

Total mercury (THg) concentrations in water, sediment and biota:

General principles, sampling, preparation and methods of analysis

Zorana Kljaković-Gašpić

Institute for Medical Research and Occupational Health (IMROH)

KICK-OFF MEETING

Integrated evaluation of aquatic organism responses to metal exposure: gene expression, bioavailability, toxicity and biomarker responses (BIOTOXMET)

Zagreb, 11th October 2021

Outline

- General aspects of total mercury analysis
- Sampling and storage
- Sample preparation: pre-processing, digestion
- Analytical methods
- Future prospects

General aspects

Determination of THg in environmental samples involves the following steps:

- (a) sample collection;
- (b) sample pretreatment/preservation/storage;
- (c) liberation of mercury from its matrix;
- (d) extraction/clean-up/pre-concentration;
- (e) quantification

Each of these 5 steps requires that we adhere to the **general requirements**:

- High purity reagents (Suprapur or 'low in mercury')
- Deionized water (for rinsing, preparation of standards, dilution of samples, etc..)
- Inert plastic (PTFE) or glass cookware and laboratory ware
- **Rigorous cleaning procedures of laboratory ware and other equipment!!!**
 - necessary to minimize contamination; common to all matrices:
 - (1) aqua regia treatment followed by soaking in diluted (~5-10%) HNO_3 for a week;
 - (2) soaking in a hot oxidizing mixture of KMnO_4 and $\text{K}_2\text{S}_2\text{O}_8$, NH_4OCl rinsing; soaking for a week in 5M HNO_3
 - (3) soaking in a 1:1 mixture of concentrated chromic and nitric acids for a few days;
 - (4) soaking in BrCl (mixture of HCl and KBrO_3);
 - (5) Teflon usually cleaned in hot concentrated HNO_3 for 48 hours, followed by soaking in dilute HNO_3 (5%) etc...
- Storing in a mercury-free place, preferably sealed in mercury-free plastic bags

Total mercury analysis at IMROH

Primary tasks:

measurement of total mercury (THg) in **5 different media**:

- water samples (total and dissolved)
- sediments
- fish muscle
- fish intestine
- intestinal parasites (acanthocephalans)

Water is **the most demanding** of these media → Hg concentrations in natural waters are very low (nanogram-per-liter levels)

→ difficult to get accurate and reliable results!!!

Natural waters are susceptible to contamination from many sources:

- improperly cleaned equipment,
- improper sample-collection techniques,
- contaminated reagents
- atmospheric inputs (dust, dirt, rain)

Clean procedures → necessary to minimize contamination!!!



Contents lists available at ScienceDirect

MethodsX

journal homepage: www.elsevier.com/locate/mex



Method Article

Cleaning and sampling protocol for analysis of mercury and dissolved organic matter in freshwater systems



Andrea G. Bravo^{a,*,1}, Dolly N. Kothawala^{b,1},
Katrin Attermeyer^b, Emmanuel Tessier^c, Pascal Bodmer^{d,e},
David Amouroux^c

^a Department of Environmental Chemistry, Institute of Environmental Assessment and Water Research (IDAEA), Spanish National Research Council (CSIC), Barcelona, Spain

^b Limnology/Department of Ecology and Genetics, Uppsala University, Uppsala, Sweden

^c CNRS/ UNIV PAU & PAYS ADOUR, Institut des Sciences Analytiques et de Physico-Chimie pour l'Environnement et les Matériaux, UMR5254, MIRA, Pau, France

^d Institute for Environmental Sciences, University of Koblenz-Landau, Landau, Germany

^e Chemical Analytics and Biogeochemistry, Leibniz-Institute of Freshwater Ecology and Inland Fisheries, Berlin, Germany

Bravo, A. G., Kothawala, D. N., Attermeyer, K., Tessier, E., Bodmer, P., & Amouroux, D. (2018). Cleaning and sampling protocol for analysis of mercury and dissolved organic matter in freshwater systems. *MethodsX*, 5, 1017–1026. doi:<https://doi.org/10.1016/j.mex.2018.08.002>

Sampling: cleaning procedures

The cleaning procedure for the amber borosilicate bottles and other plastic and glass utensils used to store water and analyze mercury species should be carried out in a **series of three baths** :

- Soaking in mild solution of **detergent** (Kemex; Kemika) for 1 h in ultra-sonic bath or 48 h without sonification
- Rinsing with Milli-Q water
- Soaking in a **10% (v:v) HNO₃** (Merck, p.a., distilled at sub-boiling temperature) bath for 2 h in ultra-sonic bath or 48 h without sonification
- Rinsing with Milli-Q water
- Soaking in a **10% (v:v) HCl** (Merck, p.a., distilled at sub-boiling temperature) bath for 2 h in ultra-sonic bath or 48 h without sonification
- rinsing with Milli-Q water

