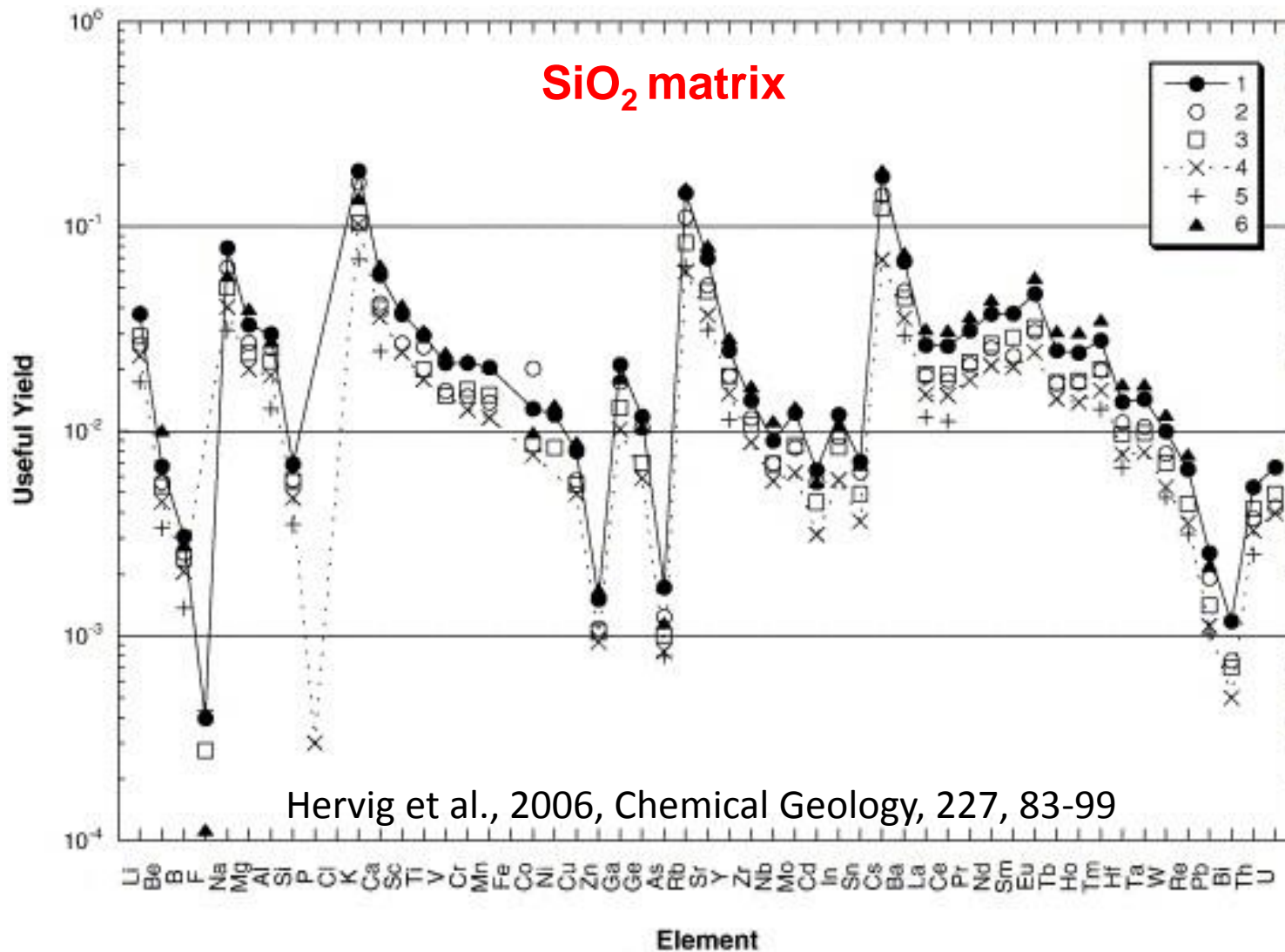
Y_i : ion yield of M_i $\frac{\text{nb ions } \mathbf{ionized}}{\text{nb atoms sputterized}}$

The SIMS ionization efficiency is called ion yield, defined as the fraction of sputtered atoms that become ionized.

T. $Y_i = \text{UY Useful Yield}$

$\frac{\text{nb ions } \mathbf{detected}}{\text{nb atoms sputterized}}$

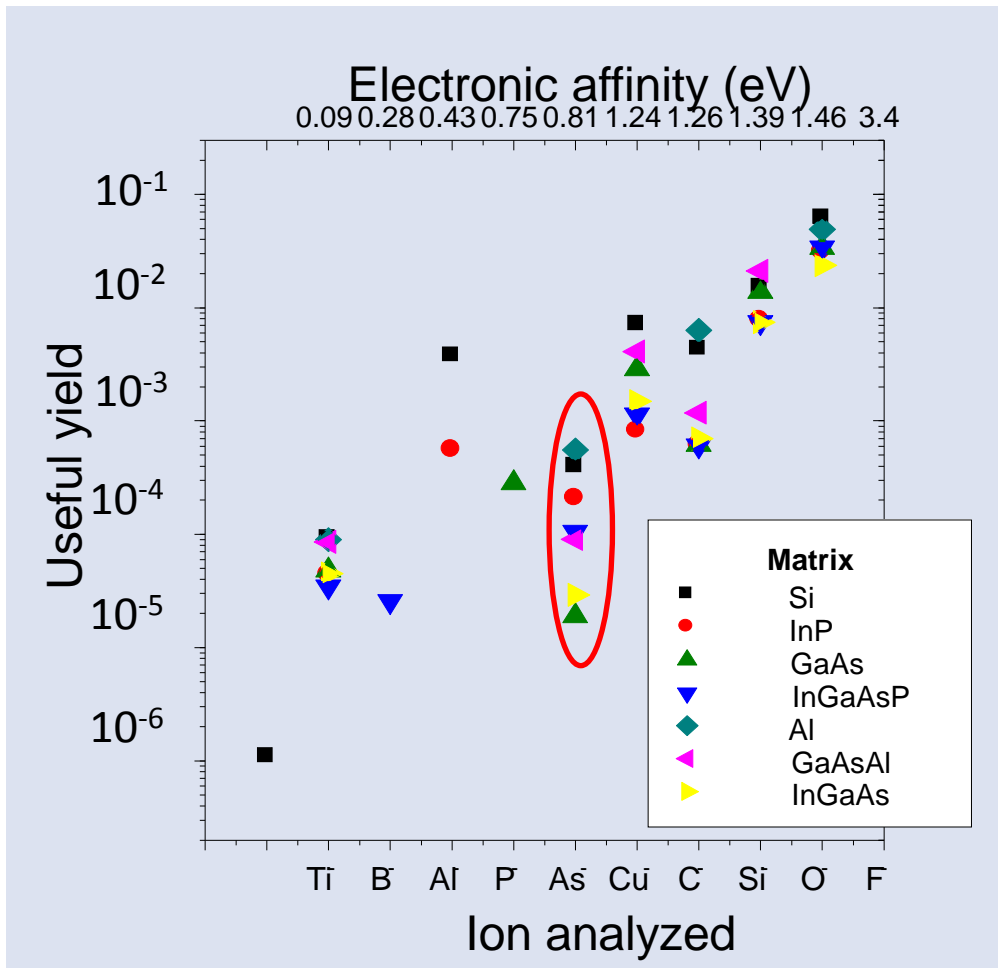
Useful Yield vs element



$$\text{UY } ^{12}\text{C} = \text{UY } ^{13}\text{C}$$

Isotopic ratio
not affected by
the UY

Useful Yield vs matrix



UY Useful Yield

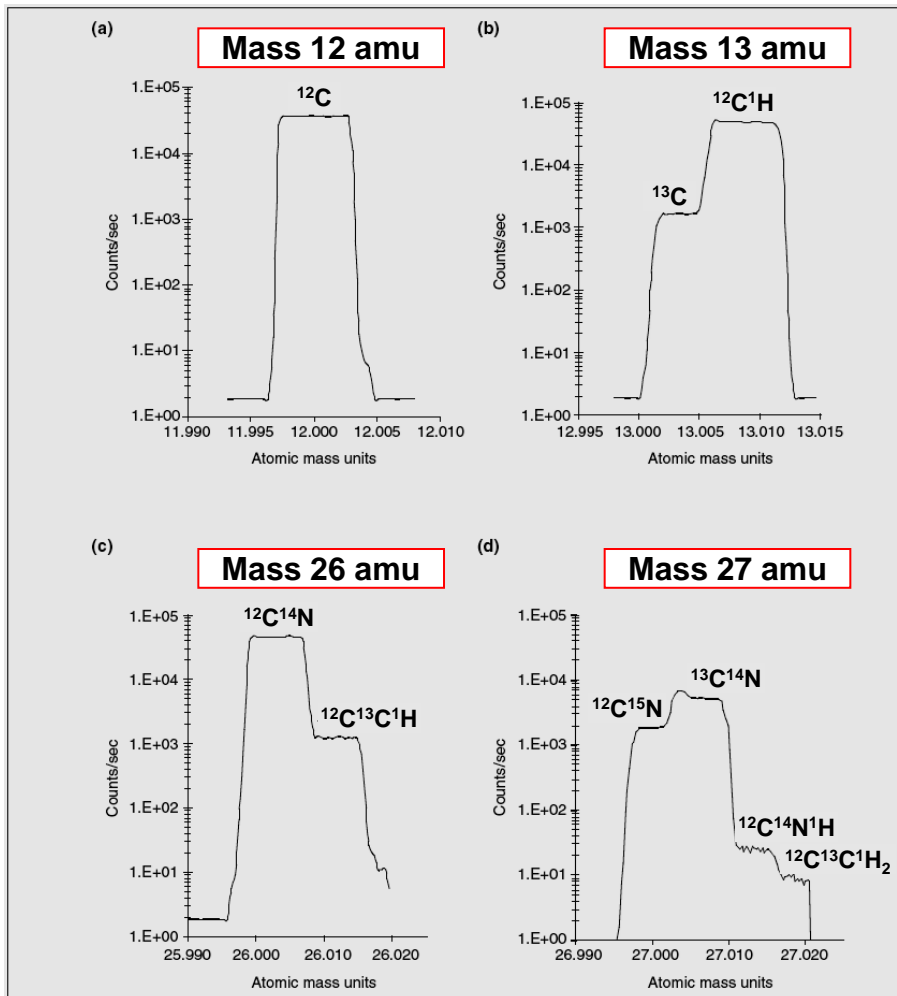
$$\frac{\text{nb ions detected}}{\text{nb atoms sputterized}}$$

Isotopic ratio not affected by the UY

Matrix effect

→ Quantification : difficult

High Mass Resolution need in SIMS



Section of embedded biological tissue.

In SIMS mass interferences are usually present at each unit mass.

High Mass Resolution is necessary to resolve such mass interference, specially for precise isotope ratios, quantitative measurements and trace level detection.

Flat top peak mode is used for better isotope ratio precision and reproducibility.

Note the dynamic range (log. scale) and peak shape.

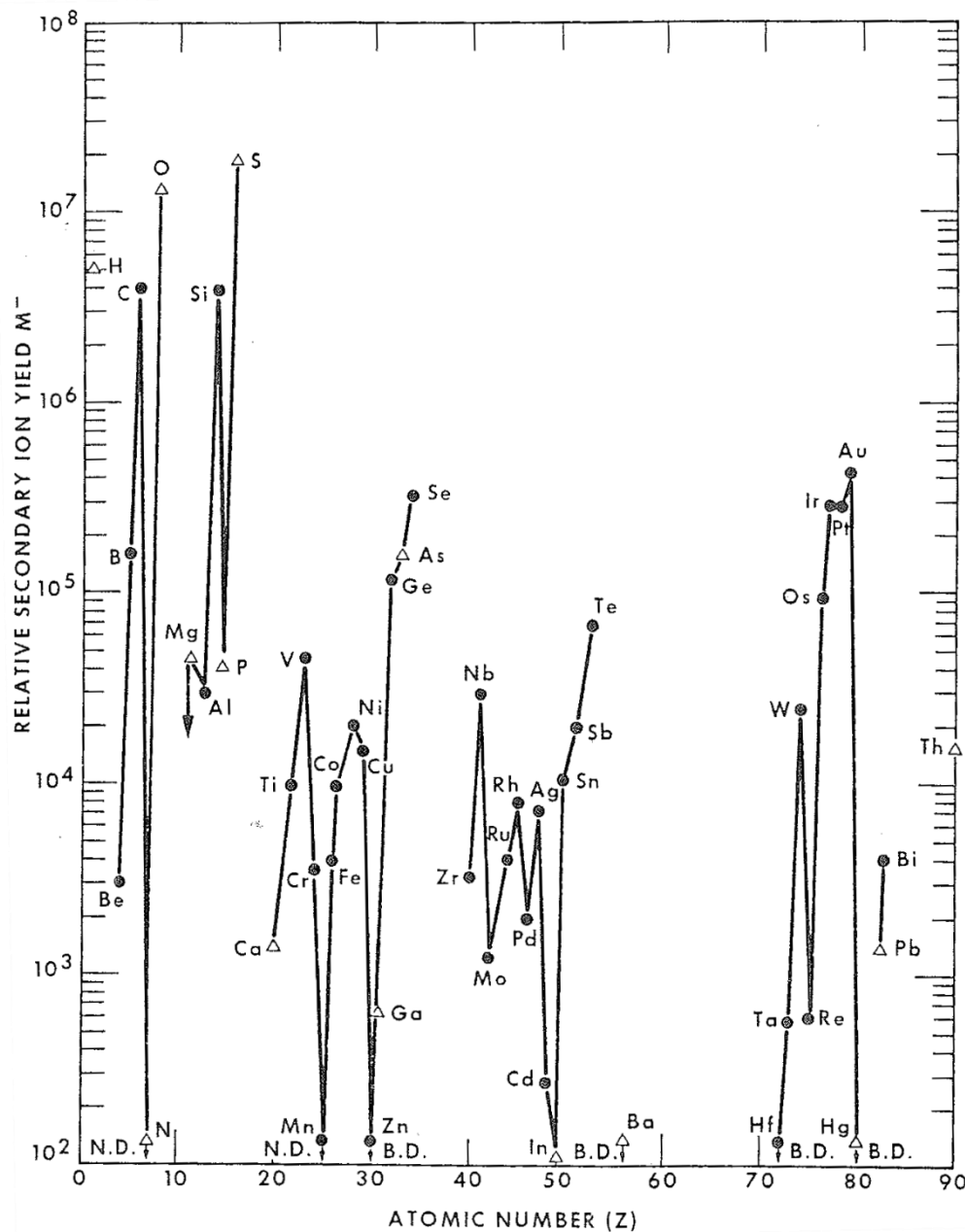
The uniqueness of the NanoSIMS is to keep nearly **full Transmission** (= High Sensitivity; **ppm level** for most elements) at **High Mass Resolution** ($M/\Delta M = 5000$) together with **High Lateral Resolution** ($< 50\text{nm}$).

