



UMS 3420



université
de **BORDEAUX**

Inserm

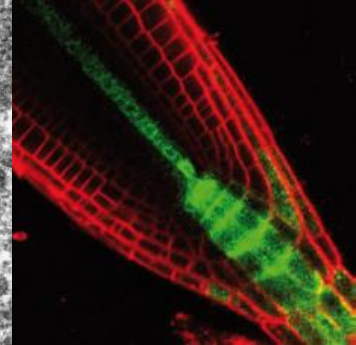
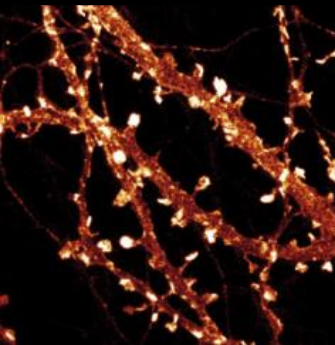
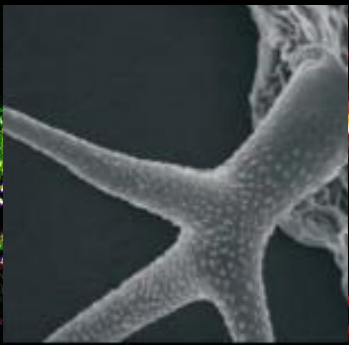
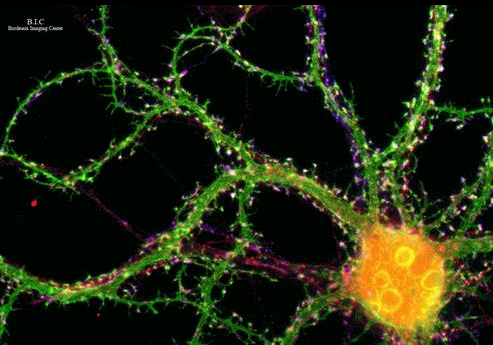
Institut national
de la santé et de la recherche médicale

US 4



bic

Bordeaux Imaging Center

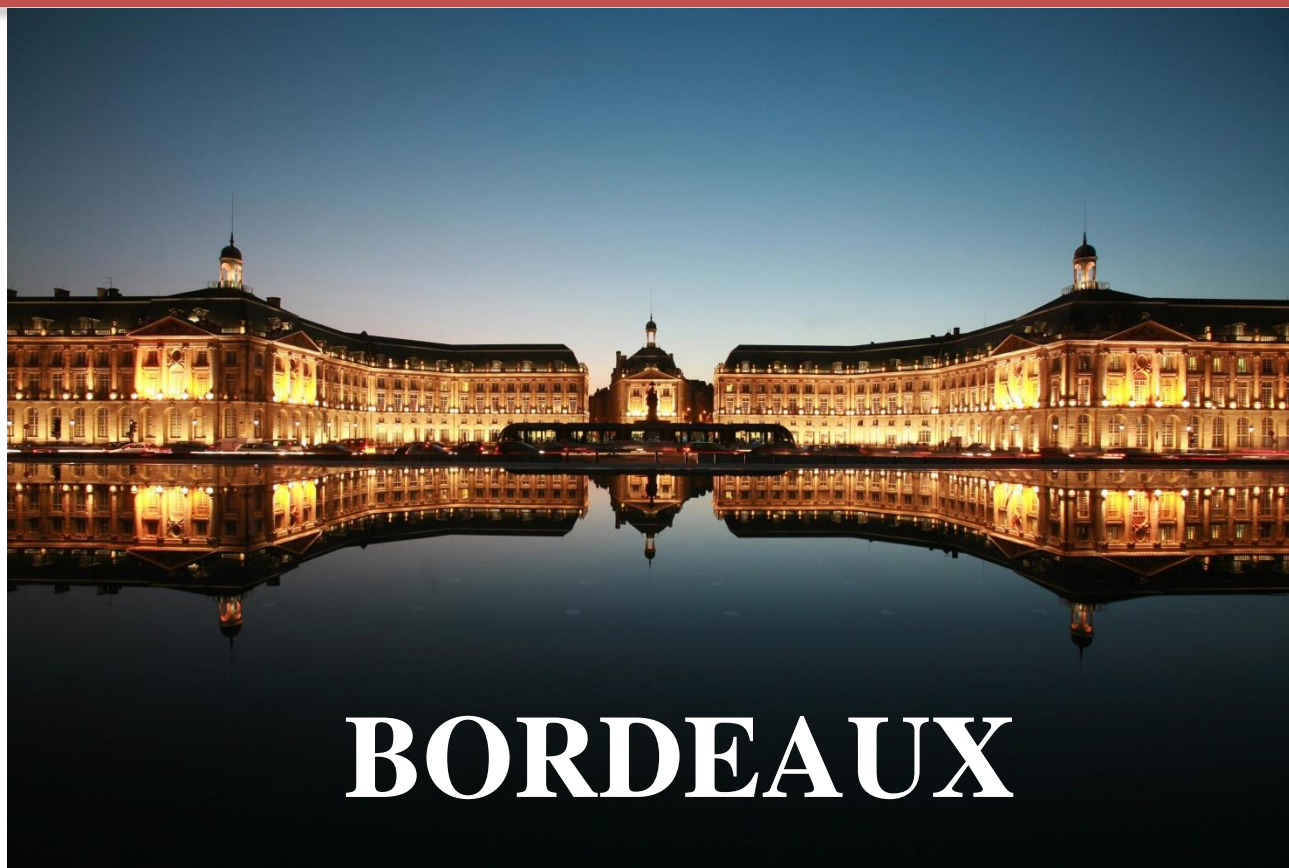


What is the BIC ?

Cell Imaging core facility of Aquitaine

- ⇒ *Gather resources in light and electron microscopy.*
- ⇒ *In the health and plant domain.*
- ⇒ *Infrastructure in health biology and agronomy (IBISA) label*

Location



Location of Bordeaux Imaging Center

Campus
of Bordeaux University

Electronic Imaging Unit

Hospital



Faculty of Sciences

What is the BIC ?