How to minimize potential sources of contamination during sampling:

- Wear clean, no-talc gloves during all operations
- Use metal free apparatus and materials (PTFE or glass bottles)
- All sampling equipment should be double bagged to reduce risk of contamination
- Avoid airborne particulate matter (dirt, dust, nearby bridges, wires and poles)
- Avoid cigarette smoke
- Avoid vapors from automobile exhaust
- Avoid breathing directly into the sample (dental mercury amalgam fillings)

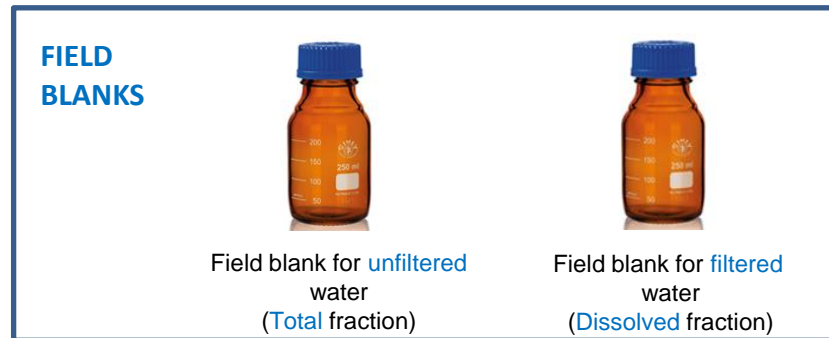
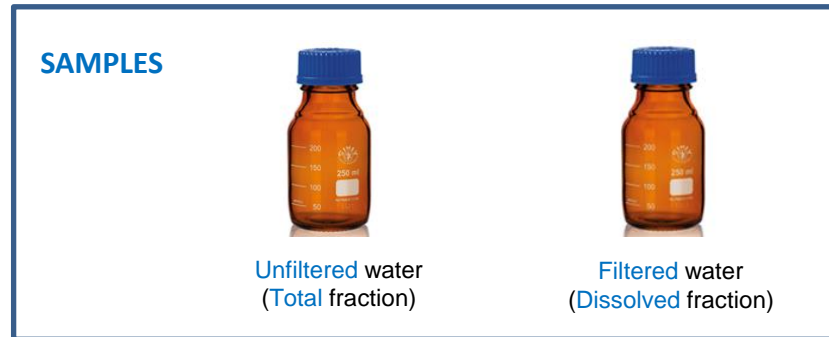


Water sampling

Material required for 1 location:



Summary of the samples collected:



Field blanks → to evaluate the potential for contamination associated with the field methods, materials used, and sampling environment.
- processed in the same manner and under the same environmental conditions as environmental samples

- Water samples were acidified (HCl, Suprapur) and stored at +4°C in clean plastic bags with zipper
- Analyzed as soon as possible, but can be stable for 30 days

Sample preparation: pre-processing

1. **Natural waters:**

Preservation: acidified in the field (HCl, Suprapur)

2. **Biological material:**

dissection conducted during the field trip; stored at -18°C or -80°C (IRB team);

no freeze-drying

3. **Sediments:**

collected in to clean zip bags, labeled and stored at -18°C
in laboratory: freeze-drying and sieving



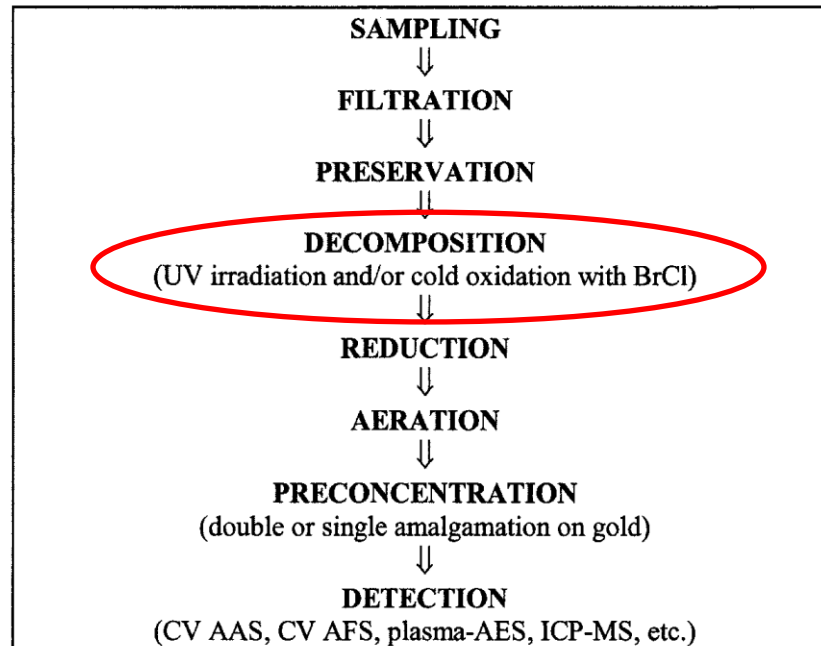
Freeze-drying
HETOSIC (Heto Ltd., Denmark)