Curves extracted from: High-resolution quantitative imaging of mammalian and bacterial cells using stable isotope mass spectrometry. C. Lechene et al, Journal of Biology 2006, Volume 5, Article 20.

Secondary Ion Yields -- Primary Beam Effects

		<div style="display: flex; align-items: center; gap: 10px;"> <div style="width: 15px; height: 15px; background-color: yellow; border: 1px solid black;"></div> O₂⁺ Primary Positive Secondary </div>										<div style="display: flex; align-items: center; gap: 10px;"> <div style="width: 15px; height: 15px; background-color: green; border: 1px solid black;"></div> Cs⁺ Primary Negative Secondary </div>																																			
H																			He																												
Li	Be											B	C	N	O	F			Ne																												
Na	Mg											Al	Si	P	S	Cl			Ar																												
K	Ca	Sc	Ti	V	Cr	Mn	Fe	Co	Ni	Cu	Zn	Ga	Ge	As	Se	Br			Kr																												
Rb	Sr	Y	Zr	Nb	Mo	Tc	Ru	Rh	Pd	Ag	Cd	In	Sn	Sb	Te	I			Xe																												
Cs	Ba	La	Hf	Ta	W	Re	Os	Ir	Pt	Au	Hg	Tl	Pb	Bi	Po	At			Rn																												
Fr	Ra	Ac																																													
<table border="1" style="margin: auto;"> <tr> <td>Ce</td><td>Pr</td><td>Nd</td><td>Pm</td><td>Sm</td><td>Eu</td><td>Gd</td><td>Tb</td><td>Dy</td><td>Ho</td><td>Er</td><td>Tm</td><td>Yb</td><td>Lu</td> </tr> <tr> <td>Th</td><td>Pa</td><td>U</td><td>Np</td><td>Pu</td><td>Am</td><td>Cm</td><td>Bk</td><td>Cf</td><td>Es</td><td>Fm</td><td>Md</td><td>No</td><td>Lr</td> </tr> </table>																				Ce	Pr	Nd	Pm	Sm	Eu	Gd	Tb	Dy	Ho	Er	Tm	Yb	Lu	Th	Pa	U	Np	Pu	Am	Cm	Bk	Cf	Es	Fm	Md	No	Lr
Ce	Pr	Nd	Pm	Sm	Eu	Gd	Tb	Dy	Ho	Er	Tm	Yb	Lu																																		
Th	Pa	U	Np	Pu	Am	Cm	Bk	Cf	Es	Fm	Md	No	Lr																																		

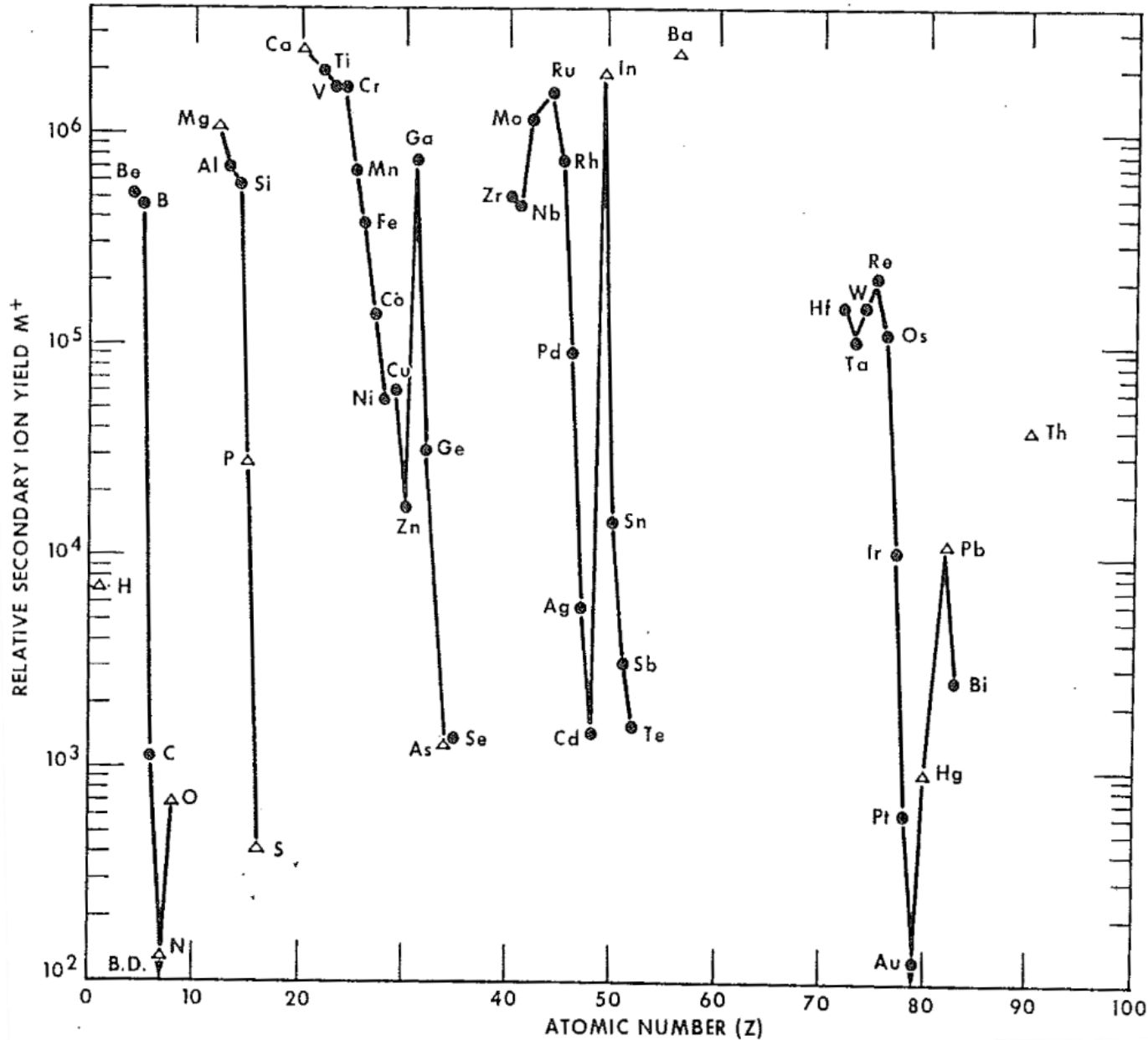
Relative negative ion yield for a Cs⁺ ion source in SIMS



Classic NanoSIMS application for cell imaging:
C, N (via CN⁻), **O, S, P, Se** and their stable isotopes for tracer studies.

Storms, H.A., K.F. Brown, and J.D. Stein
Evaluation of a Cesium Positive-Ion Source for Secondary Ion Mass-Spectrometry.
Analytical Chemistry, 1977. **49**(13): p. 2023-2030.

Relative positive ion yield for a O^- ion source in SIMS



Imaging of major
And trace elements
should be possible:
Ca, Mg, Mn, Cr, Cu

...

New O- source developed on our instrument by CAMECA

Oxygen adjustment

Oxygen inlet

RF cable

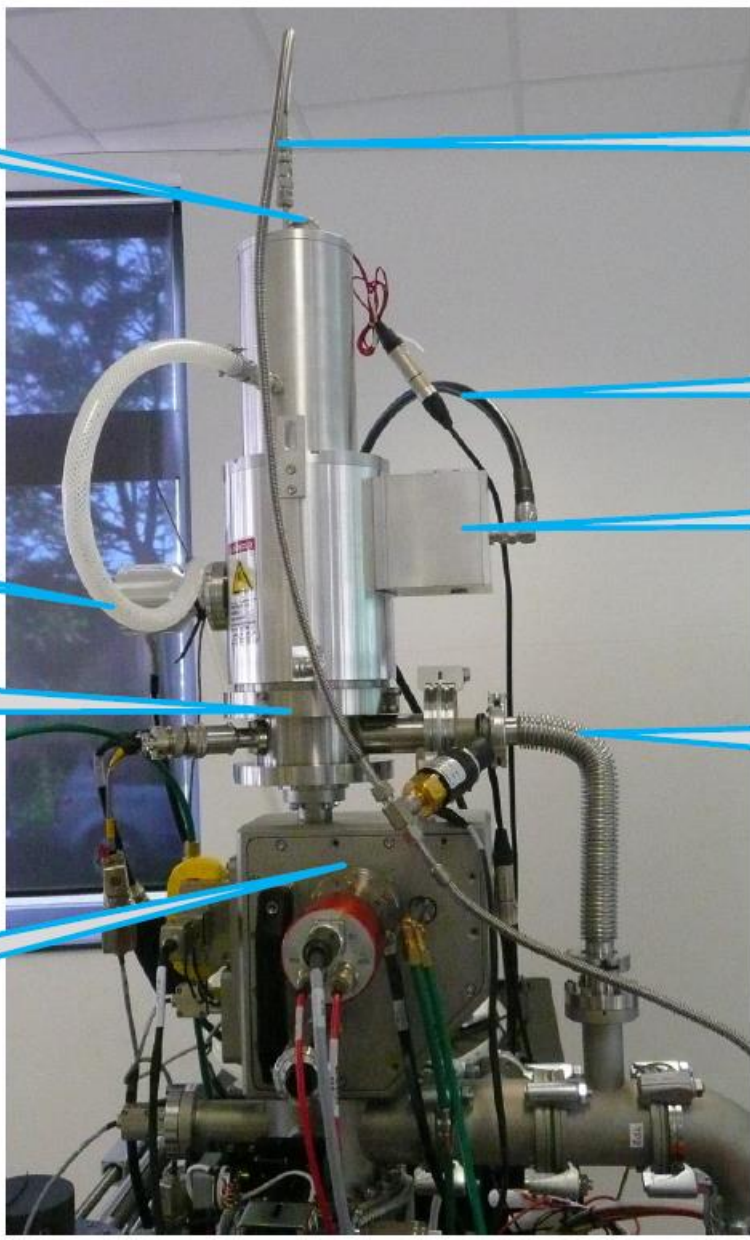
Cooling

RF match box

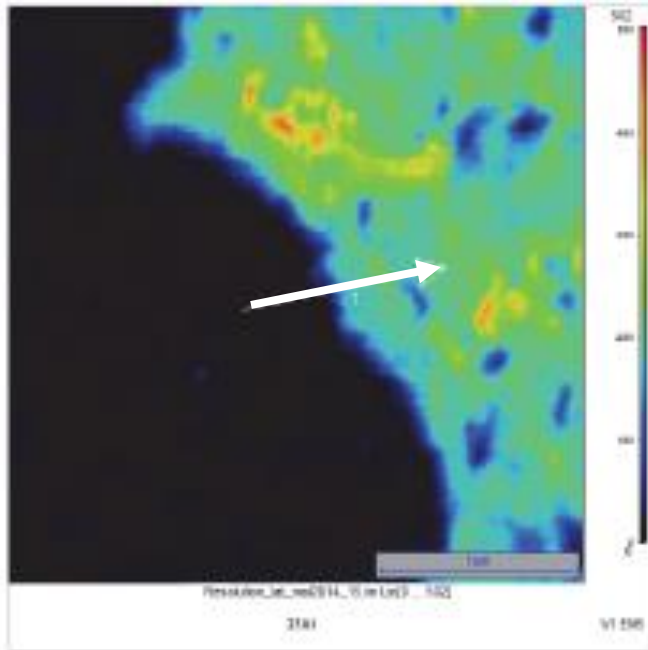
Mechanical interface

Extraction area pumping

Source chamber



Determination of the size of the O⁻ primary ion beam (probe size)