Our missions :

- **Service packages**
 - *Devices providing*
 - *Sample preparation*

- **Training**

- **R&D**

BIC – Devices

14 Microscopes:

1 STED microscopes,

3 épifluo., 1 video-FRAP, 3 confocal, 3 multiphotons, 1 macroscope, 1 « spinning disk » confocal, 1 speed multiphoton (dev)

Laser et autres

- 3 femtosecondes laser
- FLIM, FCS, patch-clamp

Image analyse

- 4 Images analyse station

4 Electron Microscopes :

- TEM FEI – Spirit 120kV (tomography & cryo-observation)
- TEM FEI Tecnai 12 120kV (EDS & tomography)
- TEM Hitachi H7650 120kV
- SEM FEI Quanta 200 (with environmental mode & EDS)

Sample prep devices :

- 5 ultramicrotomes
- 1 microwaves automate for sample preparation Leica EM-AMW
- 2 cryo-congélation hyperbare Leica EM-Pact and 1 Leica EM-HPM100
- 3 Leica EM-AFS (automated freeze substitution)
- 1 Immunolabelling automate Leica EM-IGL
- 1 sputter coater Cressington 108
- 1 critical point dryer Leica EM-CPD

Future Devices in 2016

MEB FEG



High resolution
Cryo transfer system
Serial block face
X Analysis System
Coordinate tracking system (CLEM)

MET 200kV Lab6



High resolution
Cryo transfer system
Electron Tomography
X Analysis System
STEM module

Bordeaux Imaging Center Structure

Director Daniel Choquet

Co-Director : Marc Landry & Patrick Moreau

3 Units
15 Engineers

Electronic

Etienne Gontier

Melina Petrel
Sabrina Lacomme
Isabelle Svahn
Hugo LeGuengo

Photonic

Christel Poujol

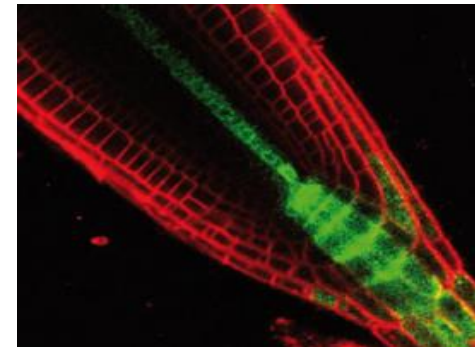
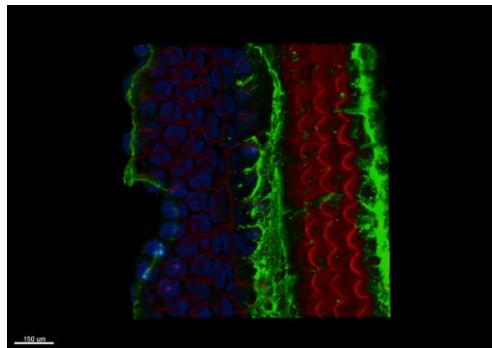
Sebastien Marais
Fabrice Cordelières
Matthieu Ducros

Plant

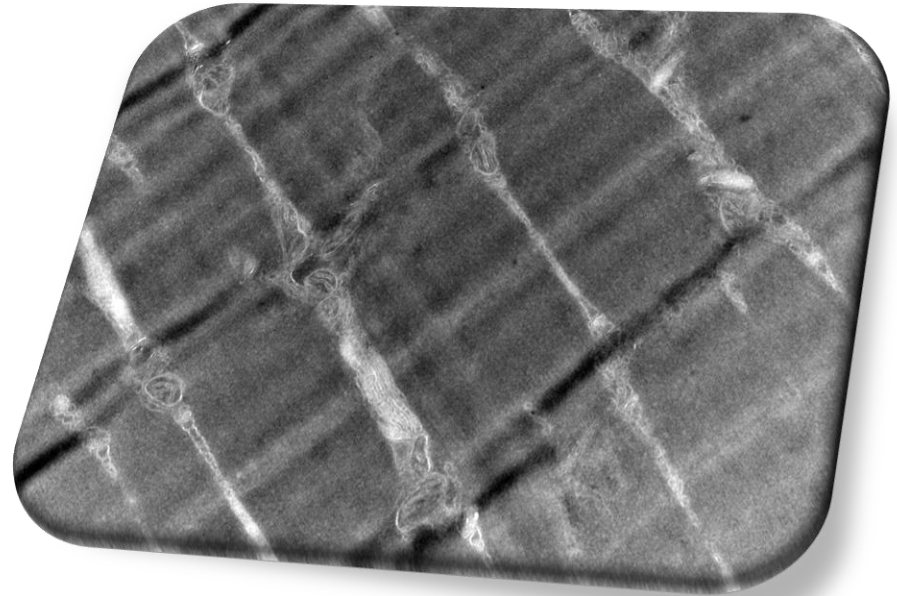
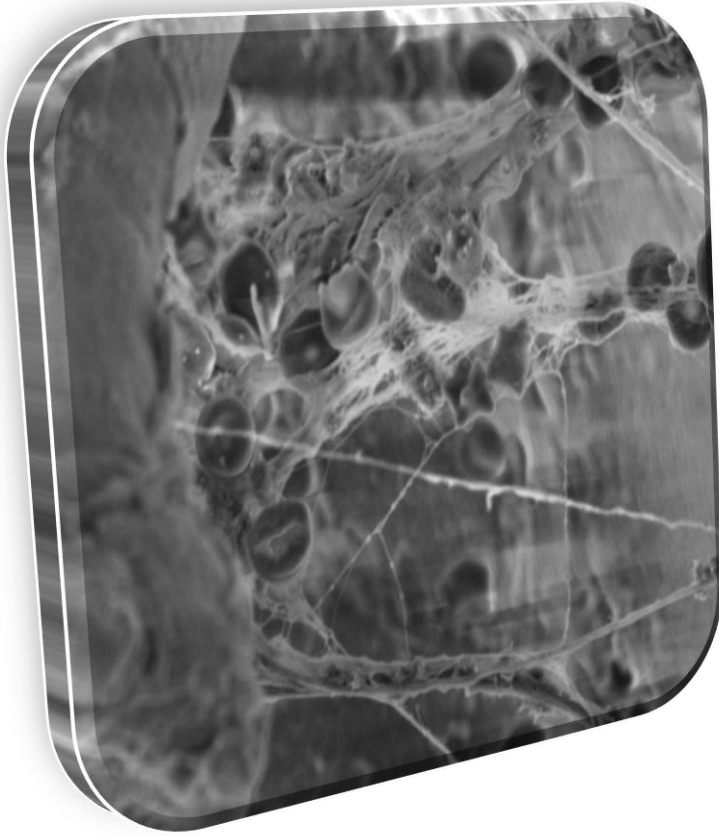
Lysiane Brocard

Catherine Cheniclet
Brigitte Batailler
Valérie Rouyère

Jennifer Petersen: R&D



**Electronic
imaging unit
of the
Bordeaux
Imaging Center**



Etienne.gontier@u-bordeaux.fr

Electronic Imaging Team

Marc Landry



Scientific Manager.

Etienne Gontier



Technical Manager.

In charge of the TEM activity



Sabrina Lacomme

In charge of the cryo-preparation / immunostaining.



Melina Petrel

In charge of the SEM activity.