Sieved ($\varnothing=2$ mm) to remove
gravel and branches

Sample preparation: digestion of samples

Principal steps for the determination of THg in water samples



Decomposition of samples for the THg analysis:

1. Natural waters:

➤ depends on the instrument used

AMA 254: no decomposition required → decomposition occurs in the instrument itself, as part of the analysis

ICP-MS: decomposition with concentrated HNO_3 (Ultraclave IV, IMROH)

Sample preparation: digestion of samples (2)

2. **Biological material** (fish muscle, intestine, acanthocephalans)

wet decomposition with acids:

- ✓ intestine, acanthocephalans: digestion at IRB facilities due to **small amount** of biological samples (agreement VFM and ZKG)
- ✓ muscle tissue of fish: IMROH (Ultraclave IV System)

3. **Sediments:**

Hg in sediments associated with humic matter and/or sulphides (HgS)

→ **strong oxidizing agents** required!! (HNO_3 , HCl, BrCl, H_2SO_4 , HClO_4 , H_2O_2 , V_2O_5 , KMnO_4 , $\text{K}_2\text{Cr}_2\text{O}_7$)

wet decomposition with Aqua regia (HNO_3 :HCl = 1:3) in Al-block on 70-90°C (IMROH)

Sample preparation: digestion of biological material

High pressure microwave destruction of samples (fresh or freeze-dried) in a high-pressure microwave device using method for biological samples



UltraCLAVE IV (Milestone, Italy)

- Samples of fresh biological material (0.250-0.500 g) are weighed in **quartz tubes**
- 4 ml of concentrated HNO_3 (65%; S.p.) is added and left to react for 10 to 15 minutes; then 2 ml of deionized water is added
- Tubes are closed with a Teflon stoppers and placed on a stand which is immersed in a stock solution (water, H_2O_2 , H_2SO_4) which serves to conduct the microwaves more evenly
- **Quality Control:** Blanks and Certified Reference Materials digested in the same manner with every batch of samples!

Temperature program for destruction of biological samples in high-pressure microwave device UltraCLAVE IV (Milestone, Italy)

	t (min:s)	E (W)	T_1 (°C)	T_2 (°C)	p (bar)
1.	3:30	700	70	60	100
2.	15:00	1000	180	60	100
3.	10:00	1000	220	60	120
4.	30:00	1000	220	60	120
5.	40:00	0	30	25	20

• $T_{\text{max}} = 220^\circ\text{C}$

• $P_{\text{max}} = 120$ bar

• Number of samples_{max} = 40

• Required time: ~2.5 h

t - time, T_1 - set temperature, T_2 - temperature in the outer vessel,
E - microwave radiation power, p - pressure in the reaction vessels

Sample preparation: digestion of sediments



TECATOR:

Aluminium block with 40 slots
Autostep 1012 Controller

Wet digestion in Al-block:

- Sediment samples (0.100–0.150 g) are weighed into [quartz tubes](#);
- 3 ml of Aqua-regia ($\text{HNO}_3:\text{HCl} = 1:3$) is added and mixture is allowed to react overnight.
- After addition of 1 ml of deionized water, the tubes are closed with a Teflon-coated stoppers, placed in the Aluminum block and heated for 5 hours at 90°C.
- After cooling, the samples are made up with Ultrapure water to a total volume of 20 mL and stored at room temperature.
- **Quality Control:** Blanks and Certified Reference Materials digested in the same manner with every batch of samples!

Sample analysis: Instrumentation

Common analytical techniques for the analysis of Total mercury:

- Atomic Absorption Spectrometry (AAS): GF-AAS, CV-AAS
 - Cold Vapor Atomic Absorption Spectrometry (CV-AAS)
 - **Total mercury analyzer** (Direct thermal decomposition – Gold Amalgamation – Cold Vapor Atomic Absorption Spectrometry)
- Atomic Fluorescence Spectrometry (AFS): CV-AFS
- Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)
- Neutron Activation Analysis (NAA)
- Gas Chromatography (GC): GC-ECD (GC with Electron Capture Detector)
-