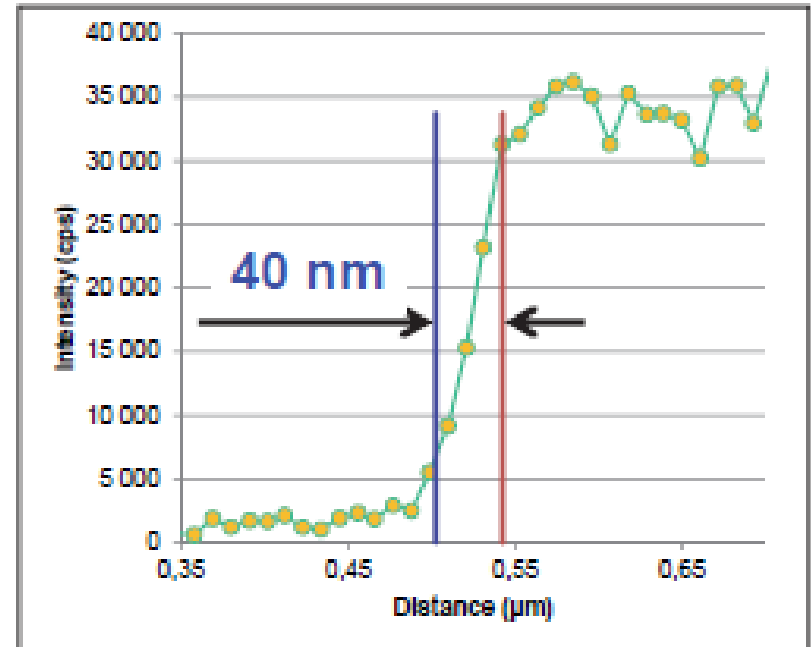


Al/Si oxide grain sample

Image size: 3 x 3 μm

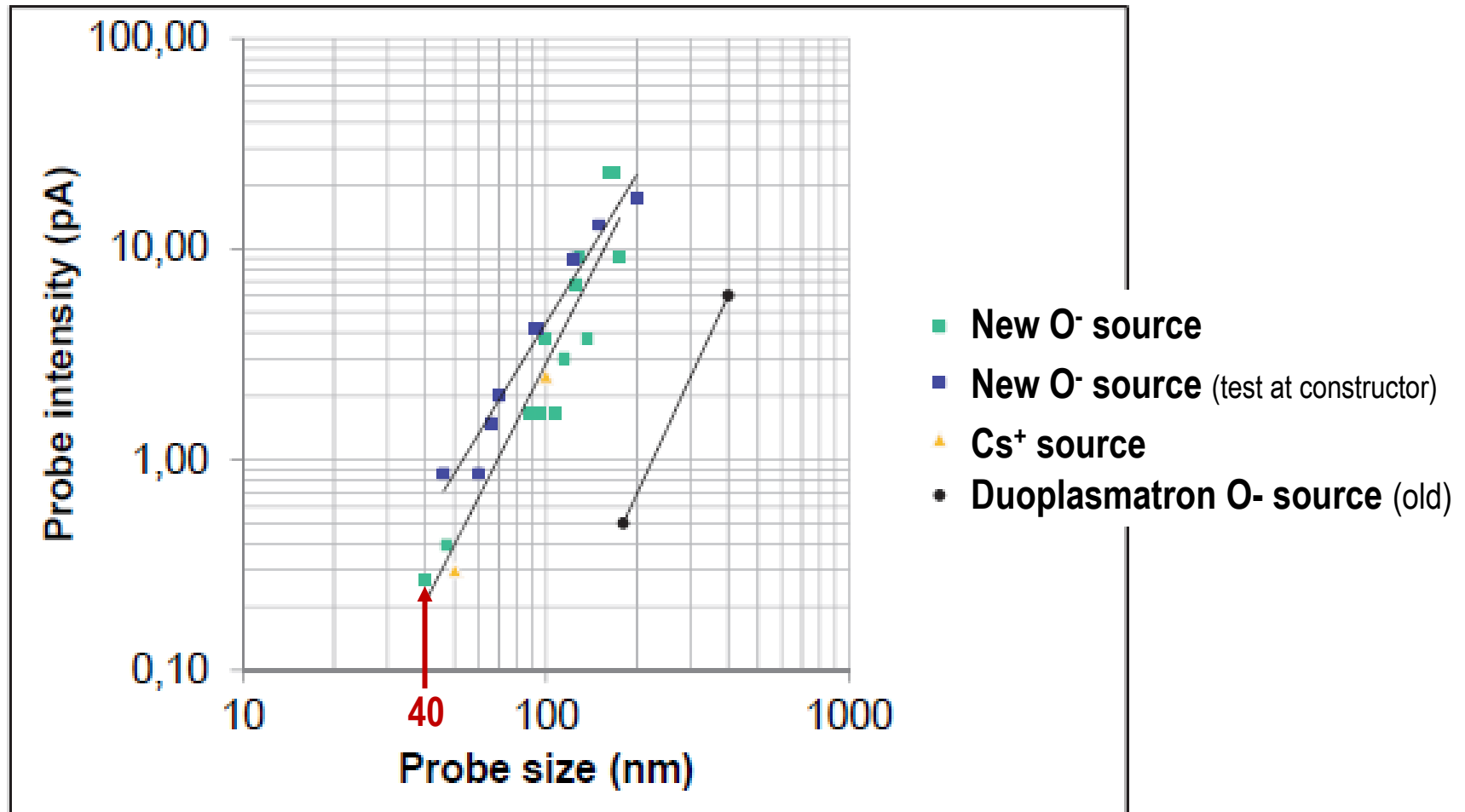
Probe size: 40 nm (16-84%)

Probe intensity: 0.3 pA



Line scan (left image) showing **intensity variation** from **16 to 84 %**:
determination of **probe size** (resolution)

Comparison of NanoSIMS primary ion sources



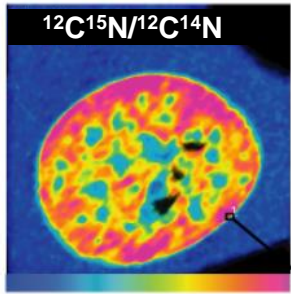
The new oxygen ion source show similar sensitivity and even better resolution (**40 nm**) as the cesium ion source and by far better characteristics (resolution and sensitivity) than the old oxygen ion source.

Advantages of the new O⁻ source

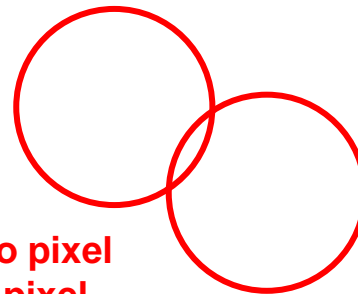
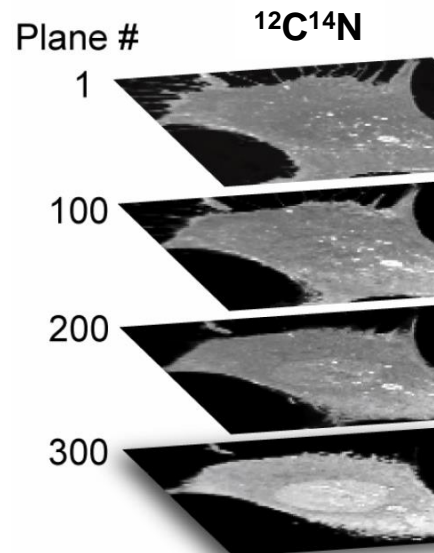
- **Higher beam density** = better sensitivity for metals (Ca, Fe, Cu, Mn....)
- **Higher lateral resolution** than conventional Oxygen sources = sharper images enabling the observation of smaller details
- **Less maintenance** = less instrument downtime
- **Stability:** < 1.6 % over 14h; **lifetime:** > 1000h (up to now)

The NanoSIMS: a scanning Ion Microprobe with a multicollection mass spectrometer

Ultra High Vacuum (→ dehydrated sample)



1454	1449	1411	1347	1239
226861	224906	219379	213200	206396
0.637%	0.640%	0.639%	0.628%	0.597%
1500	1414	1341	1163	994
222784	220467	212399	204234	198130
0.669%	0.637%	0.627%	0.566%	0.499%
1414	1265	1153	974	803
219466	212200	204972	197007	194159
0.640%	0.593%	0.563%	0.494%	0.412%
1326	1108	939	820	789
211556	204599	198366	193922	192569
0.623%	0.539%	0.471%	0.421%	0.408%



Comparison of different MS imaging techniques

Organic information
Mass resolving power



	Dynamic SIMS	Static SIMS	(MA)LDI-TOFMS	FT-MS (MALDI)
Data	Elements	Elements + organic fragments	Pseudo-molecular (low fragmentation)	Pseudo molecular (low fragmentation)
Lateral resolution	50 – 100 nm	200 nm - 10 mm	10 - 50 mm	50 - 200 mm
Field of View	80 /600 mm	9 cm	15 cm	n.a.
Typical mass range	< 250 Da	< 1 000 Da	> 100 000	> 10 000
Mass resolving Power	15 000	10 000 (28 Da)	20 000 (2000 Da)	> 1 000 000 (1000 Da)



Lateral resolution
Sensitivity
Fragmentation

The ultimate microscope

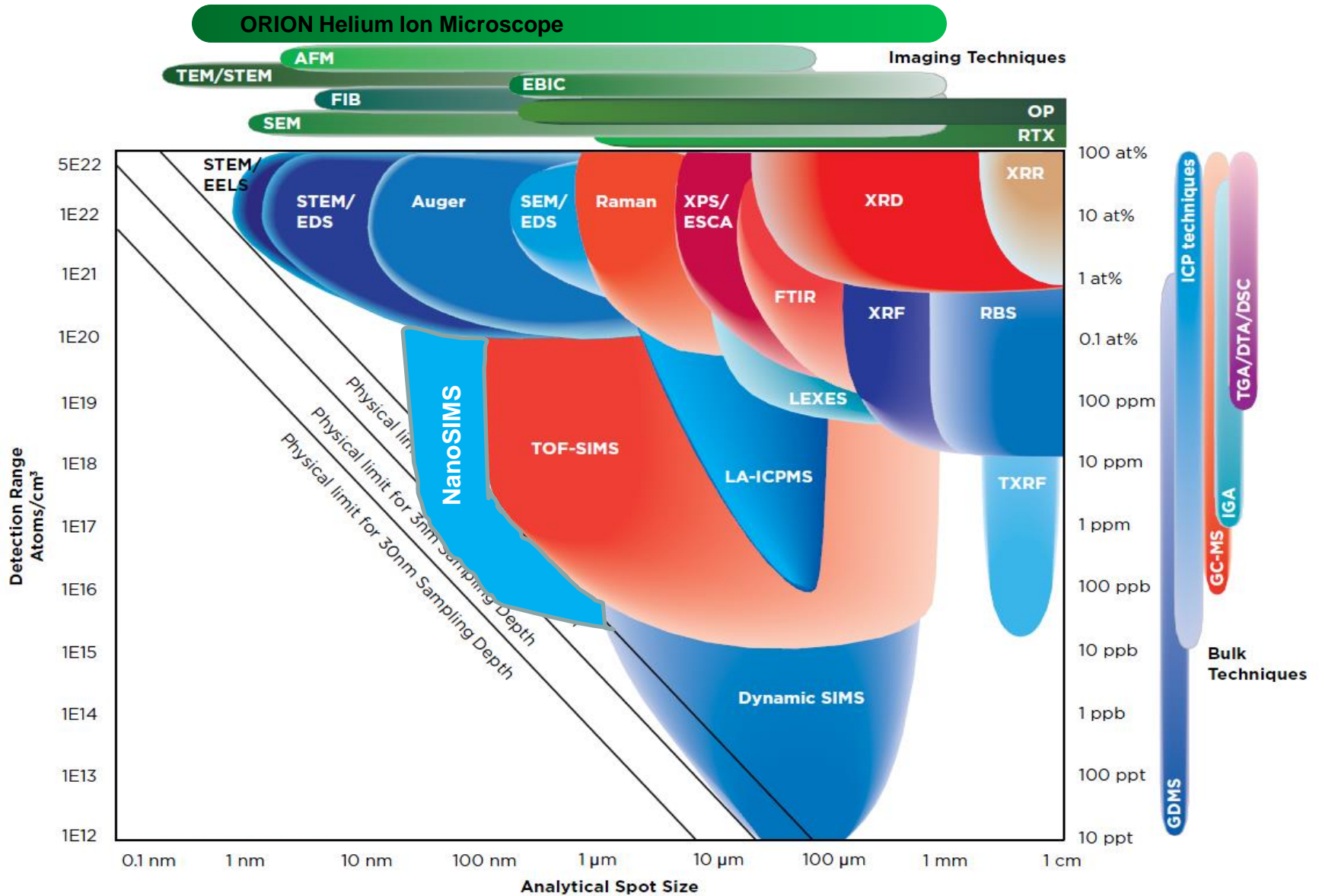


Figure: by courtesy from Jean-Nicolas Audinot, Centre de Recherche Gabriel Lippmann, Luxembourg

Strong points of NanoSIMS

- **Allmost all Elements** (from H, D, T,... up to Pu)
- **Very High Sensitivity**: down to ppb in spot analysis, ppm in imaging,
- **Quantification possible**, but difficult (Relative Sensitivity Factors)
- **Isotopic composition** (signature of origin and history in **astrophysics**, of past climate and origin and history in **geology**, of path and activity in **biology**, of path in **materials**)
- **Localized**, micro-analysis: down to 50nm lateral resolution (NanoSIMS), access to **3D** analysis with depth resolution of 10-15nm with NanoSIMS.
- Large **sample size** is possible for navigation and finding areas of interest
- Minimal **sample preparation** for solids (polishing and metal coating for minerals), dehydration or **TEM-like preparation for biological tissues**.