Isabelle Svahn

Technician in sample preparation and participates in the development



Hugo Le Guenno (CDD)

Business Areas

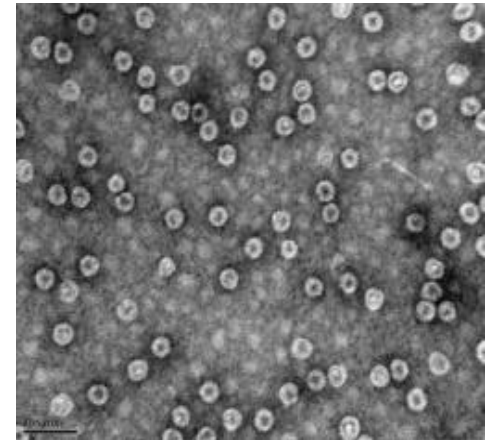
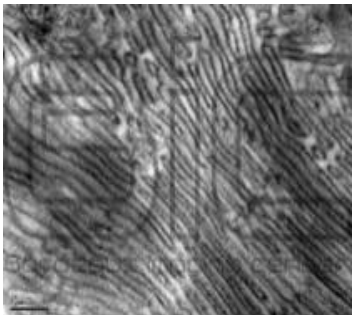
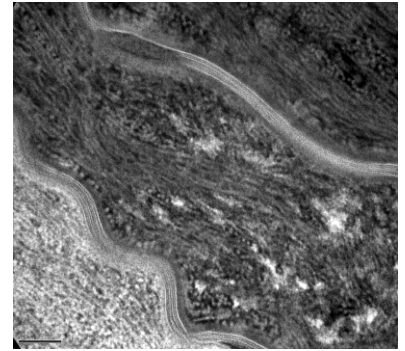
⇒ We work on transmission electron microscopy and scanning

⇒ Major Axis : Biological (Animal / Plant) :

- * Whole tissue
- * Isolated cells or in culture
- * cell suspensions, organelles or membrane fractions
- * micro-organismes (Virus, Levures...).

But also we open the activity to

- * Materials
- * Bio-materials
- * Polymers



The electronic imaging division proposes activities according to three main axis:

Service and maintenance:

- we are a **Core facility** in EM imaging and preparation equipment
- we offer **Sample preparation**
- we offer full service by **Engineers**
- we make **Assistance and advice**

Training:

- ❖ we train in the use of equipment
- ❖ we provide theoretical and practical training on sample preparation methods

Development:

- we develop new imaging techniques and provide state-of-the-art equipment and sample preparation to users.

Our expertise

⇒ **Sample preparation**

⇒ **About biology and materials**

Cryo-préparation :

- High pressure freezing, Freeze -substitution and low temperature resin inclusion
- Cryo-ultramicrotomy (Material and Biology)

Correlative microscopy:

- Dedicated support and preparation
- Localization of the ROI and ultrastructural observation

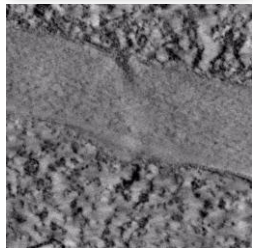
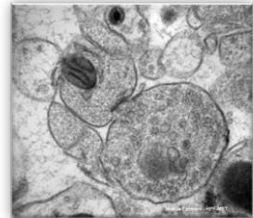
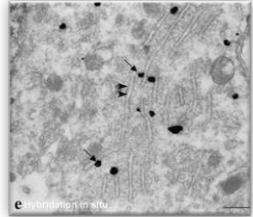
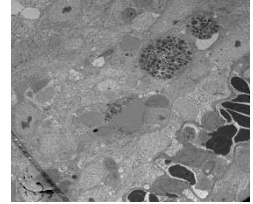
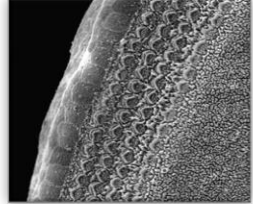
Immuno-staining :

- By pre-embedding or on section
- By Tokuyasu

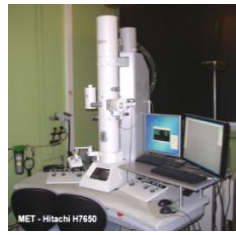
Devices of sample preparation

- 1 robot for conventionnal preparation (AMW)
- 1 High pressure freezer (HPM100)
- 3 robots for cryosubstitution
- 3 Ultra-microtomes
- 1 Cryo-ultramicrotome (UC7)





SEM



TEM



IHC



HPF



Tomography

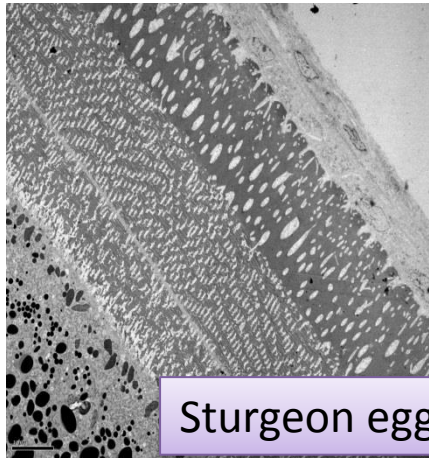
Topography

2D ultrastructural imagery

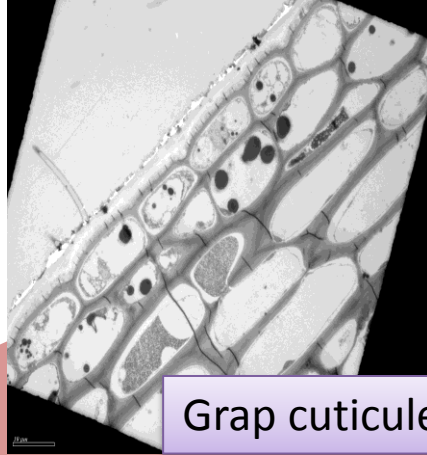
Immunolabelling 2D ultrastructural imagery