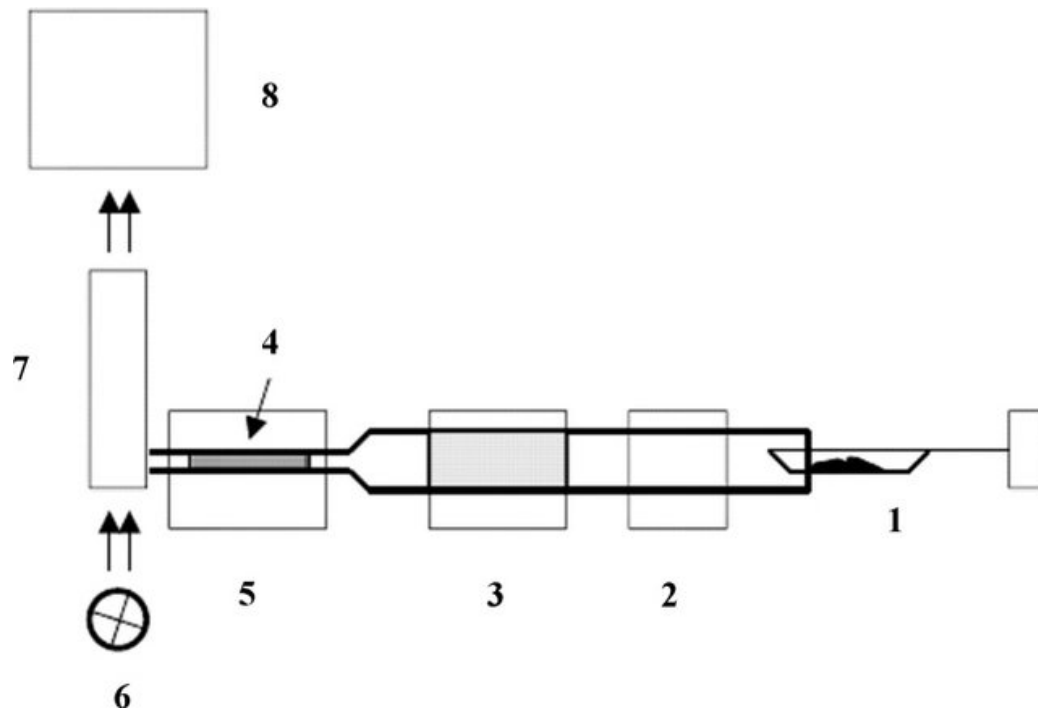
Sample analysis: AMA 254 Total Mercury Analyzer

- Principle: Direct thermal decomposition – Gold Amalgamation – CVAAS



AMA 254 Mercury Analyzer (LECO, Korea)

- **Application:** mercury content analysis in solid, liquid and gaseous samples (direct measurements are possible)
- LOD = **0.003 ng Hg**



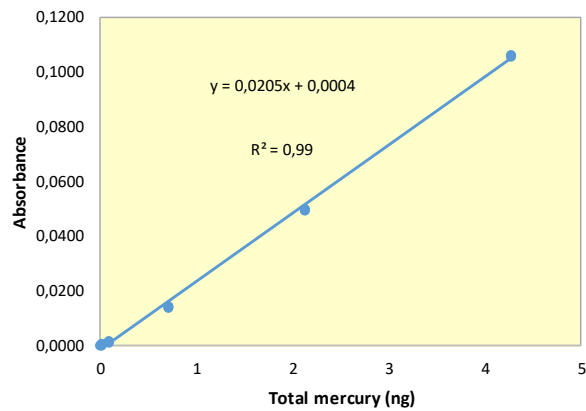
Scheme of the AMA 254 spectrometer:

(1) Sampling boat, (2) decomposition furnace, (3) catalytic column, (4) gold amalgamator, (5) releasing furnace, (6) mercury cathode lamp, (7) optical cell system, and (8) detector

(Source: Spěváčková et al. (2004). *Analytical and Bioanalytical Chemistry*, 380(2), 346–350. doi:10.1007/s00216-004-2739-2)