Biological applications

Use of the novel oxygen primary ion source for the localization of major (**Na, Ca, Mg**) and trace (**Fe, Cu, Mn, Zn**) metals
Involved in physiological processes in plant cells

Biological sample preparation

TEM-like preparation (transmission electron microscopy)

The sample is analyzed at room temperature **under vacuum**

It must be **dehydrated and fixated**.

Flat samples are required. *Thin sections* (300-400nm) are preferable to avoid sample charging under ion bombardment. **Ultramicrotome**.

Sample preparation will then depend on application:

- **chemical fixation**, resin embedding, thin sectioning.
- **fast freezing**, cryo-substitution, resin embedding and thin section deposited on metal or silicon substrate.

Biological sample preparation

less
redistribution of
highly diffusable
trace metals !

Chemical fixation

Glutaraldehyde
Formaldehyde
Osmium tetroxide

Cryo fixation

high pressure freezer
tissues
(up to 6 mm diameter,
200 μ m thick)



Dehydration

Solvent baths (acetone or ethanol/water)
with increasing solvent concentrations

Dehydration

Cryo-substitution
lyophilization



Resin embedding

Solvent baths with increasing
resin concentrations

Resin embedding

Ultramicrotomy

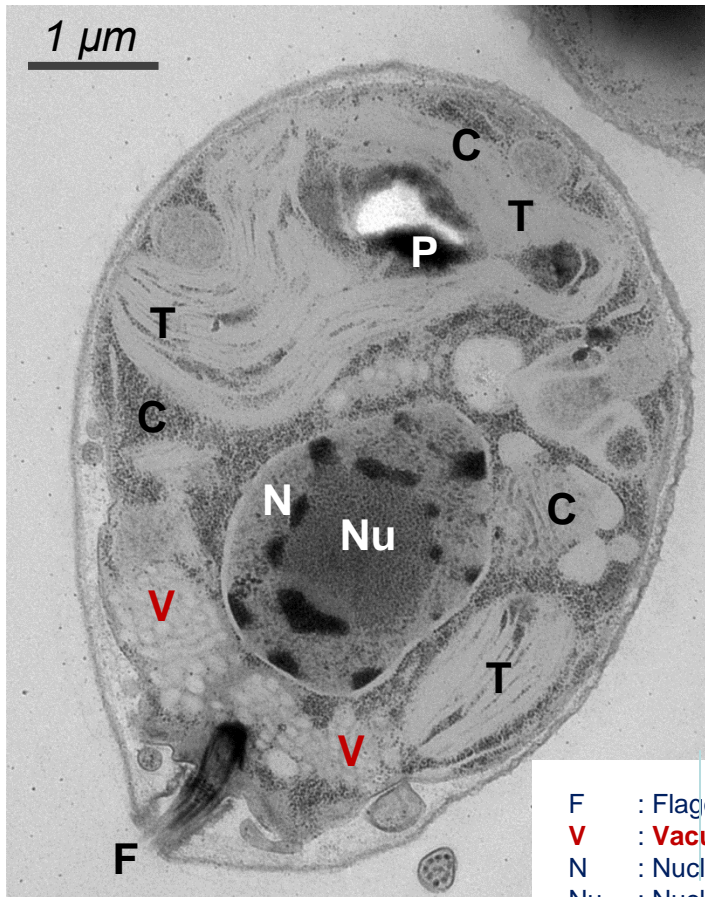
300-400 nm sections



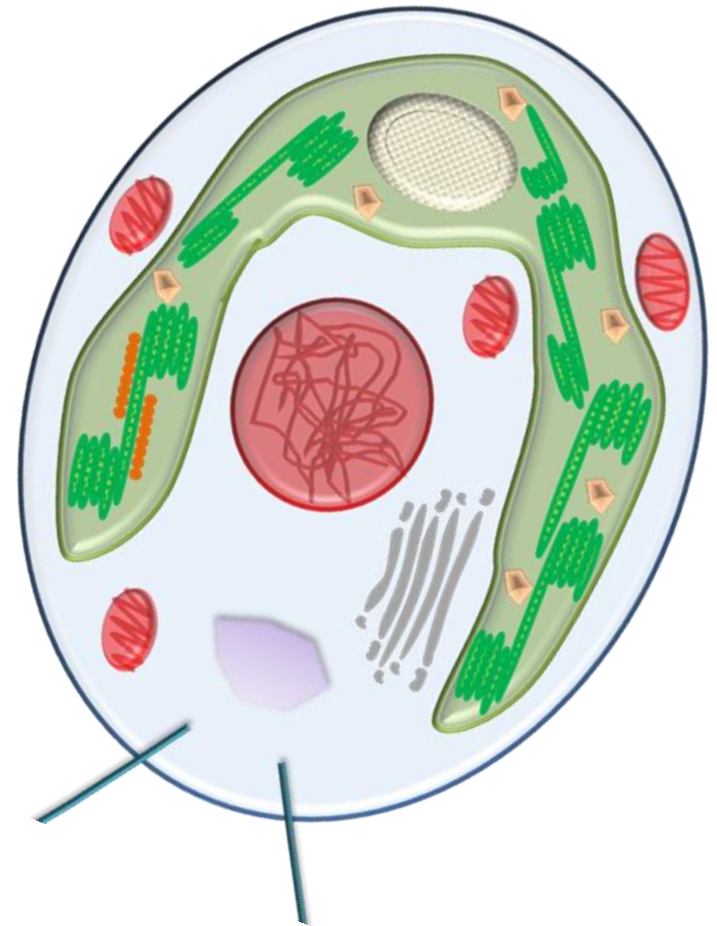
Application to a model organism: *Chlamydomonas reinhardtii* cells (unicellular green algae)

TEM analysis (70 nm thin section)
resolution down to 1 nm

Comparison with schematic view



- F : Flagella
- V : **Vacuoles**
- N : Nucleus
- Nu : Nucleolus
- C : Chloroplast
- T : Thylakoid
- P : Pyrenoid



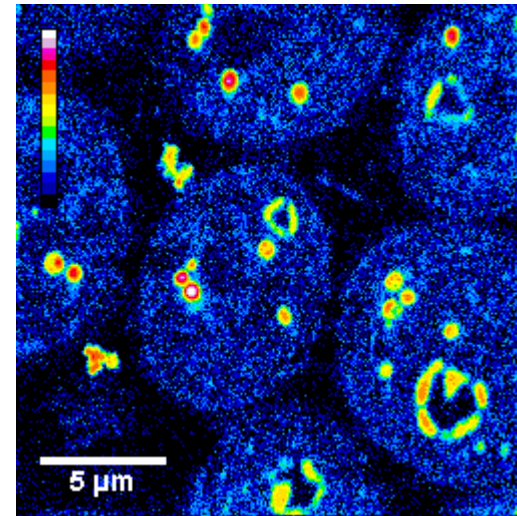
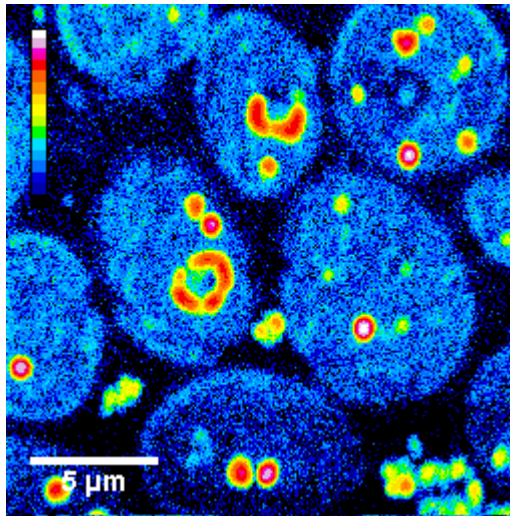
NanoSIMS analysis of *Chlamydomonas reinhardtii* cells

Comparison conventional Duoplasmatron O⁻ ion source and novel O⁻ ion source

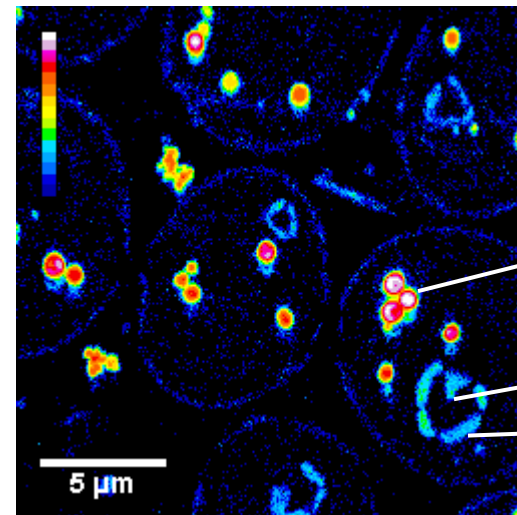
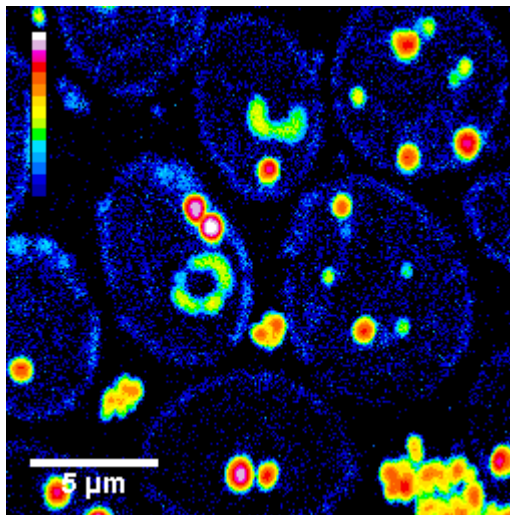
Duoplasmatron O⁻ ion source

New O⁻ ion source prototype

²³Na

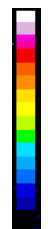


⁴⁰Ca



300 nm
thin sections

relative intensity: Max

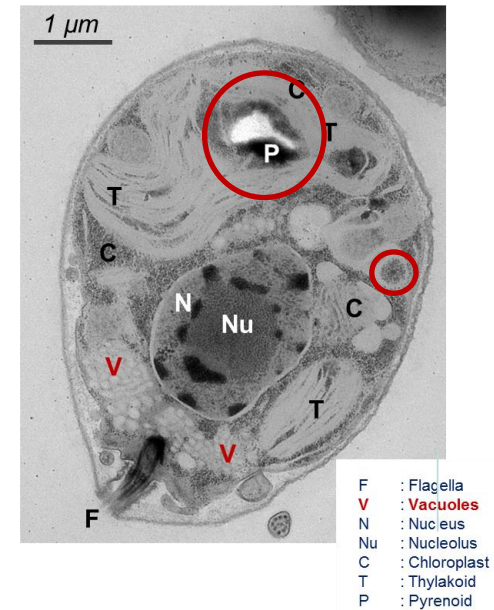
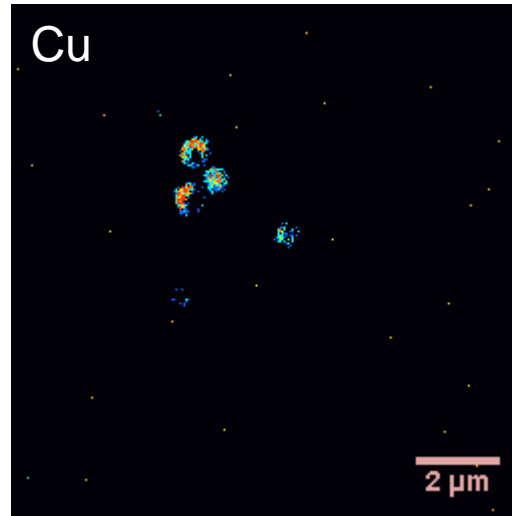
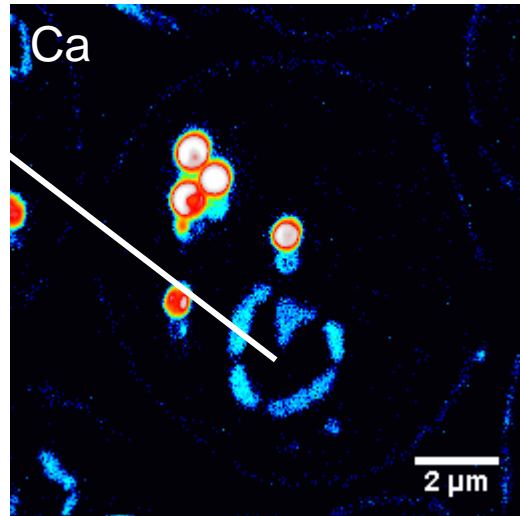
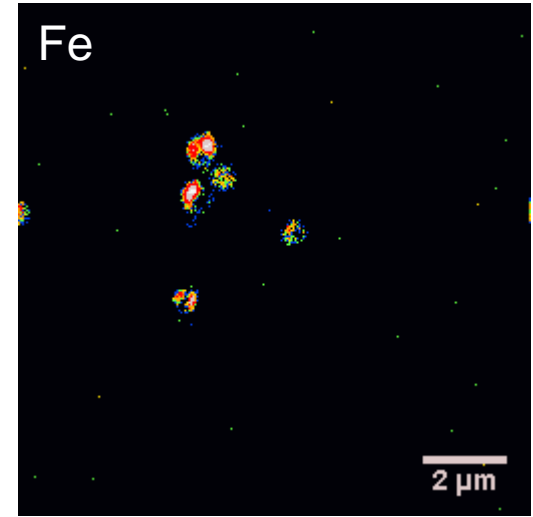
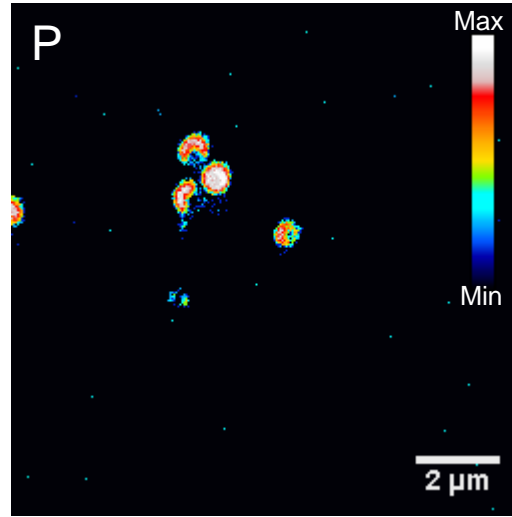
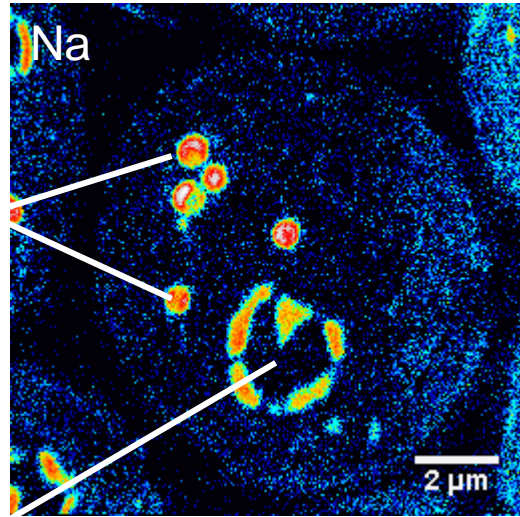