HPF 2D ultrastructural imagery

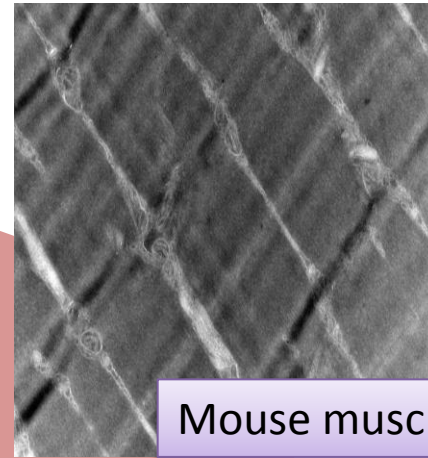
3D ultrastructural imagery



Sturgeon egg

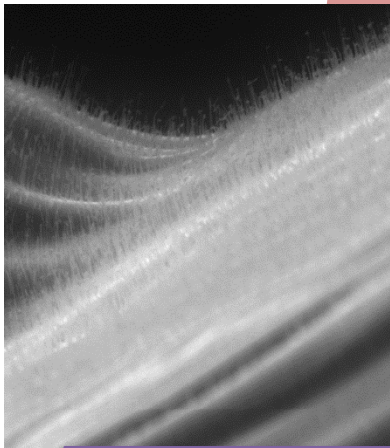


Grap cuticle

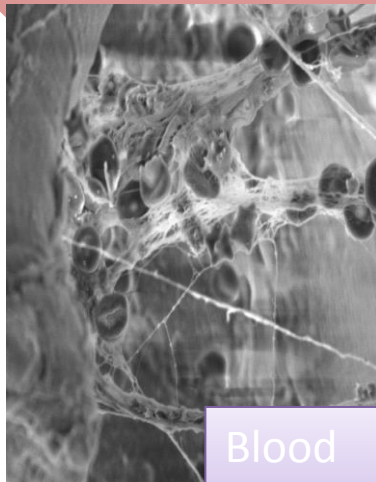


Mouse muscle

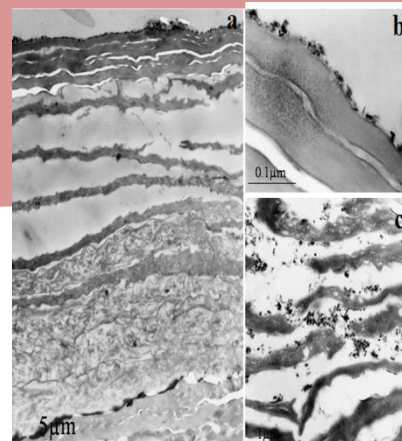
EM and Biology



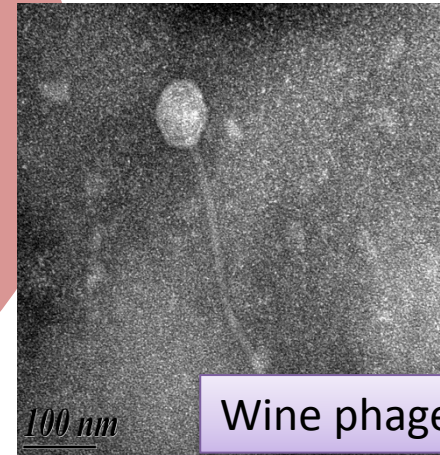
Arabidopsis stem



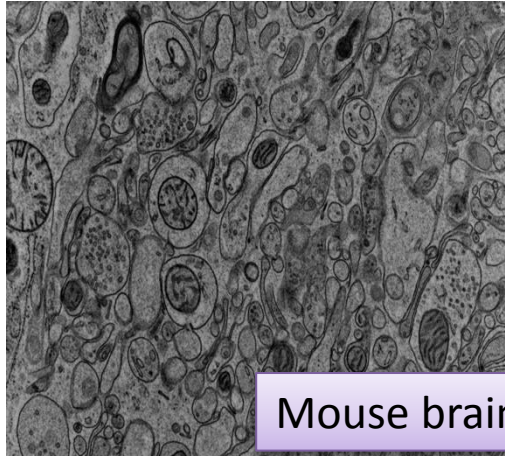
Blood cell



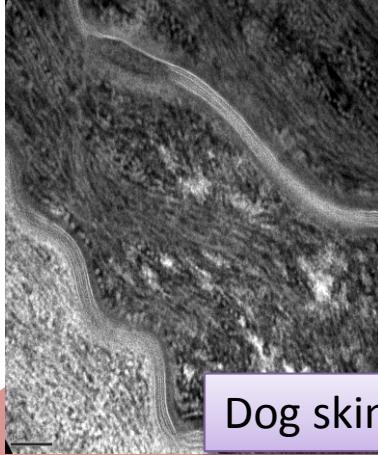
Effect of sunscreen on skin



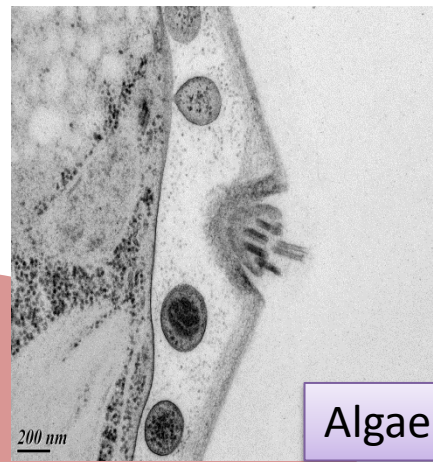
Wine phage



Mouse brain



Dog skin

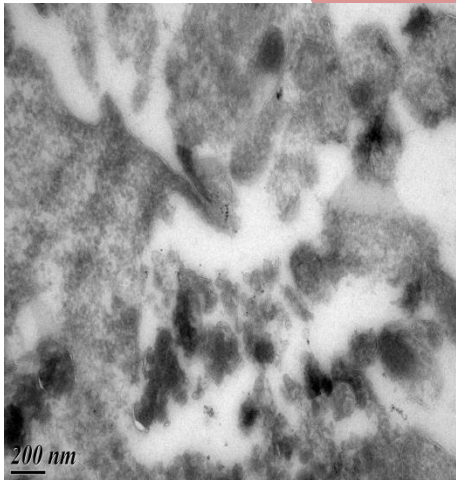
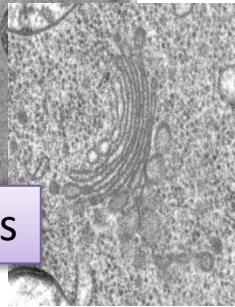


Algae

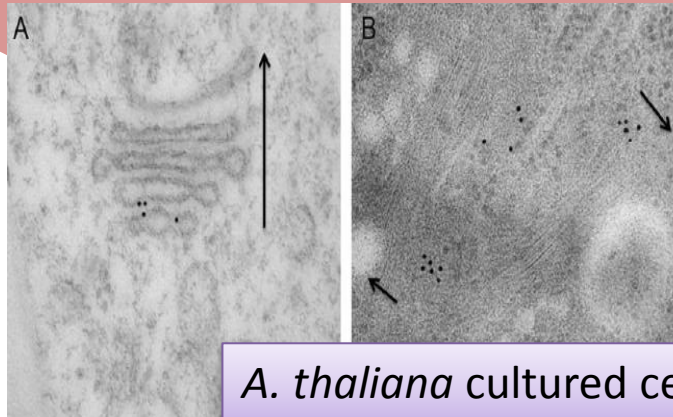
EM and Biology



Tabaco BY2 cells



Exosome in neuron cells



A. thaliana cultured cells

Physical demands of *TEM sample preparation*

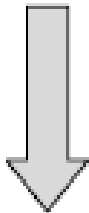
Biology

Aqueous/hydrated

Soft

Light elements
(C, O, H, N, S, P etc.)

"Large"

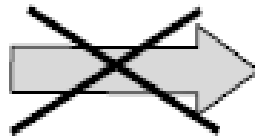


Biological samples need to be transferred into a solid state...

...which preserves the structures as a function of the living state...