AMA254 Total Mercury Analyzer: Results

Datum: 17.08.2021.		Napomena: vrijednosti THg u svim uzorcima (filtriranim i nefiltriranim) su na granici detekcije metode									
Uzorcji: Filtrirane i nefiltrirane prirodne i otpadne vode s Krke oko Knina											
Analitičar: Zorana Kljaković-Gašpić											
Instrument: AMA-254 Mercury analyser (Leco Inc.)											
	Standard concentration (µg/L)	ng Hg	Absorban	Average blank absorban ce							
Blank	0	0,0029	0,0002								
	0	0,0099	0,0002	0,0002							
STD 1	0,2134	0,02134	0,0006								
	0,2134	0,02134	0,0005								
STD 2	0,8536	0,08536	0,0014								
	0,8536	0,08536	0,0016								
STD 3	7,1133	0,7113	0,0142								
	7,1133	0,7113	0,0144								
STD 4	21,34	2,134	0,0497								
	21,34	2,134	0,0500								
STD 5	21,34*200µL	4,268	0,1060								
	21,34*200µL	4,268	0,1062								
Sample			Aliquot (mL)	Absorban ce	Absorban ce (Sample-Blank)	Total Hg (ng)	Blank substituted concentration (ug/L)	AVERAGE concentra tion (ug/L)			
NIST 1641e		r=10	0,10	0,0214	0,02120	1,0291	102,9		Certified val	Range	
			0,10	0,0213	0,02110	1,0293	102,9	102,9	101,6 µg/L	99,9-103,3	
BLF_KRS	Krka river source	Filtrirano	0,5	0,0004	0,0002	0,0098	0,020				
BLF-KRK	Krka River Knin	Filtrirano	0,5	0,0006	0,00040	0,0195	0,039				
KRS_F	Krka river source	Filtrirano	0,5	0,0005	0,00030	0,0146	0,029				
KRK_F	Krka River Knin	Filtrirano	0,5	0,0004	0,00020	0,0098	0,020				
KBL_F	Krka Brljan Lake	Filtrirano	0,1	0,0004	0,00020	0,0098	0,098				
TOR_F	tributary Orašnica	Filtrirano	0,1	0,0005	0,00030	0,0146	0,146				
TBU_F	tributary Butišnica	Filtrirano	0,1	0,0003	0,00010	0,0049	0,049				
TKO_F	tributary Kosovčica	Filtrirano	0,1	0,0002	0,00000	0,0000	0,000				
IWW_F	industrial wastewaters	Filtrirano	0,1	0,0004	0,00020	0,0098	0,098				
BL-KRS	Krka river source	Nefiltrirano	0,5	0,0003	0,00010	0,0049	0,010				
BL-KRK	Krka River Knin	Nefiltrirano	0,5	0,0004	0,00020	0,0098	0,020				
KRS	Krka river source	Nefiltrirano	0,5	0,0005	0,00030	0,0146	0,029				
KRK	Krka River Knin	Nefiltrirano	0,5	0,0005	0,00030	0,0146	0,029				
KBL	Krka Brljan Lake	Nefiltrirano	0,1	0,0003	0,00010	0,0049	0,049				
TOR	tributary Orašnica	Nefiltrirano	0,5	0,0002	0,00000	0,0000	0,000				
TBU	tributary Butišnica	Nefiltrirano	0,5	0,0003	0,00010	0,0049	0,010				
TKO	tributary Kosovčica	Nefiltrirano	0,5	0,0002	0,00000	0,0000	0,000				
IWW	industrial wastewaters	Nefiltrirano	0,1	0,0002	0,00000	0,0000	0,000				



Sample analysis: ICP-MS

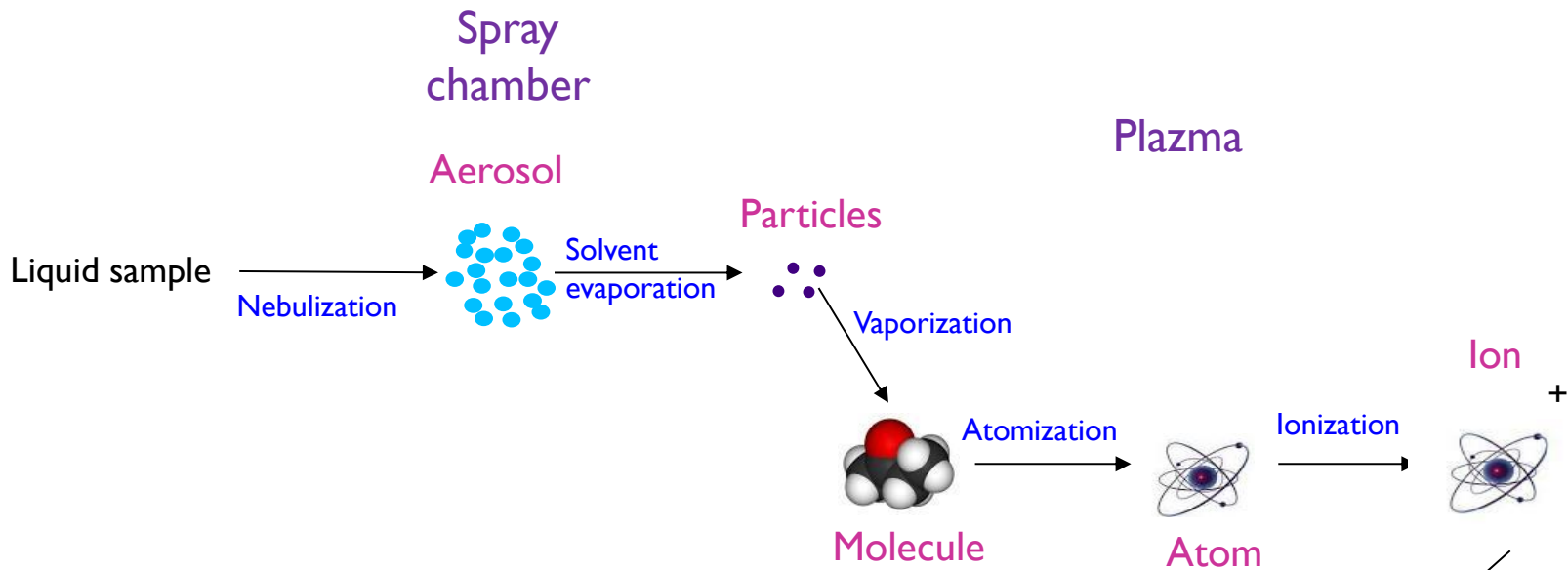
- **Water samples:** an internal standard (IS) solution is added to a given sample volume and the samples are measured directly, without dilution
- **Biological samples** are diluted with 1% (v/v) HNO_3 5-20 times, depending on the matrix
- **Sediment samples** are diluted with 1% (v/v) HNO_3 ~ 20 times
- Internal standard of 2 $\mu\text{g/L}$ is added to all samples