Min

20 x 20 μm
12 min
256x256 pixel
1 plane

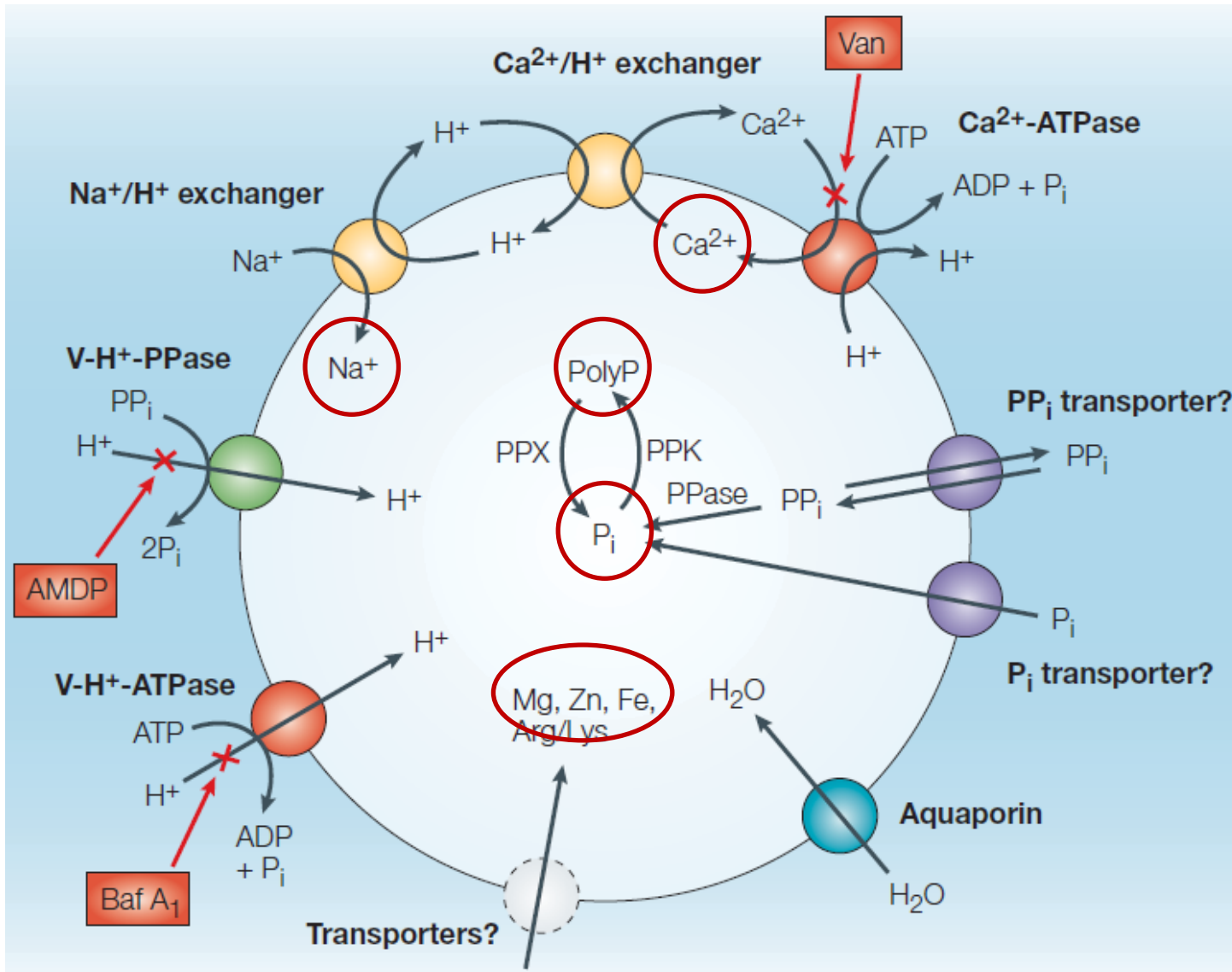
Subcellular element imaging by NanoSIMS (new ion source)

Pyrenoid
with starch plates
Acidocalcisomes
Granules ?



Single cell imaging: 12 x 12 μm, 22 min, 512x512 pixel, 2 planes

Scheme of a Acidocalcisome



Biological applications

Use of the conventional cesium primary ion source for isotopic tracer experiments (C-13):
Investigation of physiological processes
under Cd stress

Parallel TEM/X-EDS experiments

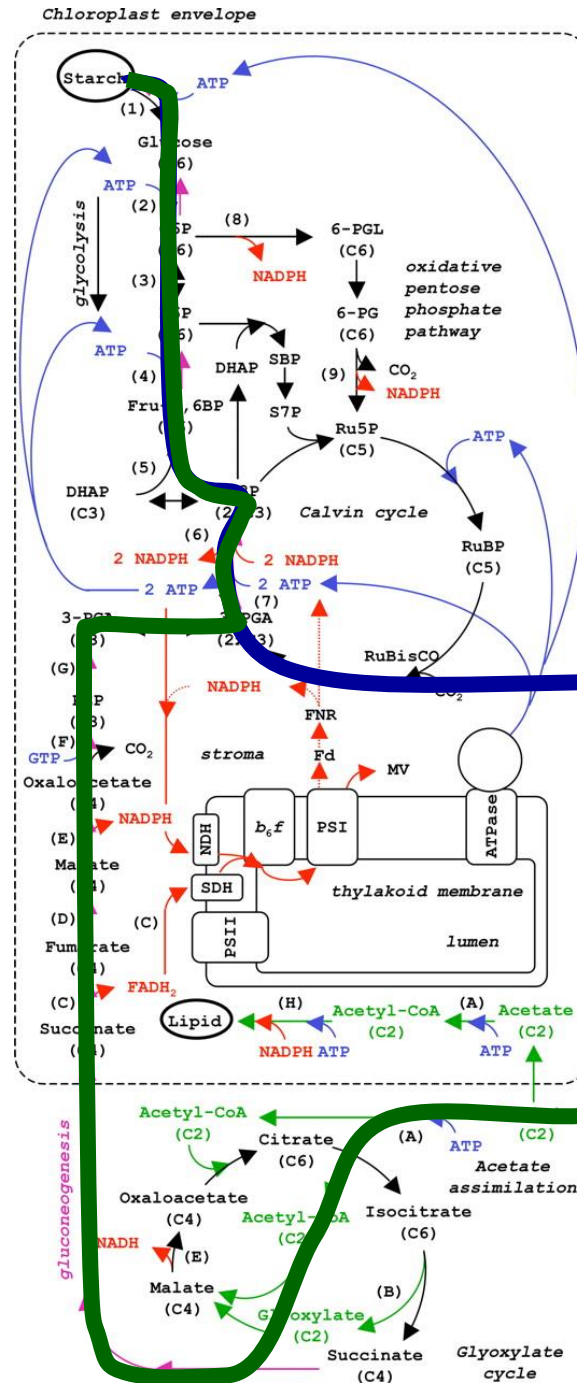
Investigation of the CO₂ fixation Under Cd stress

NanoSIMS with Cs source
TEM/X-EDS

Simplified schematic drawing of metabolic pathways in *Chlamydomonas reinhardtii*

Xenie Johnson, and Jean Alric J.
Biol. Chem. 2012;287:26445-26452

starch



Enzyme
RuBisCo

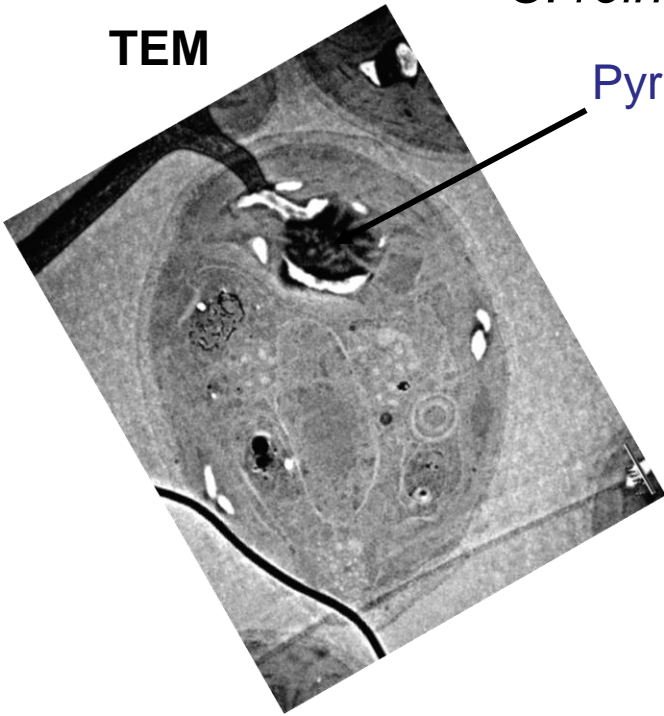
CO₂ : ¹²C

acetate : ¹³C

jbc

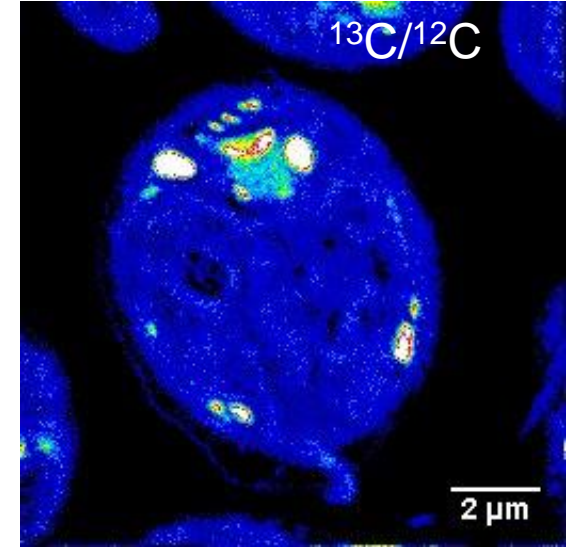
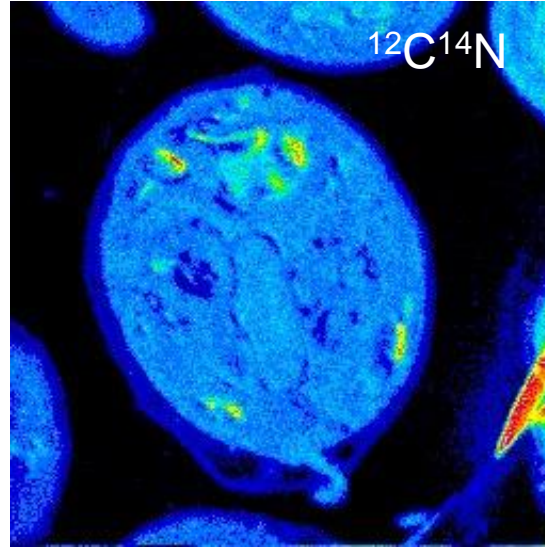
C. reinhardtii wt in mTAP medium 70 μ M Cd