...and not as a function of specimen preparation

Not suitable for EM

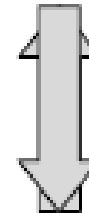


Electron microscope

High vacuum

Electron beam

Sensitive to vibration
(High magnifications)

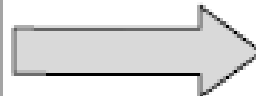


Resistant to high vacuum

Resistant in electron beam

Thin – permeable for electrons
(for TEM)

Contrast



TEM sample preparation

For biological samples → Fixation.
→ Embedding in resin.
→ Section.
(→ immunolabelling.)
→ Staining.

✓ 2 types of preparation : **Conventional or by cryomethods**

TISSUS

CELLULES

SUSPENSIONS

Cryoprotection : +/-

HPF

CRYO-SUBSTITUTION

FREEZE - DRYING

Dehydration at low T°

Inclusion at low T°

Inclusion at Room T°

Inclusion at low T°

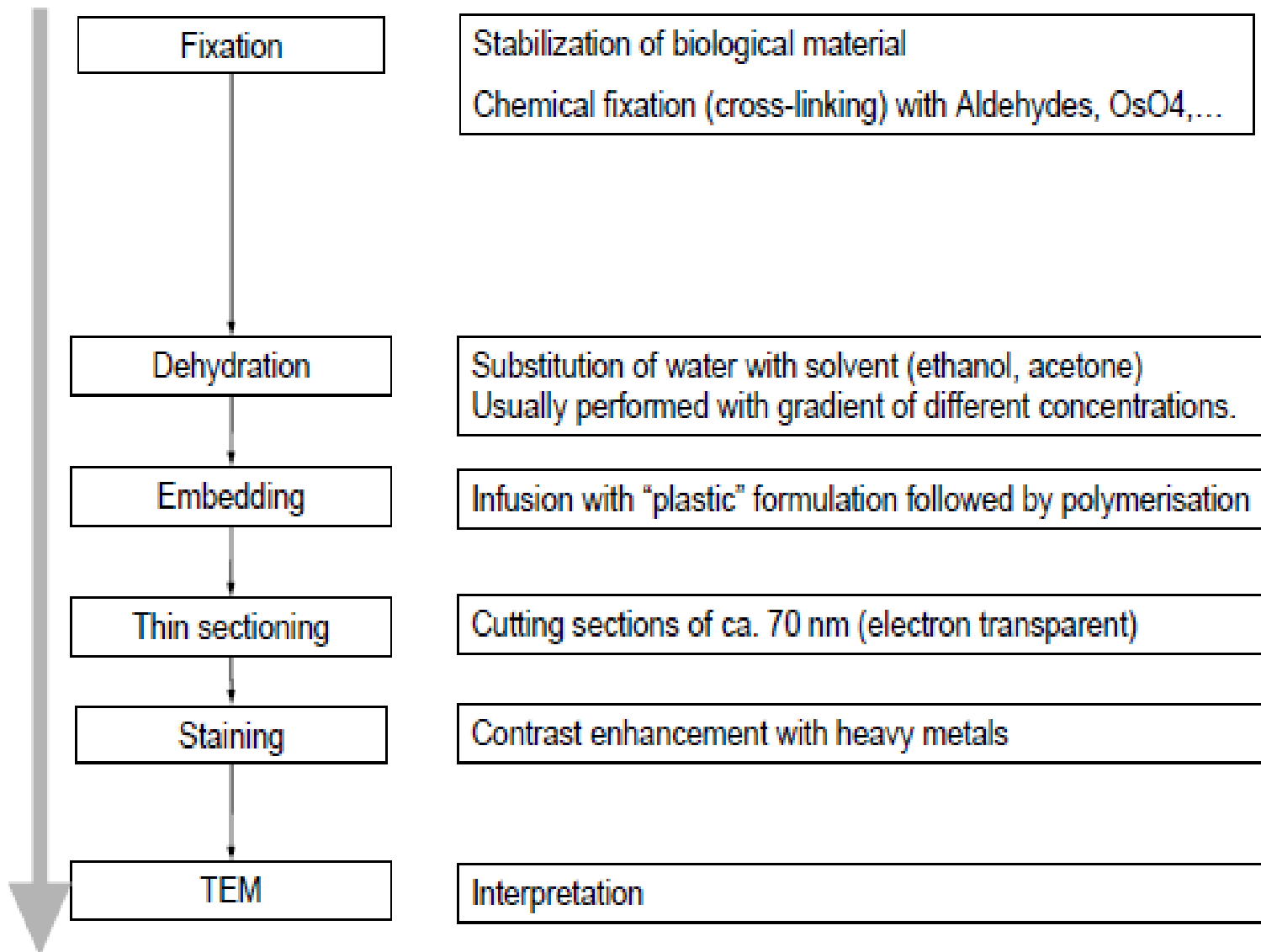
Inclusion at Room T°

Immunostaining

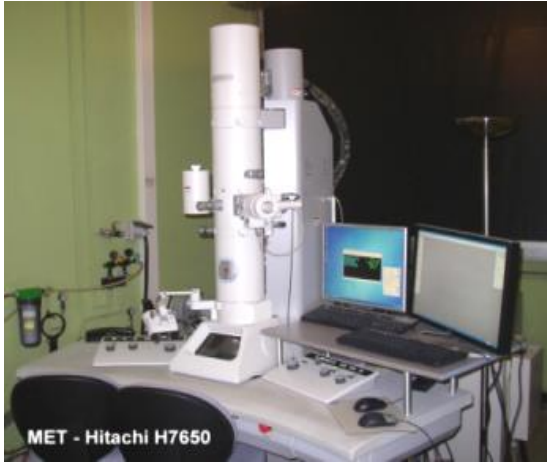
Ultrastructure
Elementary analysis EDX, EFTEM, SIMS

Ultrastructure
Elementary Analysis

TRANSMISSION ELECTRON MICROSCOPY



Transmission Electron Microscopy (TEM)



Hitachi - H7650 – 120kV

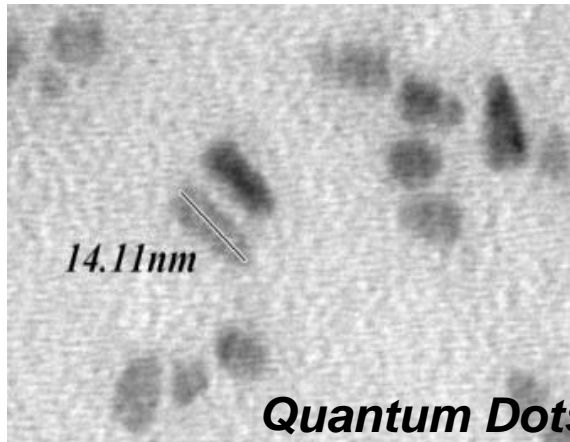
Interests :

- Resolution 1 nm
- It's **User friendly**
- Ultrastructural information and particle characterization