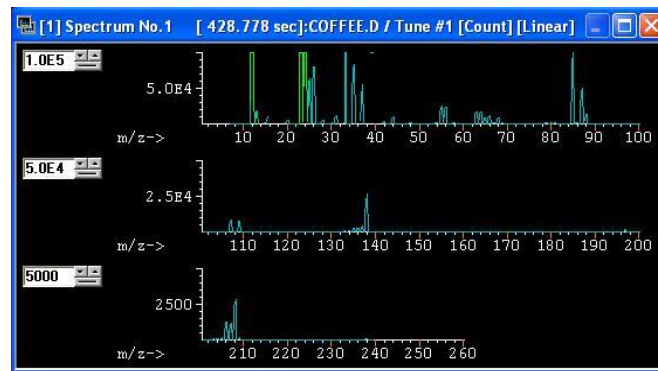
Agilent 7500cx, Agilent Technologies

- High sensitivity
- Low detection limits → **LOD** for Hg \approx **0.005 $\mu\text{g/L}$** or lower
- Can analyze many other metals at the same time (about fifty elements in one run)
- Allows for verification by analysis of multiple isotopes
- Linearity of detector response in a large concentration range (through nine orders of magnitude)
- Internal standardization improves accuracy of results
- Requires small sample volume for analysis (\sim 2-3 mL of sample solution)

Sample analysis: Processes in ICP-MS



Mass spectrum

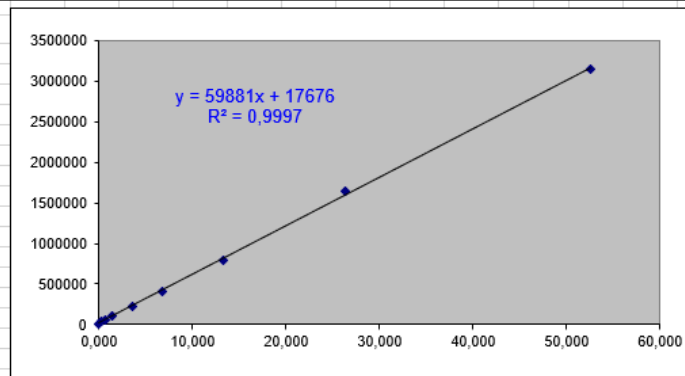


Mass spectrometer

Schematic representation of the process in ICP-MS from sample input to mass analysis

ICP-MS: Results (2)

AI		vodeni								
		vodeni		X/Sc	X/72Ge	X/74	X/Rh	X/Tb	X/Lu	X/lr
		0,000	11984	0,3	1,2	0,9	0,2	0,1	0,1	0,3
7	STD1	0,304	39916	0,8	3,4	2,7	0,6	0,4	0,4	1,0
8	STD2	0,679	58788	1,2	5,0	3,9	0,9	0,6	0,6	1,4
9	STD3	1,384	103476	2,1	8,8	6,8	1,5	1,1	1,1	2,4
10	STD4	3,551	230953	4,8	20,2	15,3	3,4	2,4	2,5	5,5
11	STD5	6,723	409939	8,4	35,5	26,7	6,0	4,3	4,4	9,9
12	STD6	13,351	795802	16,2	68,1	52,1	11,7	8,2	8,4	18,8
13	STD7	26,349	1641027	33,8	139,2	104,1	24,2	17,1	17,3	39,0
14	STD8	52,520	3146388	65,2	259,8	192,3	45,6	32,0	32,6	73,4
15	bez std 7 i 8	58549		1,19	4,98	3,80	0,85	0,60	0,62	1,38
16	Nagib=	59881		1,24	4,96	3,67	0,87	0,61	0,62	1,40
17	Odsječak=	17676		0,43	1,82	1,36	0,29	0,21	0,21	0,49
18	RSQ=	0,99967		1,000	0,999	0,999	0,999	0,999	0,999	0,999
20	Blank 1%HNO3	11984,2	12766	12553	10634					
21	BlankMQH2O	12331,5		12721	11942					
22			5826							