TEM

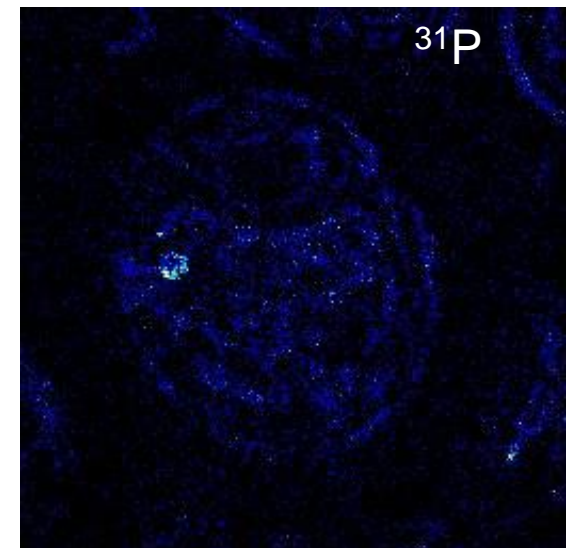
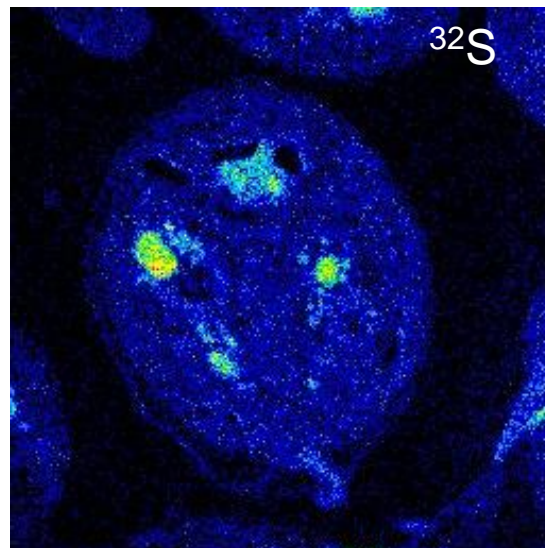
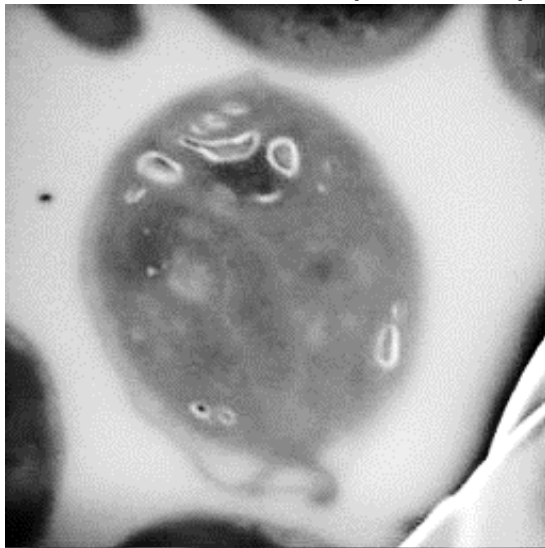


Pyrenoid: accumulation of RuBisCo

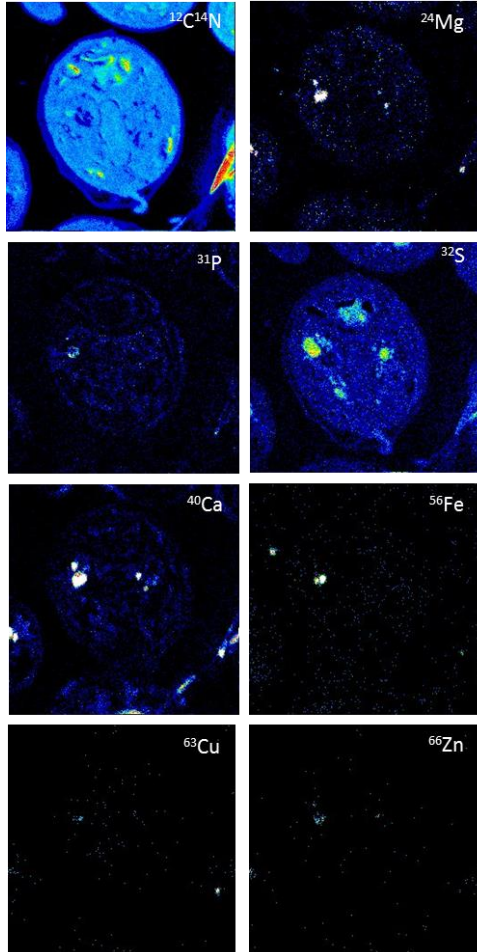
NanoSIMS



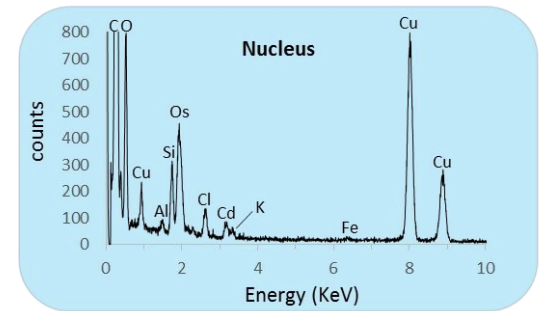
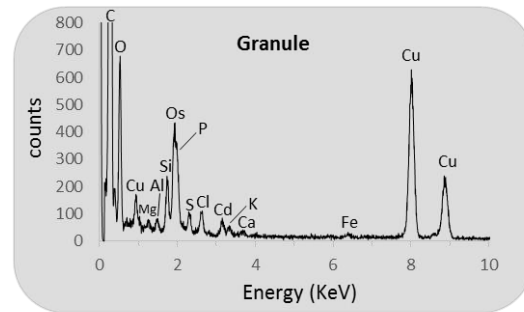
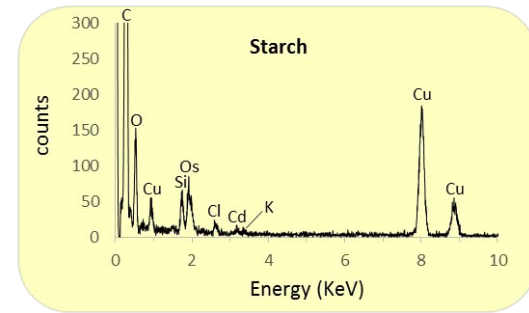
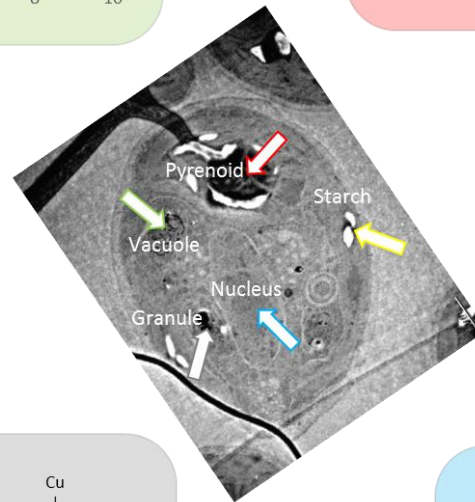
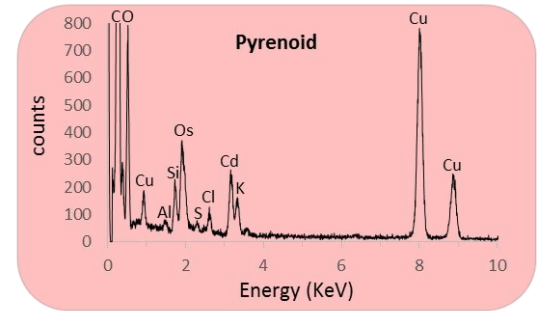
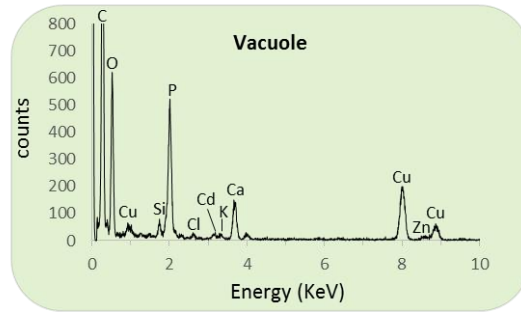
SEM (NanoSIMS)



NanoSIMS

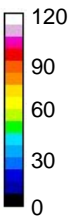


TEM/X-EDS



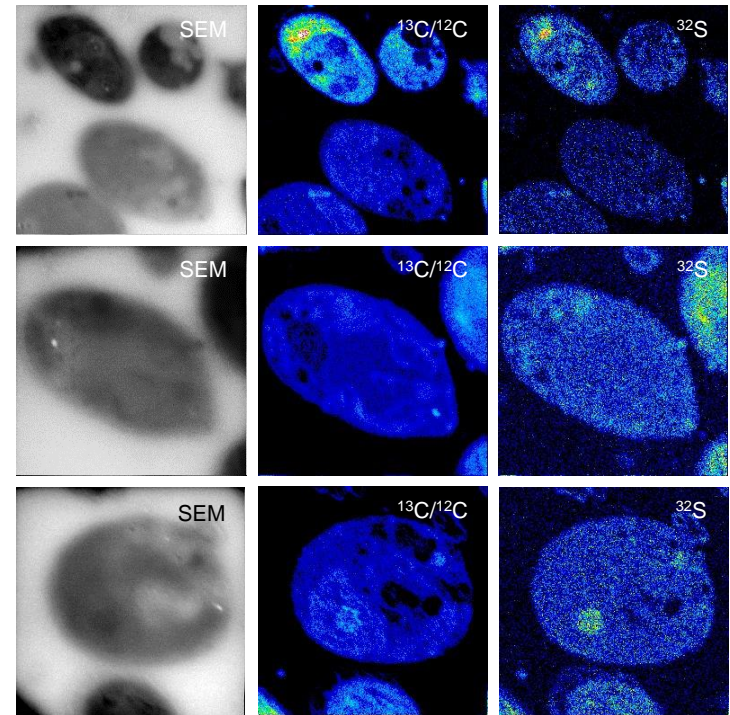
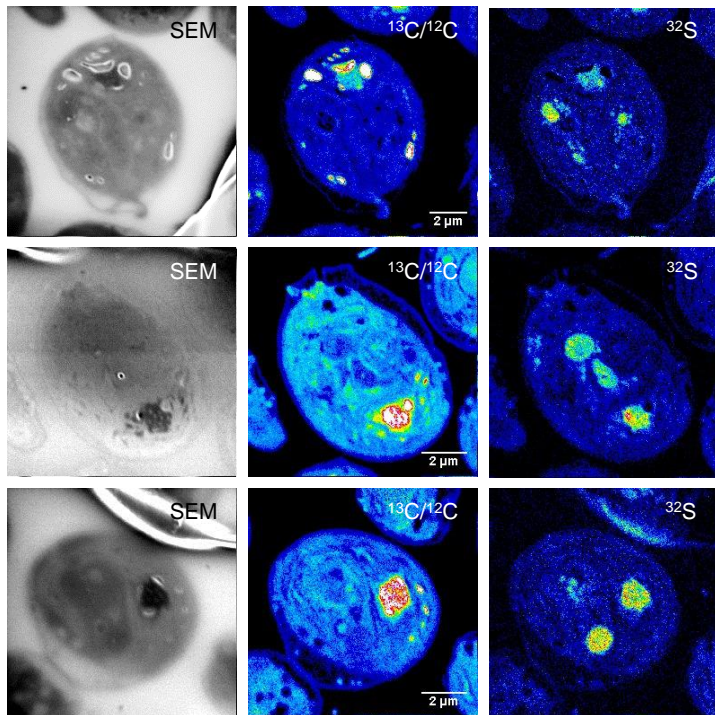
C. reinhardtii
wt in mTAP
medium
70 μ M Cd

$^{13}\text{C}/^{12}\text{C}$
x 100



A vertical color scale legend for the $^{13}\text{C}/^{12}\text{C}$ ratio. The scale ranges from 0 (black) to 120 (white), with intermediate markers at 30 (blue), 60 (green), and 90 (red).