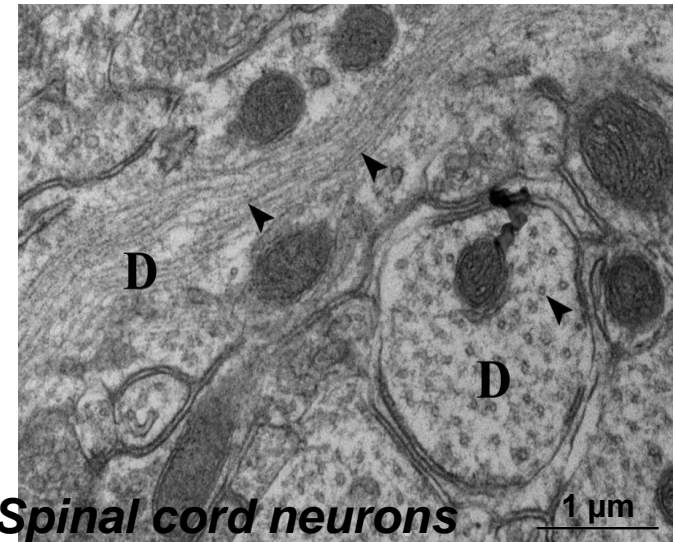
=> Conventional 2D ultra-structural imaging at high resolution

2 types of observation are possible:

High resolution for materials



High contrast for biology

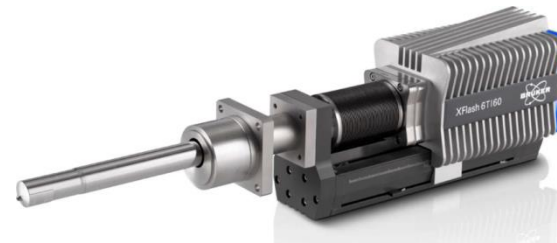


Microscopie Electronique en Transmission

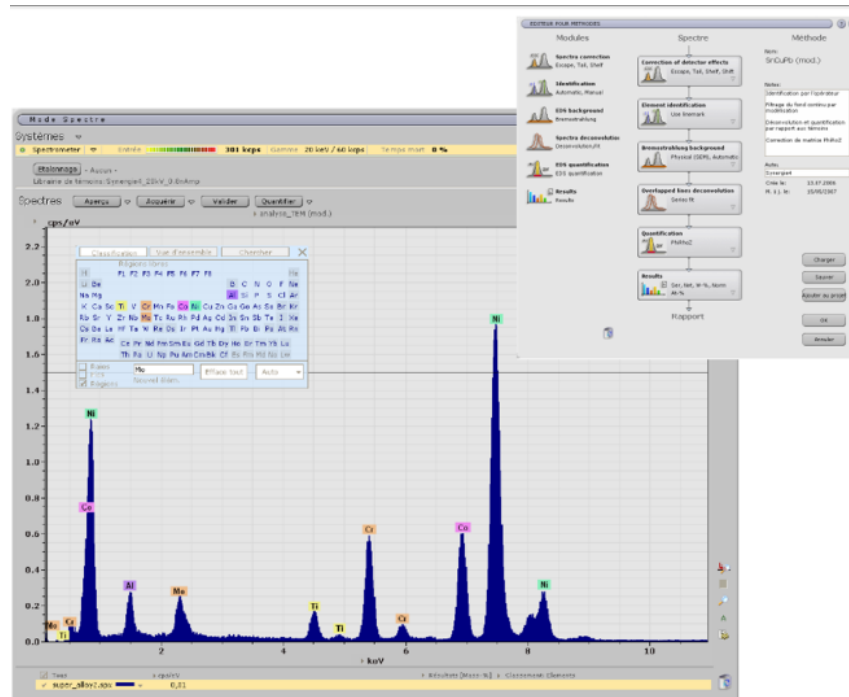
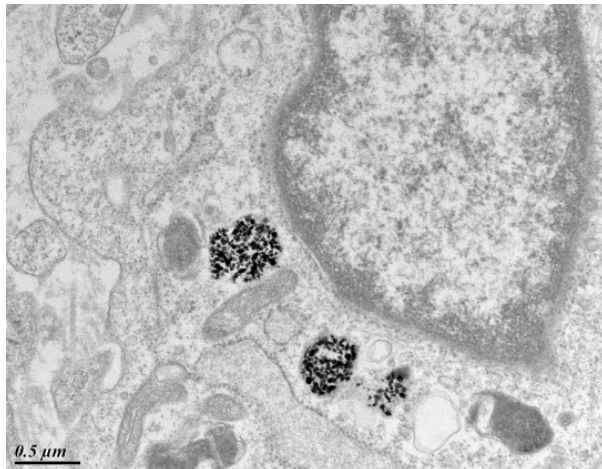