	COUNTS /CPS (Mean)	vodeni																	V1 uk/ml	V2 uk/ml	C1 (bazz1)	C2 / Sc	C2 / 72Ge	C2 / 74Ge	C2 / Rh	C2 / Tb	C2 / Lu	C2 / 193lr			
		RSD	45Sc	72Ge	74Ge	103Rh	159Tb	175Lu	193lr	X/Sc	X/72Ge	X/74Ge	X/Rh	X/Tb	X/Lu	X/lr															
27	Bi_3_P	10634	10022	11277	10603	5,9	43894	10059	13193	62101	91254	89327	39691	0,2	1,1	0,8	0,2	0,1	0,1	0,3	1	1	-0,03	-0,04	-0,03	-0,03	-0,03	-0,03	-0,03	-0,03	
28	Bi_1_MQH2O	5826	5793	5753	5931	1,6	41769	10423	13815	61602	89997	88633	40236	0,1	0,6	0,4	0,1	0,1	0,1	0,1	1	1	-0,11	-0,12	-0,13	-0,13	-0,12	-0,12	-0,12	-0,12	
29	Bi_2_MQH2O	12721	12837	12605	12721	0,9	42668	10284	13606	62272	90606	88983	39914	0,3	1,2	0,9	0,2	0,1	0,1	0,3	1	1	0,01	0,01	0,01	0,01	0,01	0,01	0,01	0,01	
30	Bi_3_MQH2O	11942	11990	11900	11935	0,4	42671	10275	13478	62266	89933	88997	39793	0,3	1,2	0,9	0,2	0,1	0,1	0,3	1	1	-0,01	-0,01	-0,01	-0,01	-0,01	-0,01	-0,01	-0,01	
31	Kal_1_trav	60520	61055		59985	1,3	70899	10799	13983	61142	93405	93083	40398	0,9	5,6	4,3	1,0	0,6	0,7	1,5	1	1	0,80	0,46	0,89	0,93	0,91	0,84	0,82	0,85	
32	Kal_stap_trav	52960	53612	51617	53653	2,2	72404	11140	14177	62370	94790	94612	41224	0,7	4,8	3,7	0,8	0,6	0,6	1,3	1	1	0,68	0,36	0,72	0,77	0,75	0,69	0,68	0,70	
33	Kor_1_trav	81690	81249	87434	76387	6,8	72133	11330	14475	63507	96557	95201	41995	1,1	7,2	5,6	1,3	0,8	0,9	1,9	1	1	1,16	0,68	1,21	1,29	1,25	1,16	1,16	1,17	
34	Kor_stap_trav	75125	78557	72583	74236	4,1	72816	11259	14580	63508	96092	94513	41857	1,0	6,7	5,2	1,2	0,8	0,8	1,8	1	1	1,05	0,60	1,10	1,16	1,13	1,05	1,05	1,06	
35	Rij_1_trav	252244	255504	249461	251766	1,2	73641	11543	14797	63313	96043	95136	42051	3,4	21,9	17,0	4,0	2,6	2,7	6,0	1	1	4,01	2,53	4,17	4,39	4,35	4,07	4,04	4,06	
36	Rij_stap_trav	255094	253285	252310	259688	1,6	74015	11548	14666	63716	96800	95917	41885	3,4	22,1	17,4	4,0	2,6	2,7	6,1	1	1	4,05	2,55	4,21	4,49	4,38	4,09	4,05	4,13	
37	Pro_1_trav	182613	179489	184966	183383	1,5	71097	11641	15038	64760	97532	96577	41832	2,6	15,7	12,1	2,8	1,9	1,9	4,4	1	1	2,84	1,84	2,92	3,06	3,01	2,84	2,82	2,90	
38	Pro_stap_trav	104492	102081	106902		3,3	71873	11644	15154	66226	98400	98009	42622	1,5	9,0	6,9	1,6	1,1	1,1	2,5	1	1	1,54	0,94	1,57	1,63	1,59	1,51	1,49	1,53	
39	Mat_1_trav	439508	444929	434946	438648	1,1	87621	11796	15325	64991	97586	97138	42551	5,0	37,3	28,7	6,8	4,5	4,5	10,3	1	1	7,13	3,81	7,27	7,56	7,55	7,14	7,05	7,15	
40	Mat_stap_trav	376278	382190	372558	374085	1,4	88020	11416	14725	64574	96903	96342	41830	4,3	33,0	25,6	5,8	3,9	3,9	9,0	1	1	6,08	3,21	6,41	6,71	6,47	6,13	6,06	6,20	
41	Pit_1_trav	336152	327380	343733	337342	2,5	74554	11903	15099	65071	97508	96771	42639	4,5	28,2	22,3	5,2	3,4	3,5	7,9	1	1	5,41	3,40	5,45	5,81	5,71	5,41	5,36	5,41	