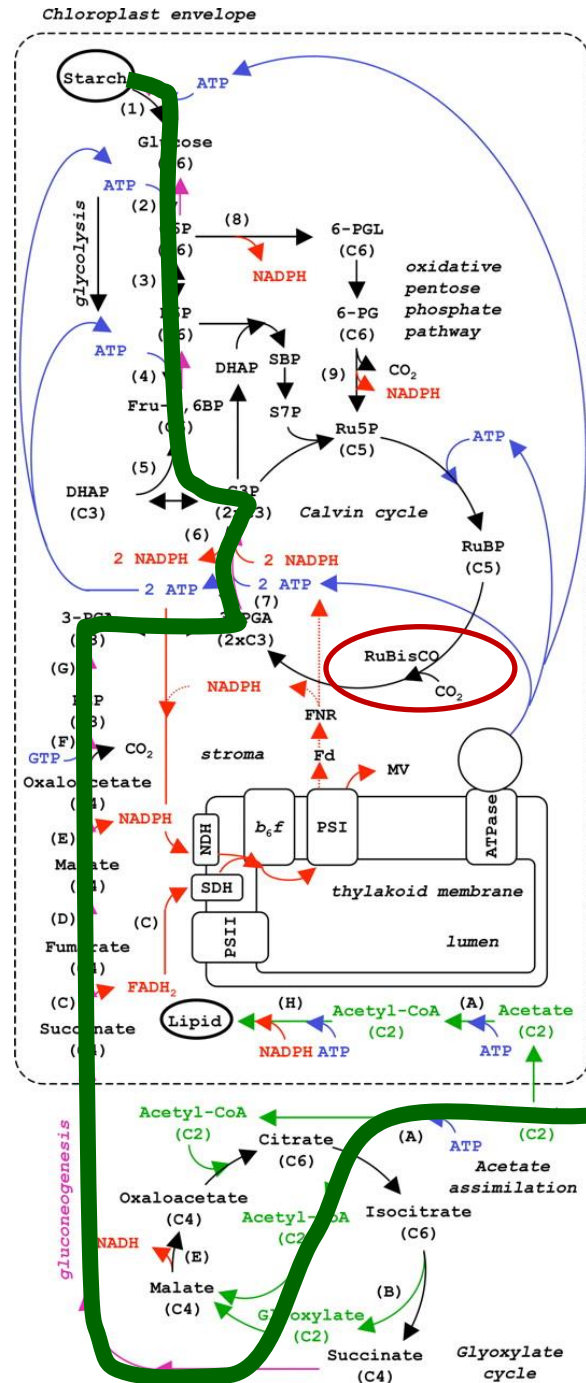
C. reinhardtii
wt in mTAP
medium
control



Simplified schematic drawing of metabolic pathways in *Chlamydomonas reinhardtii*

Xenie Johnson, and Jean Alric J.
 Biol. Chem. 2012;287:26445-26452

starch



RuBisCo blocked by Cd



acetate : ¹³C

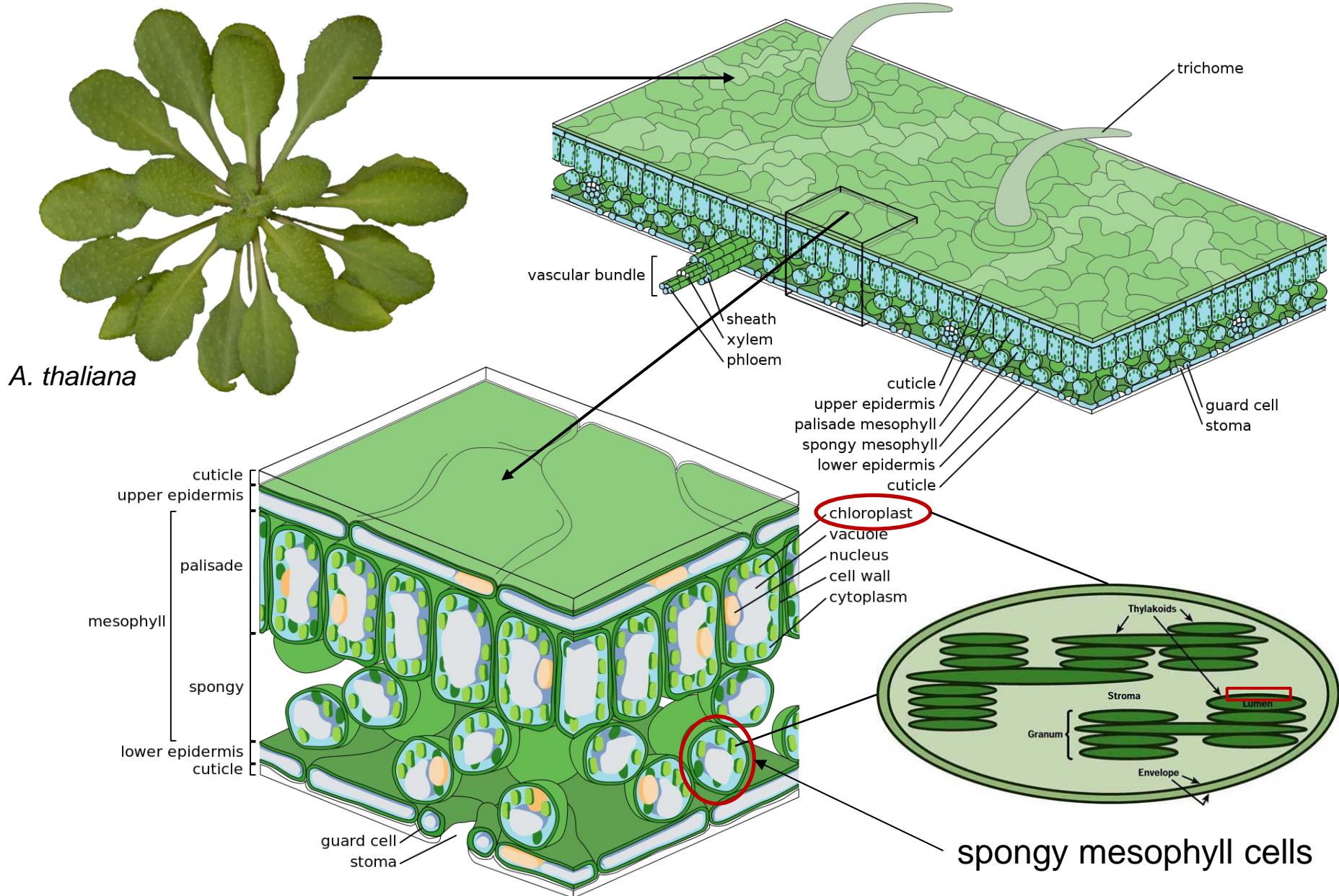
jbc

Biological applications

Use of the novel oxygen primary ion source for the localization of major (**Na, Ca, Mg**) and trace (**Fe, Cu, Mn, Zn**) metals
Involved in physiological processes in plant cells

Insight into photosynthesis

Subcellular localisation of metals in *Arabidopsis thaliana* leaf cells



Essential (trace) metals involved in photosynthesis

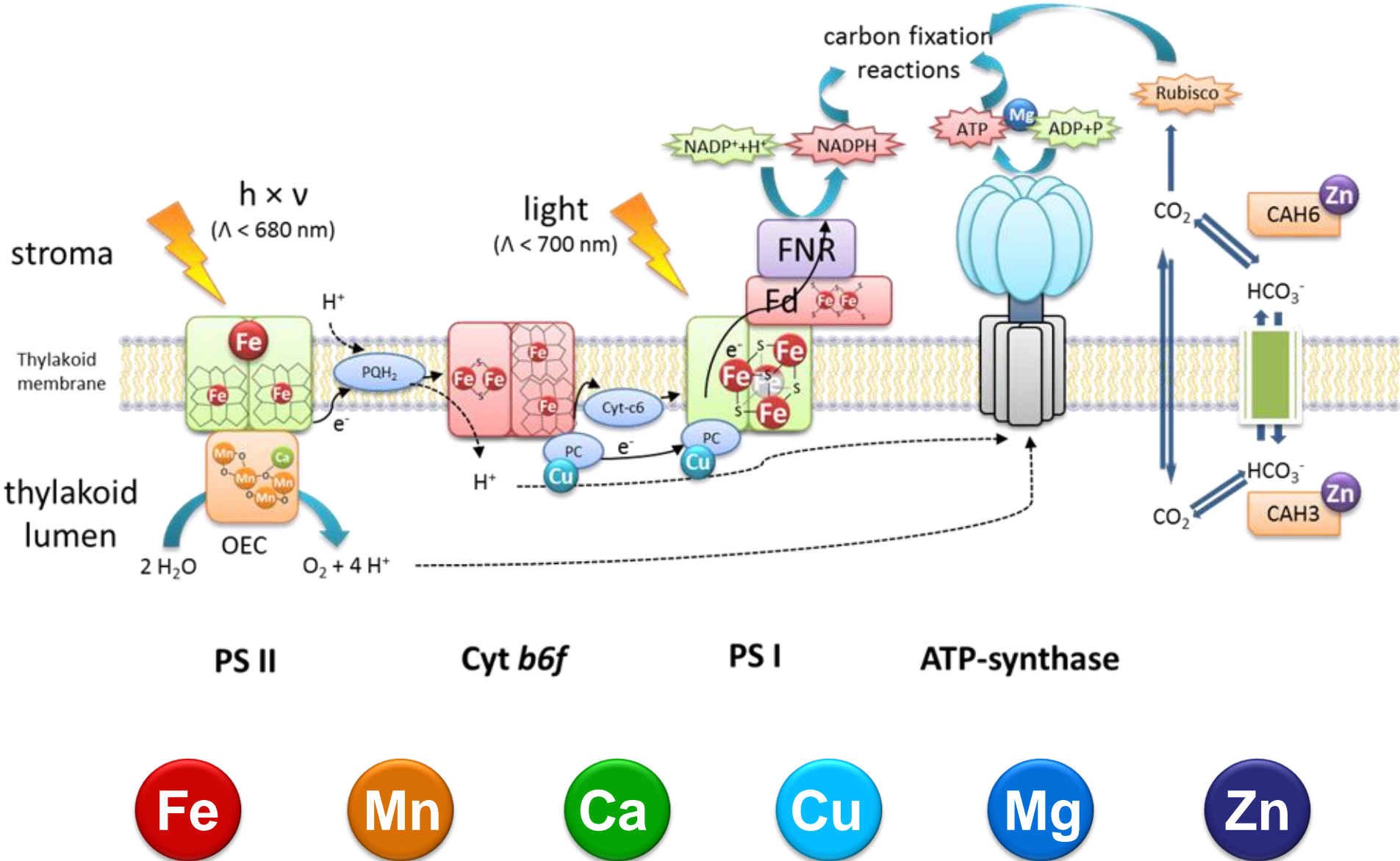
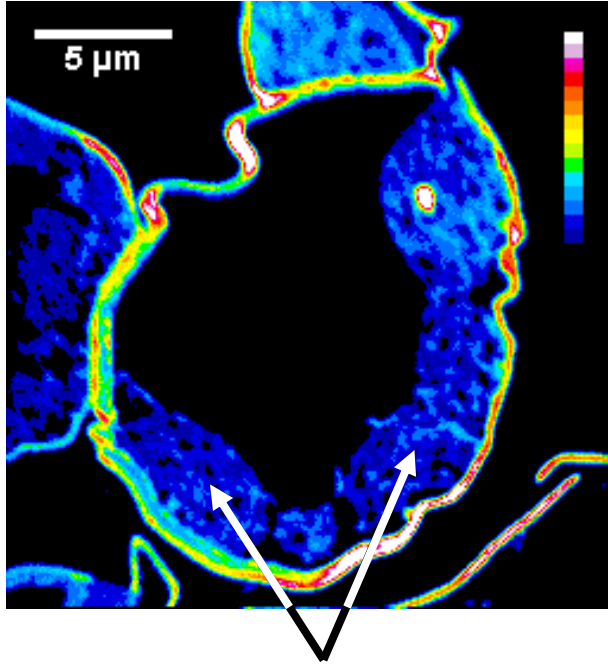


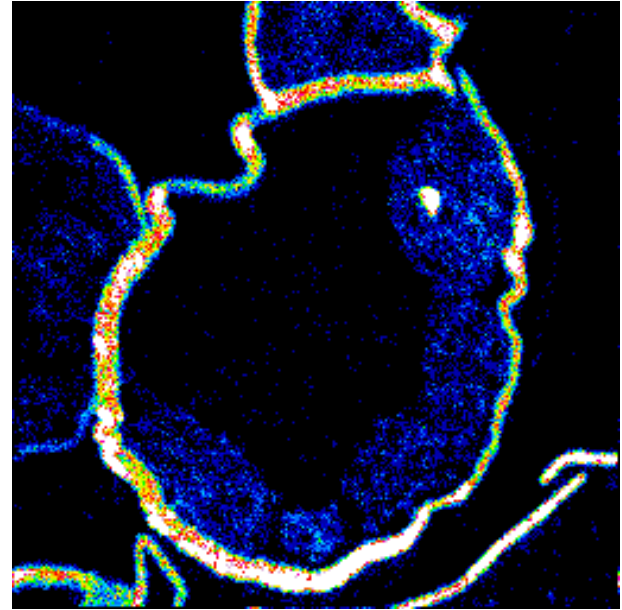
Image of a whole cell: sodium, calcium and magnesium imaging