MET 120 kV Tecnai12

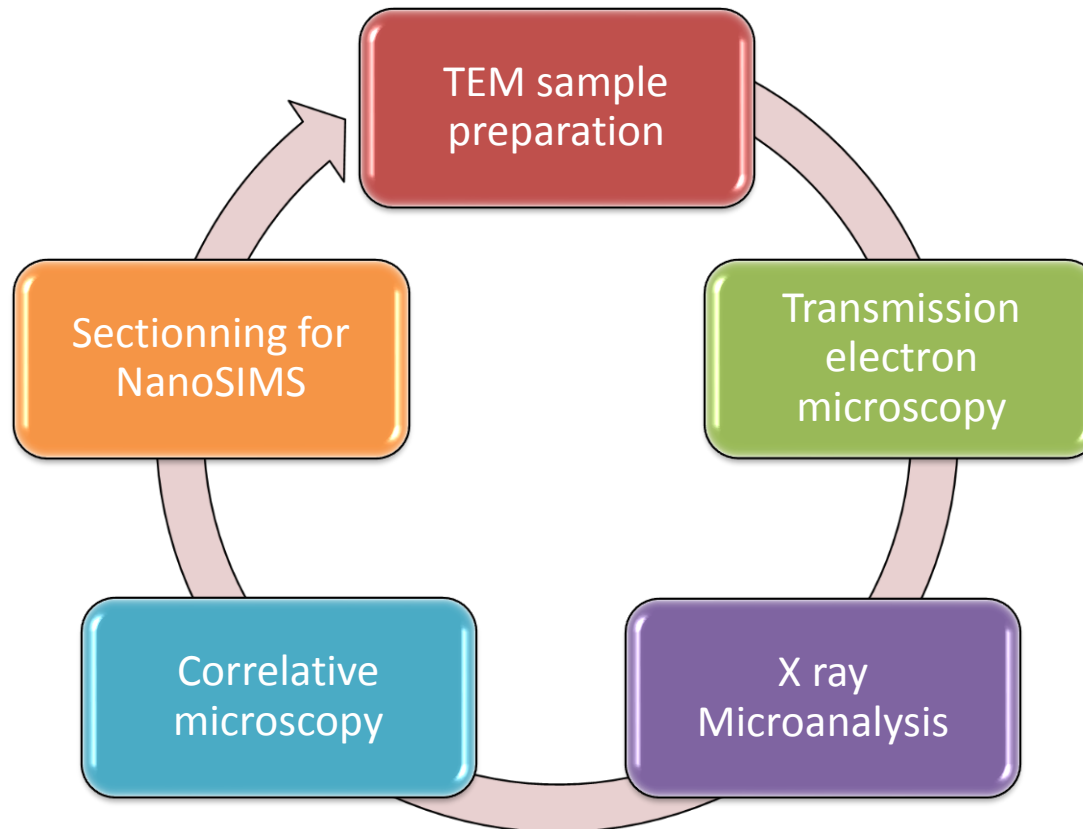
X-rays microanalysis



XFlash 6160T



BIC Field of work for AQUAMAPMET



Sample preparation possible for the project

Chemical Fixation

RT inclusion

Cryofixation

Freeze-drying

Cryosubstitution

Cryosection

First Results for TEM

Liver and gills of common carp (*Cyprinus carpio*)

RT method:

After 3-4 days, samples were prepared according to the following protocol :

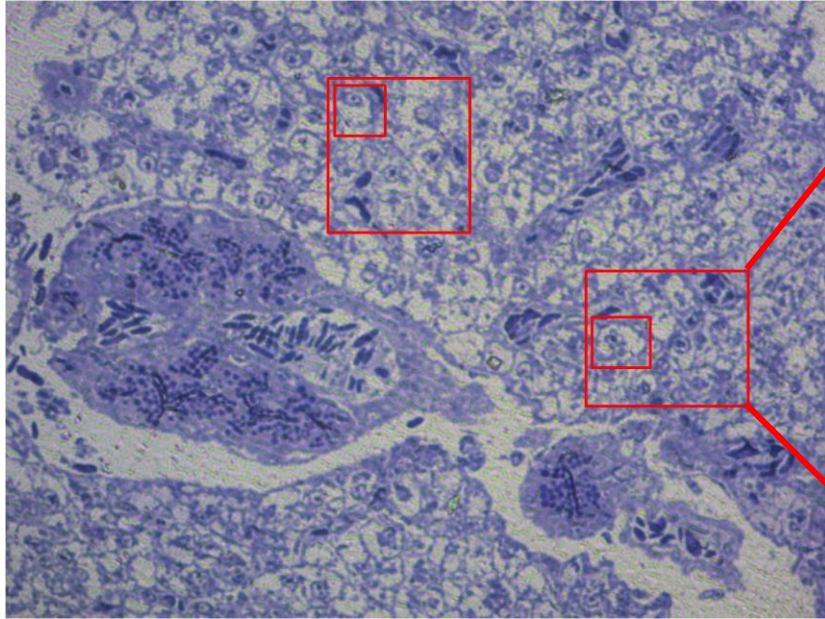
1. Infiltration by glutaraldehyde 4-5% in PBS (phosphate buffer) 0.1M (over night)
2. Washing during 10 min with PBS
3. Fixation with OsO₄ 1% for 1 hour
4. Washing
5. Dehydration steps:

1 ml Aqua dest + 0.5 ml Aceton (=30%)	10min
take out 300 µl, add 300 µl Aceton	10min
take out 600 µl, add 600 µl Aceton	10min
take out 1 ml, add 1ml Aceton	10min
take the whole volume out and refill with Aceton	2hrs
6. Resin (using EMbed-812 kit) infiltration steps:

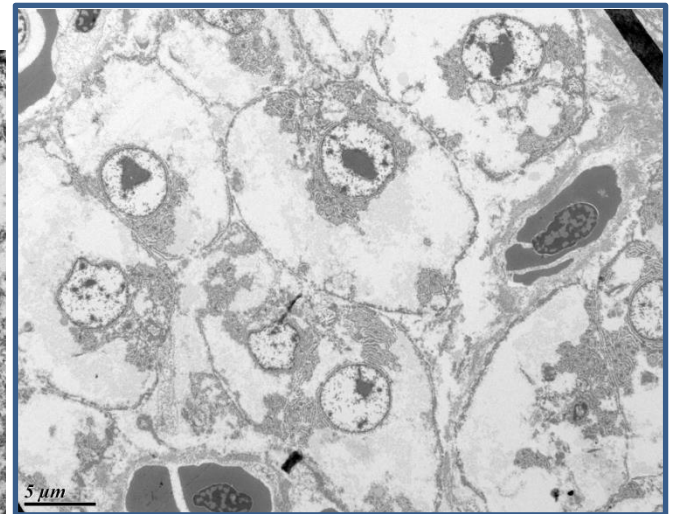
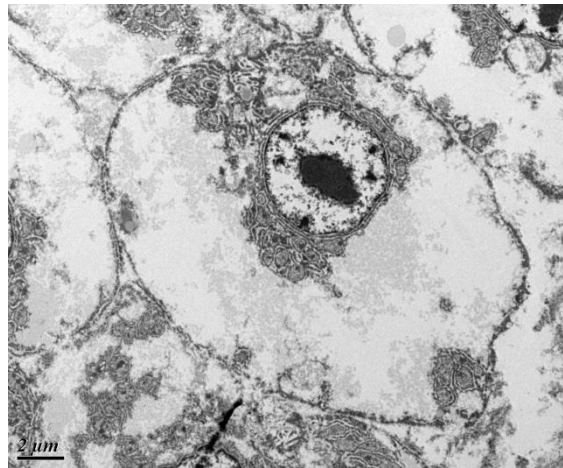
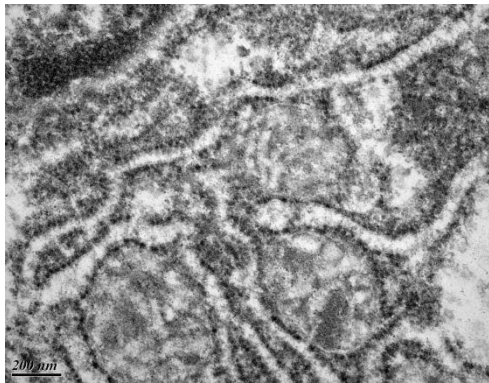
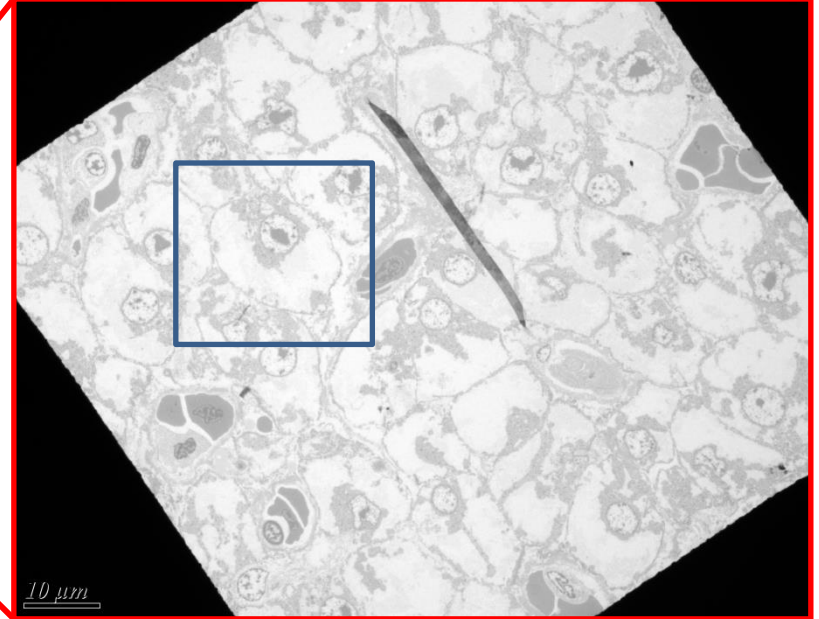
1:1 Aceton:Resin	30min
1:3 Aceton:Resin	1hrs
100% Resin	over night
7. Put the samples in moulds and bake out for 24hrs at 60°C