42	Pit_stap_trav	352371	359434	346848	350829	1,8	73801	11675	14778	64660	97660	96527	41892	4,8	30,2	23,8	5,4	3,6	3,7	8,4	1	1	5,68	3,62	5,85	6,24	6,04	5,68	5,65	5,78	
43	Koz_1_trav	70838	70103		71572	1,5	74404	11556	15065	64698	97044	96799	42123	1,0	6,1	4,7	1,1	0,7	0,7	1,7	1	1	0,98	0,53	0,99	1,03	1,03	0,97	0,95	0,98	
44	Koz_stap_trav	74946	74315	75670	74853	0,9	75205	11651	15027	64980	96787	96240	42191	1,0	6,4	5,0	1,2	0,8	0,8	1,8	1	1	1,05	0,57	1,06	1,11	1,10	1,04	1,03	1,05	
45	sum_a_1_trav	14312527	15734630	12438274	14764676	11,8	57481	11573	14953	64179	95959	95244	41332	249,0	1236,7	957,2	223,0	149,2	150,3	346,3	1	1	238,8	200,6	249,2	260,4	256,2	243,6	241,3	247,0	
46	sum_a_2_trav	12317054	12900168	12245353	11805641	4,5	55791	11631	14639	63684	94496	93827	41208	220,8	1059,0	841,4	193,4	130,3	131,3	298,9	1	1	205,5	177,8	213,4	228,9	222,2	212,9	210,8	213,2	
47	sum_b_1_trav	35064986	35899016	32001822	37294210	7,8	62175	12235	14544	62618	94584	91853	40605	564,0	2866,1	2410,9	560,0	370,7	381,8	863,6	1	1	585,4	454,6	577,8	656,2	643,7	605,9	613,4	616,3	
48	sum_b_2_trav	28433147	29190760	28148236	20960446	4,8	59540	11650	14384	62111	94894	91701	40679	494,3	2526,6	2046,2	473,9	310,2	321,0	723,5	1	1	491,3	398,4	509,3	556,9	544,7	506,9	515,7	516,4	
49	Dummy (1%HNO3)	27705	27298	28873	26944	3,7	4843	651	83	33	12	22	39	5,7	42,5	332,4	831,1	2266,7	1246,7	712,4	1	1	0,3	4,4	8,3	90,3	955,5	3705,7	2003,8	508,4	
50	SLRS-5_I	49,5 (44,5-54,5)	2909327	2894185	2927770	2906028	0,6	#####	11434	14954	64609	96729	95004	41186	27,7	254,5	194,6	45,0	30,1	30,6	70,6	1	1	48,38	22,09	51,08	52,73	51,55	48,95	49,00	50,21
51	NIST_1	141,8 (133,2-150,4)	843130	840113	845207	844070	0,3	46492	10858	14590	65091	94749	93287	41303	18,1	77,7	57,8	13,0	8,9	9,0	20,4	0,4	4	138,74	143,91	154,20	154,88	146,67	143,25	143,06	143,54
52	seroporm.urin	100 (94,1-106)	181677	186188	188370	190473	2,1	57674	11708	15054	65173	97585	96026	42088	3,3	16,4	12,7	2,9	2,0	2,0	4,6	0,1	1	148,75	122,35	153,00	160,66	157,74	148,40	149,30	154,53

What was already performed and future prospects

What we have done so far:

- Concentrations of THg in water samples (January, April and June of 2021)

To be achieved:

1st Project period:

- Analysis of THg in water samples (field trip in October 2021)
- Analysis of THg in sediments (will be finished by the end of the 1st Project period) → samples are already digested

2nd Project period

- Digestion and analysis of THg in biological material (fish muscle, intestine and acanthocephalans)

My wishes that are not included in the project tasks:

? Development of the quick method for the analysis of **MeHg** in biological samples (at least in muscle tissue) with AMA 254

? Application of this method for the analysis of **MeHg** content in biological samples