^{23}Na

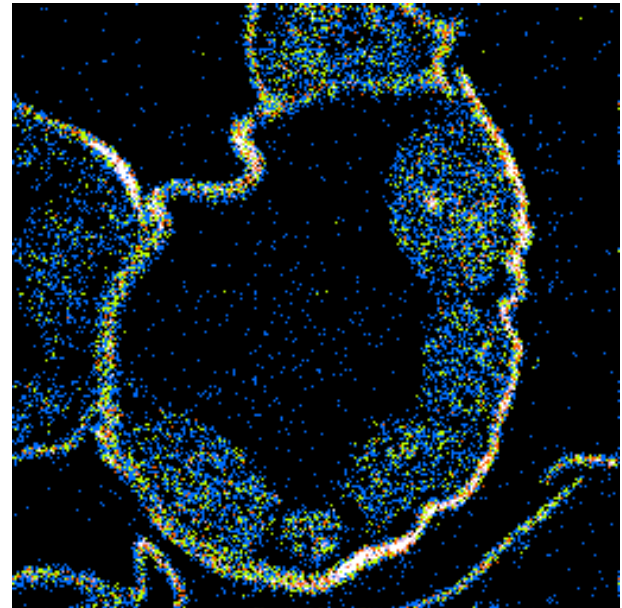


Chloroplasts

^{40}Ca



^{24}Mg



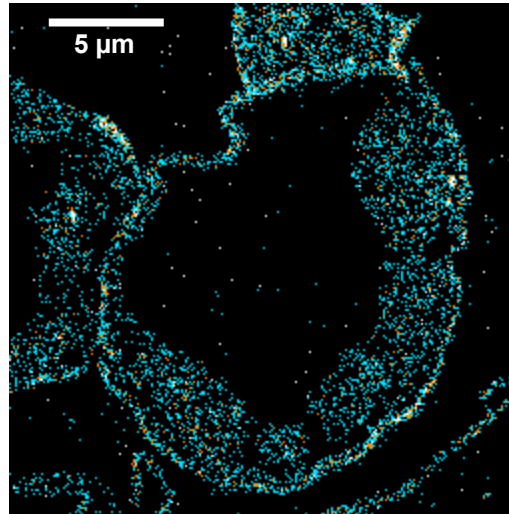
256 x 256 pixel
22 x 22 μm

1 plane

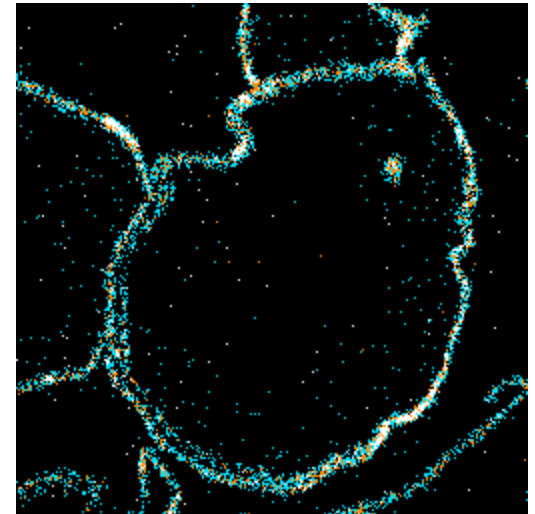
Total acquisition time: 5.5 min

Trace element imaging

^{56}Fe



^{63}Cu

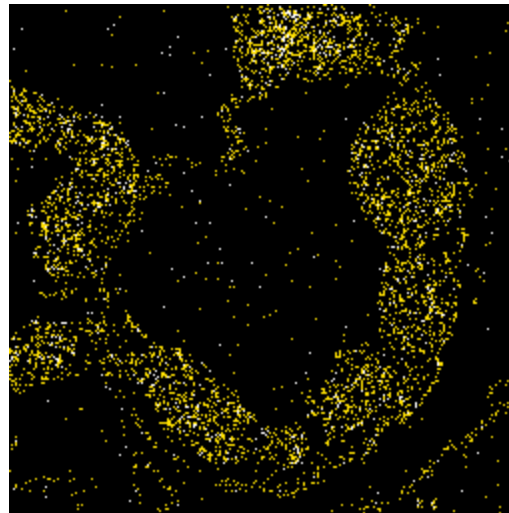


256 x 256 pixel
22 x 22 μm

Overlay of 12 planes
(single images)

Total acquisition
time: **1.5h**
(7.6 min/plane)

^{55}Mn



^{66}Zn

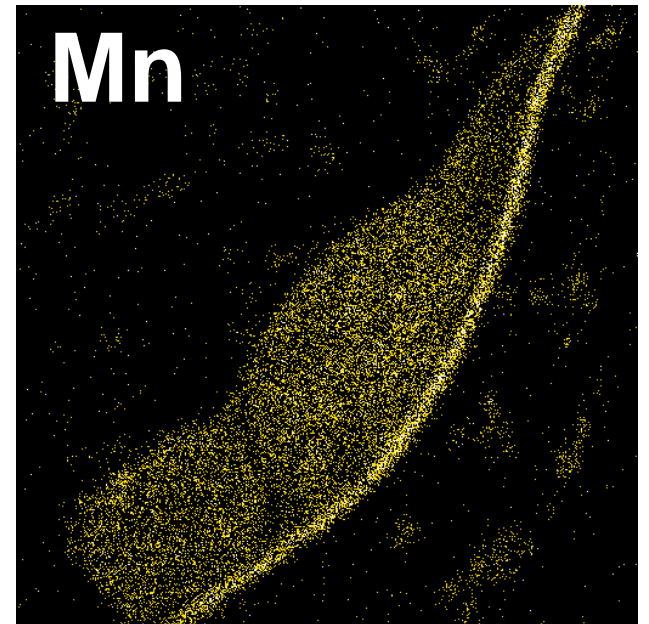
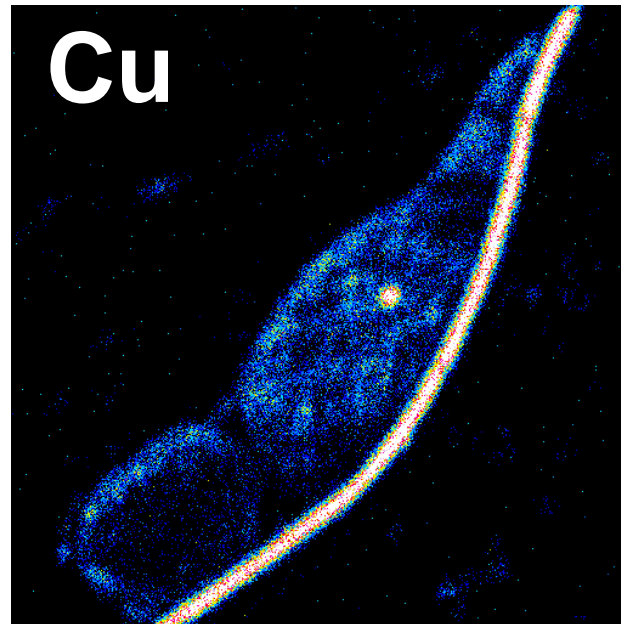
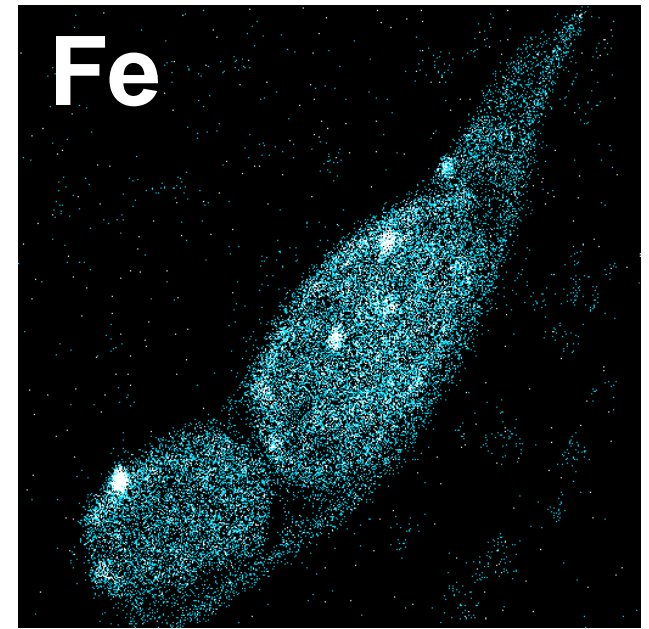
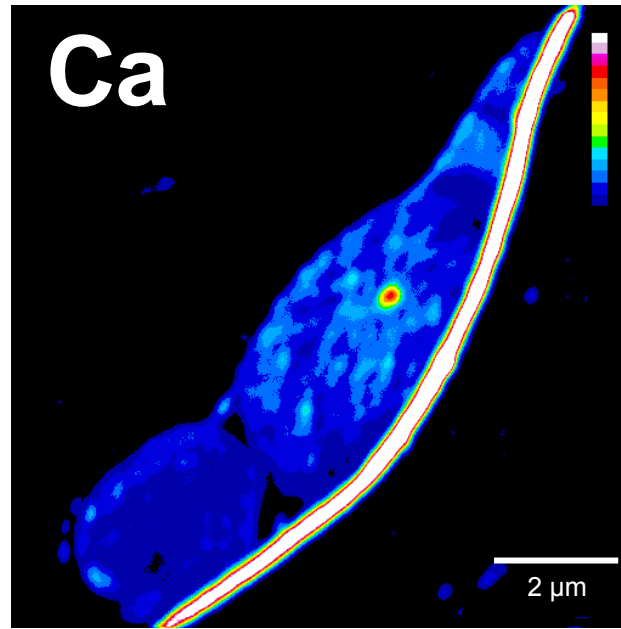


Insight into the chloroplast

512 x 512 pixel
10 x 10 μm

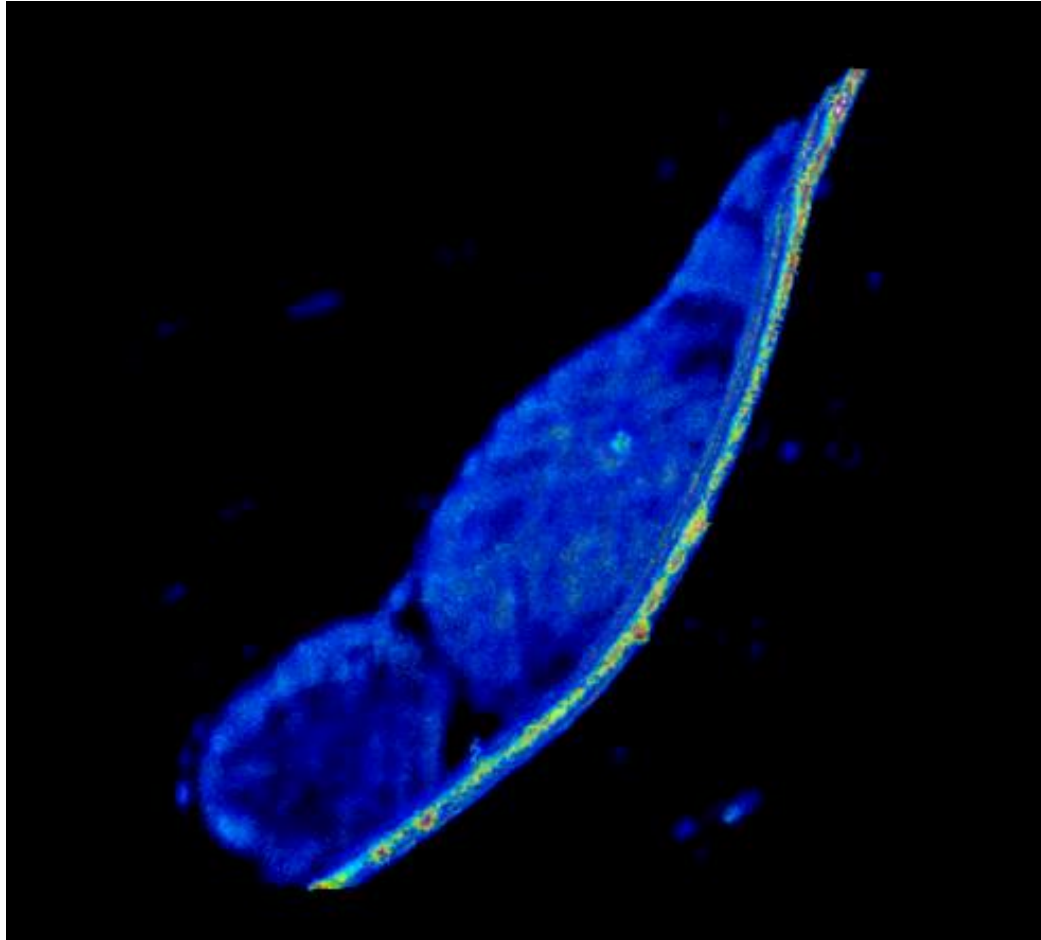
Overlay of 30 planes
(single images)

Total acquisition
time: **11h**
(21.8 min/plane)



3D reconstruction of 30 successive planes :

^{40}Ca



Conclusions

NanoSIMS offers many applications in different scientific fields :

Microbiology, Cell Biology, Environment

but also Material Sciences, Cosmology, Geochemistry :

- Surface analysis by imaging, element quantification, and isotope analysis
- High spatial resolution: 50 x 50 nm in *2D*
50 x 50 x 10 nm in *3D*
- Parallel detection up to 7 masses
- High sensibility
- High mass resolution

A **novel Oxygen primary ion source** shows high stability and high resolution and allows a parallel sensitive detection of biologically relevant major and trace elements at subcellular level.

Acknowledgements

University of Pau/CNRS-IPREM

Julien Malherbe NanoSIMS Engineer
Florent Penen PhD student
Marie-Pierre Isaure Lecturer

University Halle/Saale (Germany)

Dirk Dobritzsch (plant biochemistry)
Julia Frank (plant biochemistry)
Ivo Bertalan (plant biochemistry)
Gerd-J. Krauss (plant biochemistry)
Martin Hertzberg (biochemistry)
Gerd Hause (sample preparation)

Bordeaux Imaging Center

Etienne Gontier (TEM, sample prep.)

University of Bordeaux

Philippe Le Coustumer (TEM/X-EDS)

CAMECA Paris (NanoSIMS)

François Horreard
François Hillion

French ANR-EQUIPEX program (Equipment of Excellence)

Project: **ANR-11-EQPX-0027 – Mass Spectrometry Center MARSS**



**Thank you for
your attention !**