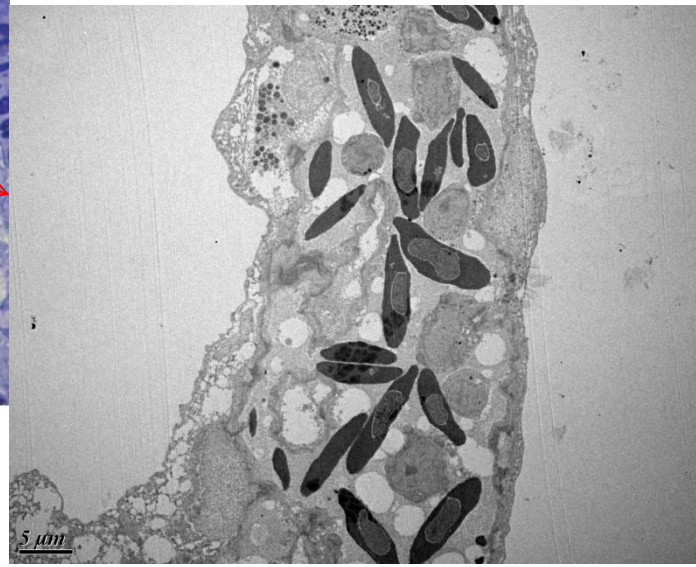
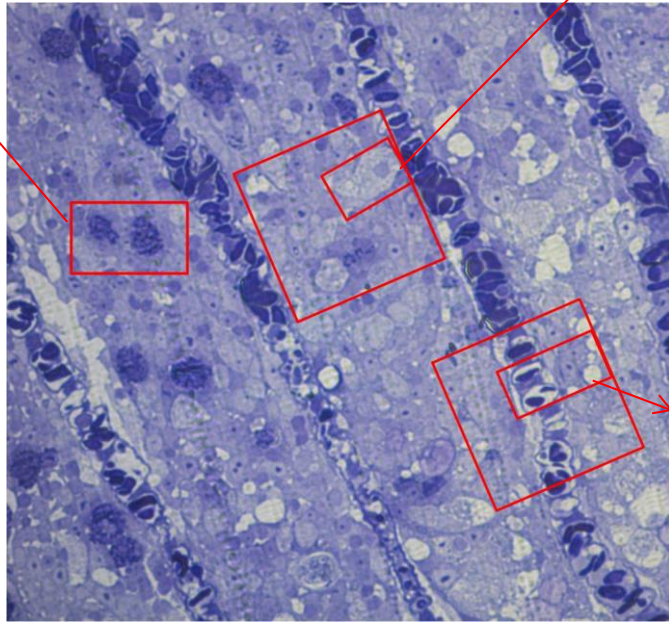
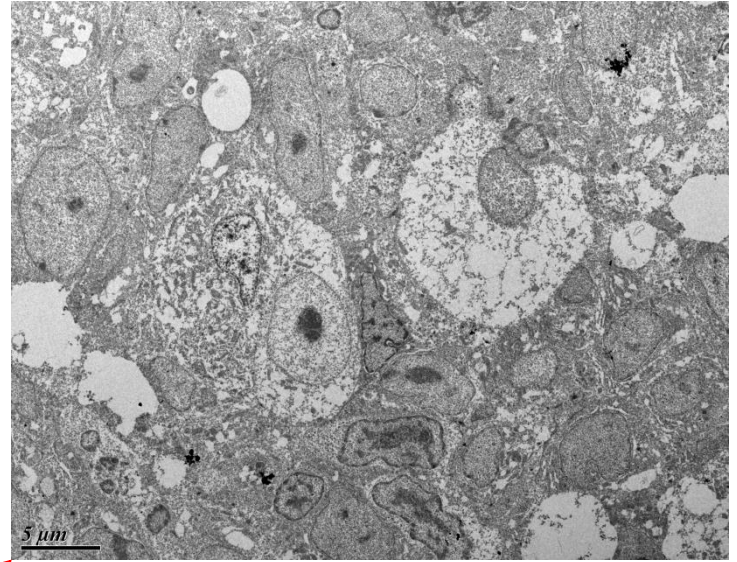
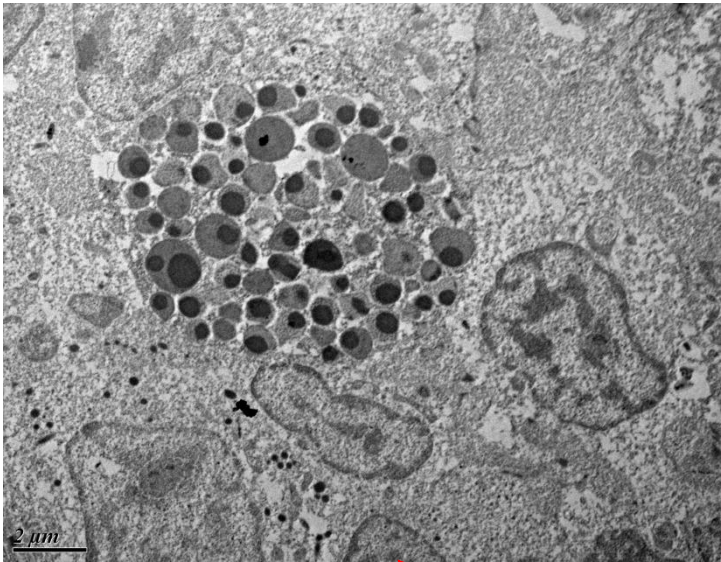
JETRA = LIVER

Semithin section



Ultrathin section





ŠKRGE = GILLS.



THANK YOU FOR YOUR ATTENTION



www.bic.u-bordeaux.fr

